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1 **Development of an SPME-GC-MS method for the specific quantification of dimethylamine and**
2 **trimethylamine: use of a new ratio for the freshness monitoring of cod filets.**

3 Running title: **Quantification of TMA & DMA by SPME-GC-MS for cod freshness monitoring**

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18

ABBREVIATIONS

19 CAR: Carboxen ®; DMA: Dimethylamine; DMA-N: Dimethylamine expressed as nitrogen equivalent;
20 DTR: DMA-N/TMA-N ratio; DVB: Divinylbenzene; FWHM: Full-width at half-maximum; GC: Gas
21 chromatography; LRI: Linear retention indices; MS: Mass spectrometry; PDMS: Polydimethylsiloxane;
22 PVB-N: Partial volatile basic nitrogen; QIM: Quality index method; SIM: Single ion monitoring;
23 SPME: Solid phase microextraction; TCA: Trichloroacetic-acid; TF: Tailing factor; TIC: Total ion
24 chromatogram; TMA: Trimethylamine; TMA-N: Trimethylamine expressed as nitrogen equivalent;
25 TVB-N: Total volatile basic nitrogen.

26

ABSTRACT

27 BACKGROUND: Fish is a highly perishable food, so it is important to be able to estimate its freshness
28 to ensure optimum quality for consumers. The present study describes the development of an SPME-
29 GC-MS technique capable of quantifying both trimethylamine (TMA) and dimethylamine (DMA),
30 components of what has been defined as partial volatile basic nitrogen (PVB-N). This method was used
31 together with other reference methods to monitor the storage of cod fillets (*Gadus morhua*) conserved
32 under melting ice.

33 RESULTS: Careful optimisation enabled definition of the best parameters for extracting and separating
34 targeted amines and an internal standard. The study of cod spoilage by sensory analysis and TVB-N
35 assay led to the conclusion that the shelf-life of cod fillet was between 6 and 7 days. Throughout the
36 study, TMA and DMA were specifically quantified by SPME-GC-MS; the first was found to be highly
37 correlated with the values returned by steam distillation assays. Neither TMA-N nor DMA-N were able
38 to successfully characterize the decrease in early freshness, unlike dimethylamine/trimethylamine ratio
39 (DTR), whose evolution is closely related to the results of sensory analysis until the stage where fillets
40 need to be rejected.

41 CONCLUSION: DTR was proposed as a reliable indicator for the early decrease of freshness until fish
42 rejection.

43 **Keywords:** SPME-GC-MS, TMA, DMA, TVB-N, PVB-N, QIM, fish freshness, storage assessment

44

INTRODUCTION

45 Fish is increasingly consumed around the world despite disparities recorded between continents. ¹ It is a
46 very convenient food, with good nutritional properties but is nevertheless highly perishable, with a short
47 shelf-life ranging from a few days to around twenty days in cold storage, depending on fish species and
48 packaging. ^{2,3} Freshness estimation is a key issue concerning this food matrix, as in fish there is a close
49 link between freshness and overall quality. ⁴ Consumers are more and more concerned about the quality
50 of what they eat, with specific attentions to price and freshness. ⁵ Accurate indicators and tools charac-
51 terizing the decrease in freshness are therefore required. To date, numerous techniques have been devel-
52 oped including sensory, ^{6,7} physical, ^{8,9} biochemical ¹⁰⁻¹³ and microbial ^{14,15} methods, but sensory analy-
53 sis remains the most widely used technique for freshness estimation by the industry. ¹⁶ Unfortunately,
54 these techniques have certain drawbacks. Concerning sensory analysis, the need for a constantly trained,
55 objective and available sensory panel can be problematic. ¹⁷ Physical methods usually require whole fish
56 or fillets with skin, which makes them less efficient with processed fish. The main problem encountered
57 with chemical analysis is their lack of reliability during the early stages of freshness decay. ³ Finally,
58 microbial methods require very long processing periods and these methods are highly variable regarding
59 numerous parameters. ¹⁸

60 After fish death and rigor mortis resolution, several processes occur, leading to freshness decrease and
61 fish spoilage. Among these processes, endogenous enzyme activities, nucleotide degradation, lipid oxi-
62 dation or development of specific spoilage organisms have been described as causative agents. ^{3, 19-21}
63 Marine fish are characterized by the presence of trimethylamine N-oxide (TMAO) in their flesh, though
64 its role remains uncertain: osmoregulation, buoyancy regulation, cryoprotection, etc. ²² In cod (*Gadus*
65 *morhua*), the mean TMAO content has been calculated between 60 and 120 mg nitrogen 100 g⁻¹, de-
66 pending on season or catching area. ³ TMAO can be catabolized in two principal ways: an exogenous
67 and an endogenous pathway. The first pathway results in its reduction to trimethylamine (TMA) by
68 TMAO reductase (EC 1.6.6.9), a molybdoenzyme presents on the periplasm of specific spoilage organ-
69 isms. ²³ The second pathway involves the action of endogenous TMAO demethylase (EC 4.1.2.32) and

70 leads to the production of dimethylamine (DMA) and formaldehyde in equimolar proportions; this en-
71 zyme activity is particularly important in gadoid fish species.²² Additional minor synthesis pathways
72 were described for TMA, which might originate from the degradation of choline, acetylcholine, carnitine
73 or betaines.²⁴ TMA and DMA are directly involved with quality and freshness issues as they result di-
74 rectly from the spoilage of fish through TMAO catabolism. On the one hand TMA is responsible of the
75 characteristic fishy-odour, with a rejection level in chilled fish comprised between 10 and 15 mg nitro-
76 gen 100 g⁻¹^{3, 25} while on the other hand DMA is produced soon after fish harvest, contributing to the
77 overall flavour of fish, with no specific odour described.²⁵ In the case of freezing, DMA is also an indi-
78 rect marker of flesh texture as it is produced together with formaldehyde, indeed methanal forms cross-
79 links with proteins resulting in a marked toughening of fish flesh.³

80 Historically; several methods have been developed to quantify TMA and DMA using capillary electro-
81 phoresis (CE), high-performance liquid chromatography (HPLC) or gas chromatography (GC).^{24, 26, 27}
82 Unfortunately, these methods usually necessitate an onerous sample preparation step with the use of
83 hazardous solutions, but some, such as CE and HPLC, are usually more informative than GC concerning
84 quantities of TMAO and some cations. Headspace analysis can markedly reduce sample preparation
85 time.^{26, 27} More recently, solid-phase micro-extraction (SPME) headspace methods have been developed
86 for quantification of volatile amines in fish.²⁸⁻³² Extraction steps together with separation of amines are
87 usually well described and transferable, although two studies did not manage to clearly differentiate
88 TMA and DMA signals.^{28, 29} Concerning fibres, most of the studies used fibres coated with
89 CAR/DVB/PDMS,³⁰⁻³² only one study used a non-available, laboratory-developed fibre.²⁹ Regrettably,
90 very few published studies described the optimization step for the sample preparation.^{29, 30} SPME meth-
91 ods were applied to numerous fish species involving fresh water or mangrove fish²⁹⁻³¹ and marine fish,
92^{28, 31, 32} but except for one study,²⁸ it was not applied to the most consumed marine species in Europe.
93 Among these studies, very few applied these methods to monitor the spoilage of fish.^{28, 30} Finally, when
94 used, comparative methods consist mostly of TVB-N analysis,^{28, 30, 32} **TMA-N** assay being overlooked,
95 and only one study used sensory tests.³⁰

96 This paper deals with the development of a new SPME-GC-MS method that specifically quantifies
97 TMA and DMA. This method, using a CAR/PDMS fibre, a new internal standard and fully commercial-
98 ly available consumables, was applied; together with QIM score evaluation, TVB-N and TMA-N assays,
99 to the concrete case of a spoilage study on cod (*Gadus morhua*). The SPME-GC-MS method was able to
100 quantify partial volatile basic nitrogen (PVB-N), the sum of TMA-N and DMA-N concentrations and to
101 define the DMA-N/ TMA-N ratio (DTR) used to characterize the early decrease in freshness.

102 EXPERIMENTAL

103 Chemicals

104 Concerning the chemicals used in this study, a C₅ - C₁₅ n-paraffin mix, and ammonium chloride, were
105 acquired from Sigma-Aldrich (St-Quentin-Fallavier, France). Other chemicals were used as pure com-
106 pounds solutions: trimethylamine hydrochloride (PubChem CID: 1146), dimethylamine hydrochloride
107 (PubChem CID: 674), or internal standards: 2-phenylethylamine (PubChem CID: 1001), sec-butylamine
108 (PubChem CID: 24874), N, N-diisopropylmethylamine (PubChem CID: 547370), 1,2-
109 dimethylpropylamine (PubChem CID: 61236), all of which were purchased from Sigma-Aldrich. Potas-
110 sium hydroxide pellets used to raise pH levels were also acquired from Sigma-Aldrich. Formaldehyde
111 solution was purchased from VWR (Fontenay-sous-Bois, France). Trichloroacetic-acid (TCA), 4% boric
112 acid with methyl red and bromocresol green indicator, 0.1 N Titrim® sulphuric acid and 10% sodium
113 hydroxide solutions were purchased from Dislab (Lens, France). Finally, sodium chloride (NaCl) was
114 acquired from Oxoid (Dardilly, France).

115 Solutions of pure compounds were prepared and diluted, if required, using a saturated NaCl solution at 4
116 °C. Eleven mL of solution was then poured into 20 mL SPME vials from Chromoptic (Courtaboeuf,
117 France), and a potassium hydroxide pellet (117 ± 14 mg, 85% purity) was added. Standard solutions
118 were prepared on the day of analysis as follows, 10 µL of pure compound solution was added to 10 mL
119 of bidistilled-grade water from Dislab and gently mixed, then 4.4 µL of standard solution was added in
120 SPME vials. Finally, vials were hermetically sealed.

121 Fish sample preparation

122 As a first step of method optimization, the effects of NaCl concentration and of the solution's pH were
123 evaluated. 50.0g of fish was blended with 100.0 mL of 30.0% (w/w) NaCl solution. After centrifugation
124 at 10,000g for 10 min at 4 °C, 11 mL of supernatant was poured into SPME vials hermetically sealed
125 after addition or not of a potassium hydroxide pellet.

126 During the storage study a single step of sample preparation was chosen, based on Regulation (EC) No.
127 2074/2005, ³³ to better compare SPME-GC-MS and steam distillation assays. Fillet was cut into 1 cm-
128 edge cubes, which were then mixed together to obtain a matrix representative of the overall freshness
129 state of fillet. 100.0 ± 0.2 g of flesh was then put into a Waring blender and approximately 200 g of a 7.5
130 % (w/w) TCA solution was added to ensure a weight ratio of 2:1. The whole was then blended and fil-
131 trated through a 5 µm cut-off filter paper, the filtrate being used for both steam distillation and SPME-
132 GC-MS assays.

133 **SPME-GC-MS procedure**

134 Vials were managed automatically thanks to a CTC Analytics CombiPAL (Zwingen, Switzerland). First-
135 ly, a sample was transferred to the extraction chamber, heated and shaken at 500 rpm for 10 min for the
136 headspace equilibration. A 23 Ga, 75 µm, Carboxen®/Polydimethylsiloxane (CAR/PDMS) SPME fibre
137 from Supelco (Saint-Germain-en-Laye, France) was then inserted into the headspace and volatile ad-
138 sorption was carried out. Times and temperatures of extraction were tested from 1 to 40 min at 35 °C
139 and 50 °C in order to determine the best temperature and extraction time. Once the volatiles had been
140 extracted, the SPME fibre was transferred to a Merlin Microseal injector from Supelco, where volatiles
141 were desorbed at a temperature of 210 °C for 1 min. Finally, after each injection SPME fibre was heated
142 to 300 °C, under a helium flow, for 20 min in the fibre conditioner device. Two types of liner were tested
143 during this study: regular SPME glass liner and deactivated SPME glass liner from Restek (Lisses,
144 France). Volatile compounds were separated and identified using a GC-MS QP2010 Plus from Shimad-
145 zu (Noisel, France). The GC device was successively equipped with two capillary columns; an SLB-5ms
146 capillary column 60 m x 0.25 mm x 0.25 µm from Sigma-Aldrich and a Stabilwax DB capillary column
147 30 m x 0.25 mm x 0.25 µm from Restek. Helium was used as the carrier gas for both, with respective

148 flows of 1.78 mL min⁻¹ and 1.20 mL min⁻¹. The GC conditions applied with SLB-5ms were the same as
149 described in a previous study.³⁴ The Stabilwax DB capillary column was used under the following con-
150 ditions: the oven temperature was initially set at 60 °C for 5 min, then increased to 210 °C at 15
151 °C.min⁻¹ and maintained for 5 min. To optimize the method, an experimental plan-like approach was
152 used on two levels of three parameters: split value (splitless or split of 5), injection temperature (150 °C
153 and 210 °C) and initial oven temperature (45 °C and 60 °C). To determine the impact of these parame-
154 ters on peaks, two shape indicators were used: tailing factor (TF) and full width at half maximum
155 (FWHM). MS conditions were set as follows: interface temperature equal to the final oven temperature
156 to avoid any recondensation, 70 eV of ionization voltage and for the early development steps, a mass
157 range from 33 to 200 m/z for total ion chromatogram (TIC) tracing. In the later phases, single ion moni-
158 toring (SIM) was preferred with specific ions of TMA, DMA and internal standard (42 m/z, 44 m/z, 45
159 m/z, 55 m/z, 58 m/z, 59 m/z and 72 m/z). Volatile compounds were identified using the NIST08 spectral
160 database and comparison of linear retention indices (LRI) with those of the corresponding pure chemi-
161 cals.

162 **Shelf-life assessment**

163 Eleven gutted, whole cod (*Gadus morhua*) were purchased at a local fish retailer in Boulogne-sur-Mer
164 (France). Fish were caught in the north-east Atlantic, during spring, and were packed into two polysty-
165 rene boxes, covered by crushed ice and insulated by a plastic film. Fish were acquired from the auction
166 of the day to ensure the best freshness grade. Based on QIM attributes, the initial quality of raw fish was
167 very good; skin was bright and flesh was greyish, flat eyes clearly revealing a black pupil thanks to a
168 transparent cornea, with only the gills displaying a slight discoloured aspect. Fish preparation was car-
169 ried out at the innovation platform *PFI-Nouvelles Vagues* (Boulogne-sur-Mer, France). In a room with
170 ambient temperature maintained at 10 °C, each fish was filleted with skin. Three fillets were placed in
171 new polystyrene boxes, taking care to place the two fillets of any given fish in separate boxes. After
172 insulation with plastic film, boxes were filled with fresh crushed ice and placed in a cold room at +2.0
173 °C. The day of fillet preparation was the day of the first analysis and was considered as D01 of the study.

174 On days D01, D04, D07, D09, D11, D14 and D16, a single box was withdrawn and the three fillets were
175 analysed.

176 On the day of analysis each of the fillets was processed in the same way, as follows. Firstly, the fillet
177 was subjected to sensory analysis, carried out by three judges, using QIM score evaluation as described
178 by Cardenas Bonilla *et al.* ³⁵After fish preparation as previously described, 1 mL of filtrate was con-
179 served for SPME-GC-MS analysis and the remainder was used for total volatile basic nitrogen (TVB-N)
180 and TMA-N assays, in duplicate, by steam distillation as previously described. ³⁴ Briefly, before the tube
181 to be placed in a Gerhardt Vapodest distillation unit (Les Essarts-le-Roi, France), 25 mL of filtrate were
182 loaded in tube. In case of TMA-N assays, with 20 mL of formaldehyde were added. Then, 6 mL of 10%
183 NaOH was poured and the distillation was performed. The distillate was collected into a beaker contain-
184 ing 10 mL of 4% boric acid, before titration with a 0.1 N sulfuric acid solution. According to Regulation
185 (EC) No. 2074/2005, ³³ a solution of NH₄Cl at 1.91g.L⁻¹ was used as a control for steam distillation as-
186 says. Results were considered as valid for recovery rates comprised between 98% and 101%. For
187 SPME-GC-MS, the filtrate was diluted with a saturated NaCl solution from 1/100 to 1/4000 depending
188 on the day of analysis concerned. On each day, filtrate was analysed in triplicate as follows: 11 mL of
189 diluted filtrate was poured into SPME vials, 4.4 µL of 1,2-dimethylpropylamine standard solution was
190 added together with one potassium hydroxide pellet and vials were hermetically sealed.

191 **Descriptive statistics, inferential statistics and modelling**

192 All of the statistical work was carried out using R software 3.1.1 (<http://www.r-project.org>). Except
193 where otherwise indicated, error bars shown on bar charts and line graphs illustrate twice the standard-
194 error value (standard deviation of the sampling distribution), thus showing the 95% confidence limits of
195 the measurement. When least significant difference (LSD) tests are performed, two results tagged with
196 different letters are significantly different with an error risk of 5%. ANOVA were carried out after care-
197 fully establishing normality of the distribution using Shapiro-Wilks test and distribution homoscedastici-
198 ty using Bartlett's test, all under the independent sample hypothesis. Interdependence of the values

199 yielded by this research was evaluated by calculating Pearson's correlation or Spearman's correlation
200 coefficients depending on the linear relationships between them.

201 TMA production being a reflection of the specific spoilage organisms' metabolism activity, modelling of
202 **TMA-N** by a microbial growth model was tested. Using a modified Gompertz model,³⁶ a non-linear
203 regression was performed with R software (**Supporting information – Algorithm S1**). The three pa-
204 rameters A , μ_m and λ were entered as input variables in accordance with the observed data and an itera-
205 tive process was followed to refine the parameters in order to fit the model to observed data. The **DMA-**
206 **N/TMA-N** ratio (DTR) was modelled against QIM using an exponential equation with another non-
207 linear regression (**Supporting information – Algorithm S2**), **the data structure suggesting this type of**
208 **approach**. In both cases a special care was taken to enter the most probable parameter values to reduce
209 the risk of reaching local minima during the iterative process.

210 **RESULTS AND DISCUSSION**

211 **Sample preparation optimization**

212 In a first step of method optimization, special attention was applied to the chemical conditions of the
213 sample. **In absence of potassium hydroxide** (KOH), pH values of solutions were lower than 8.0 and no
214 detectable peak was recorded on chromatogram. **It was already known that TMA and DMA are only**
215 **volatiles in their conjugate base form**²⁸ and have respective pKa of 9.76 and 10.64.³⁷ **Addition** of KOH
216 in the solution **increased** the pH above **12.0 and clear peaks appeared** on chromatograms. NaCl tended to
217 increase both TMA and DMA recovery with a clear improvement of the peak area for TMA and a slight-
218 er increase concerning DMA which is in accordance with previous observations.²⁹ It was thus decided
219 to retain the use **of a saturated solution of NaCl, with** the addition of a KOH pellet into the vial for the
220 preparation of samples.

221 **Optimization of headspace extraction**

222 The second group of parameters optimized concerned the conditions for headspace extraction, mostly
223 concerning time and temperature of extraction as, these have already been described as key parameters,
224 although few studies include a documented optimization step.³⁰ Two temperatures (35 °C and 50 °C)

225 and seven extraction times (1, 2, 5, 10, 20, 30 and 40 min) were tested, as illustrated in **Figure 1**. Com-
226 pared with similar extraction times at 50 °C, bar graphs clearly show that an extraction temperature of
227 35 °C produced peaks higher intensities from time of 5 min and longer (p-value < 0.001). Concerning
228 time of extraction, longer times led to higher peak intensities. Least significant difference tests tended to
229 conclude that a significant increase of peak area was observed between 20 min and 30 min of extraction,
230 whereas area increase between 30 min and 40 min was not significant. Regarding these results, extrac-
231 tions at 35 °C for 30 min were chosen as the best compromise for extracting volatiles. Extraction by
232 SPME is the result of two concomitant equilibria: first between matrix and headspace and then between
233 fibre and headspace. Results giving smaller areas observed with an extraction temperature of 50 °C
234 could be explained by the fact that equilibrium between fibre and headspace displaced towards the head-
235 space, as previously reported.³⁸ Following this observation, increasing the time of extraction at a fixed
236 low extraction temperature resulted in the adsorption of more volatile compounds on SPME fibre. The
237 temperature of 35 °C was confirmed by two other studies,^{30,31} but in contrast the time of extraction was
238 slightly higher than the range from 5 to 22 min described elsewhere.²⁸⁻³² This difference could be ex-
239 plained by the fact that a CAR/PDMS fibre was used instead of Carbox-
240 en®/Polydimethylsiloxane/Divinylbenzene (CAR/PDMS/DVB) or Polydimethylsilox-
241 ane/Divinylbenzene (PDMS/DVB) fibres, and in addition no saturation was noted after 10 min, as was
242 observed by Chan *et al.*³⁰

243 **Injection and separation optimizations**

244 A comparison of the above-mentioned columns concluded that Stabilwax-DB columns offer the best
245 peak shapes (data not shown). Improvement of peak shapes could be explained by the phase coated on
246 the Stabilwax-DB columns composed of Carbowax® polyethylene glycol, with higher polarity than 5%
247 diphenyl/95% methyl-siloxane coated on the SLB-5ms columns. Apart from the phase coated on the
248 columns, peak resolutions and shapes are also influenced by numerous parameters, including injector
249 temperature, oven temperature or split value. **Table 1** reports ANOVA results assessing the impact of the
250 three parameters tested on TMA and DMA peak shapes. Except for the initial temperature of the col-

251 umn, which has no significant impact on the tailing factor of TMA, the three parameters had a signifi-
252 cant impact on both tailing factor and full-width at half-maximum of TMA and DMA. The shape of
253 DMA peaks is significantly influenced by all the interactions between the parameters, whereas the shape
254 of TMA peaks is more likely significantly influenced by the injection parameters, *i.e.* injector tempera-
255 ture and split value. For each assessed parameter, the highest levels led to the best values concerning
256 shape indicators. An injection at 210 °C with a split value of 5 and an initial column temperature of 60
257 °C were thus chosen to perform the next analysis.

258 Deactivated SPME glass liner (data not shown) led to greater areas for both TMA and DMA; it was thus
259 selected for analysis of volatile amines.

260 **Internal standard screening and quantification range**

261 To overcome signal variation, an internal standard was added for sample preparation. Four compounds,
262 2-phenylethylamine, sec-butylamine, N,N-diisopropylmethylamine and 1,2-dimethylpropylamine were
263 tested, as they share structures or chemical properties close to the amines studied. As special attention
264 was paid to safety issues when selecting internal standards, sec-butylamine was rapidly discarded be-
265 cause of its toxic, polluting and highly corrosive properties. Among the remaining compounds 1,2-
266 dimethylpropylamine was the molecule that led to best linearity of the signal for both TMA and DMA
267 (data not shown). Using 1,2-dimethylpropylamine as internal standard enabled a quantification range
268 from 100 µg kg⁻¹ to 900 µg kg⁻¹ for TMA and from 150 µg kg⁻¹ to 1300 µg kg⁻¹ for DMA with good
269 linearity characterized by respective determination coefficients of $R^2_{\text{TMA}} = 0.98$ and $R^2_{\text{DMA}} = 0.99$ (**Sup-**
270 **porting information – Figure S1**). No further investigations were brought to LOD estimation, consid-
271 ering that lower TMA and DMA values, observed in fish preparation, will rarely be lower than 1 mg kg⁻¹.
272 ³⁹

273 **Application to fish freshness assessment monitoring**

274 Sensory analysis results are show in **Figure S2 (Supporting information)**. A marked evolution is rec-
275 orded from D01 to D11 with an average QIM score increase from 0 to 13; no significant increase being
276 observed afterwards, this might reflect a final spoilage state of cod fillets. Throughout the study the av-

277 erage QIM score values were linear in comparison with conservation days with an acceptable determina-
278 tion coefficient (adjusted $R^2 = 0.97$); these results are consistent with those of previous published results.
279 ³⁵ Based on the Cardenas Bonilla *et al.* method, the rejection limit, corresponding to a QIM score of 8,
280 was observed in the neighbourhood of Day 7. Priority has been given to mean QIM score values in the
281 development below.

282 Results concerning steam distillation assays are presented on **Figure S3 (Supporting information)**.
283 Unfortunately, a technical problem on D09 (Day 9) prevented the acquisition of workable data concern-
284 ing biochemical analysis; this day does not appear in the charts. Based on TVB-N measurement and EC
285 Regulation No. 2074/2005, ³³ fillets were considered as spoiled between D04 and D07, probably after
286 D06. These results are consistent with the sensory analysis and previous observation with the same
287 methods of fillet processing and storage. ³⁵ Comparison of TMA-N concentrations measured by steam
288 distillation assay and SPME-GC-MS (**Figure 2 a**) led to highly correlated values ($r = 0.98$; p -value $<$
289 0.001). Moreover, there is a linear relationship between values of the TMA-N obtained by the two meth-
290 ods (adjusted $R^2 = 0.99$) (**Supporting information – Figure S4**), and no significant difference was ob-
291 served between the two methods throughout the study (p -value = 0.42). In addition, TMA-N concentra-
292 tion is highly correlated with both QIM values ($r = 0.97$; p -value < 0.001) and days of storage ($r = 0.93$;
293 p -value < 0.001). A modified Gompertz model enabled TMA-N production to be modelled accurately
294 throughout the study and additional information was inferred from this model. Thanks to modelling, the
295 lag phase of TMA-N production was estimated to be 4.8 days. During the exponential phase, the daily
296 average increase of TMA-N concentration was $8.5 \text{ mg nitrogen } 100 \text{ g}^{-1}$. Finally, theoretical initial and
297 final TMA-N concentrations in cod fillets were respectively estimated to 4.4 and $66.3 \text{ mg nitrogen } 100$
298 g^{-1} . Compared with steam distillation, PVB-N monitoring by SPME-GC-MS was capable of the specific
299 detection of DMA-N, so it was possible to monitor concentration throughout the storage period (**Figure**
300 **2 b**). Compared with TMA-N, DMA-N levels remained lower than $20 \text{ mg nitrogen } 100 \text{ g}^{-1}$ in the flesh
301 of cod fillet. Additionally, DMA-N correlation values are lower than those of TMA-N, but correlation
302 factors remain acceptable for QIM values ($r = 0.93$; p -value < 0.001) and days of storage ($r = 0.88$; p -
303 value < 0.001). Recorded concentrations of DMA-N follow the same linear trend mentioned in a previ-

304 ous study on gutted whole cods,⁴⁰ but values are almost ten times higher; it could be hypothesized that
305 fish filleting might have discharged TMAO demethylase and induced an increase in its activity in com-
306 parison with the one in whole fish.

307 **Proposition of the DTR ratio**

308 Despite acceptable correlations with QIM scores and number of days of storage throughout the storage
309 period, neither TMA-N nor DMA-N evolved markedly in the early stages of fish freshness decrease. It
310 was decided to further investigate the quantities of PVB-N compounds using ratio as described in an
311 earlier study.¹⁰ Many ratios were tested including: TMA-N/PVB-N, DMA-N/PVB-N, PVB-N/TMA-N
312 and PVB-N/DMA-N, but none of them provided sufficiently accurate information. By contrast, the
313 DMA-N/TMA-N ratio (DTR) led to interesting results; it illustrates the relative proportion of autolytic
314 activity in the degradation of TMAO relative to the spoilage activity. Plotting DTR against QIM score
315 values (Figure 3) showed a marked decrease of DTR values for QIM scores comprised between 0 and 7,
316 corresponding to fish suitable for human consumption. DTR values were found to be homogeneous with
317 an exponential function of QIM values (Figure 3). Globally, DTR values decreased from 0.9 to 0.2
318 throughout storage, illustrating the increasing preponderance of the activity of specific spoilage organ-
319 isms in comparison with autolysis. The limit of 0.2 corresponded to QIM values higher than the maxi-
320 mum storage time estimated for the fillets of this study. It should be mentioned that three outlier values
321 were recorded for a QIM value in the neighbourhood of 0 with DTR value slightly lower than 0.4; these
322 values correspond to the analysis in triplicate of a single fillet performed on D01. These atypical values
323 are the result of TMA-N values markedly superior to those for the other fish and might be the fingerprint
324 of an early development of spoilage flora.

325 No recovery rate was calculated in this study, as it was in another studies,²⁶ to assess the matrix effect
326 on volatile amine extraction and analysis method. Nonetheless, the SPME-GC-MS method was applied
327 together with steam distillation assays, which is well characterized and used as a reference in Europe,³³
328 and TMA-N results from both methods were not significantly different.

329 The authors note that further storage studies, species, catch seasons, and fish processing techniques need
330 to be analysed with this method in order to confirm the suitability of DTR in assessing fish freshness.
331 The developed method made it possible to completely dispense with the use of formaldehyde, sulphuric
332 acid and boric acid. For steam distillation assays, all these compounds require special care concerning
333 the safety of technicians and waste management, because of their polluting, highly corrosive and toxic
334 properties. In the future the use of TCA, which is polluting and corrosive, could be avoided; a procedure
335 for fillet preparation with TCA solution replaced by a saturated NaCl solution should be considered.
336 This method of sample preparation was not used in this study to enable a better comparison of SPME-
337 GC-MS with steam distillation assays. Nonetheless, preliminary results (data not shown) demonstrated
338 that the use of a saturated NaCl solution for fillet preparation did not affect TMA and DMA results rec-
339 orded on chromatograms. The main drawback of the developed method is analysis time compared with
340 steam distillation assays; nonetheless this is quickly offset by the specificity of the detection of volatile
341 amines and the possibilities it opens up. Moreover, based on the results shown in **Figure 1**, it might be
342 possible to reduce extraction times considerably by reducing dilutions during samples preparation; nev-
343 ertheless, linearity of the signal would have to be checked beforehand.

344 CONCLUSION

345 A new SPME-GC-MS procedure for quantification of both TMA and DMA was developed. Special care
346 was paid to optimize different parameters such as solution preparation, extraction, volatile compounds
347 separation and signal normalization, with the use of a new internal standard. The developed method
348 proved capable of analysing PVB-N, and quantifying of both TMA-N and DMA-N, in a sample within
349 fifty minutes. PVB-N measurement by the SPME-GC-MS procedure was applied, together with refer-
350 ence steam distillation assays and sensory analysis, to a shelf-life monitoring study on cod fillets con-
351 served under melting ice. First results are encouraging, indeed TMA-N results using SPME-GC-MS and
352 steam distillation assay were very highly correlated. Modelling of TMA-N production throughout the
353 storage period was successfully carried out using a modified Gompertz model. Finally, the study of DTR
354 evolution in regard to QIM score values made it possible to visualize the early freshness decrease of cod

355 fillets. The authors thus propose DTR as a new tool for monitoring the early freshness evolution in fish.
356 The Authors also note that further investigations need to be carried out to confirm the reliability of the
357 method; nevertheless, the use of hazardous chemicals can already be significantly reduced by adopting
358 this SPME-GC-MS method.

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365 REFERENCES

- 366 1. Food and Agriculture Organization, The state of world fisheries and aquaculture. (2012)
- 367 2. Ashie I, Smith J, Simpson B and Haard NF, Spoilage and shelf-life extension of fresh fish and
368 shellfish. *Critical Reviews in Food Science & Nutrition* **36**:87-121 (1996)
- 369 3. Huss HH, Quality and quality changes in fresh fish. *FAO fisheries technical paper* (1995)
- 370 4. Oehlenschläger J and Sørensen NK, Criteria of seafood freshness and quality aspects, in
371 *Methods to Determine the Freshness of Fish in Research and Industries, Proceedings of the Final*
372 *Meeting of the Concerted Action "Evaluation of Fish Freshness"*, AIR3CT94-2283, ed. by Ólafsdóttir
373 G, Luten J, Dalgaard P, Carreche M, Verrez-Bagnis V, Martinsdóttir E and Heia K, Nantes (France), pp
374 30-35 (1997).
- 375 5. Brunsø K, Consumer research on fish in Europe, in *Quality of Fish from Catch to Consumer:*
376 *Labelling, Monitoring and Traceability*, ed. by Luten J, Oehlenschläger J and Ólafsdóttir G. Wageningen
377 Academic Publisher, Wageningen, pp 335-344 (2003).
- 378 6. Warm K, Boknass N and Nielsen J, Development of quality index methods for evaluation of
379 frozen cod (*Gadus morhua*) and cod fillets. *Journal of Aquatic Food Product Technology* **7**:45-59 (1998)
- 380 7. The Commission of the European Communities, Council Regulation (EC) No 2406/96 laying
381 down common marketing standards for certain fishery products. *Official Journal of the European Union*
382 **L334** (1996)
- 383 8. Oehlenschläger J, Luten J and Ólafsdóttir G, Measurement of freshness quality of fish based on
384 electrical properties, in *Quality of Fish from Catch to Consumer: Labelling, Monitoring and*
385 *Traceability*, ed. by Luten J, Oehlenschläger J and Ólafsdóttir G. Wageningen Academic Publisher,
386 Wageningen, pp 237-249 (2003).
- 387 9. Autheman C and Widmann L, Method and device for checking the freshness of a fish Patent No
388 EP2189789 A1 (2010).
- 389 10. Malle P and Poumeyrol M, A new chemical criterion for the quality control of fish:
390 trimethylamine/total volatile basic nitrogen (%). *Journal of food protection* **52**:419-423 (1989)

- 391 11. Valle M, Malle P and Bouquelet S, Evaluation of fish decomposition by liquid chromatographic
392 assay of ATP degradation products. *JAOAC Int* **81**:571-575 (1998)
- 393 12. Aubourg SP, Recent advances in assessment of marine lipid oxidation by using fluorescence.
394 *Journal of the American Oil Chemists' Society* **76**:409-419 (1999)
- 395 13. Duflos G, Leduc F, N'Guessan A, Krzewinski F, Kol O and Malle P, Freshness characterisation
396 of whiting (*Merlangius merlangus*) using an SPME/GC/MS method and a statistical multivariate
397 approach. *Journal of the Science of Food and Agriculture* **90**:2568-2575 (2010)
- 398 14. Dalgaard P, Mejlholm O and Huss HH, Conductance method for quantitative determination of
399 *Photobacterium phosphoreum* in fish products. *Journal of Applied Bacteriology* **81**:57-64 (1996)
- 400 15. Dehaut A, Krzewinski F, Grard T, Chollet M, Jacques P, Brisabois A and Duflos G, Monitoring
401 the freshness of fish: development of a qPCR method applied to MAP chilled whiting. *Journal of the*
402 *Science of Food and Agriculture* (2015).10.1002/jsfa.7322
- 403 16. Consortium "Fish Freshness", Future recommendations, in *Methods to Determine the Freshness*
404 *of Fish in Research and Industries, Proceedings of the Final Meeting of the Concerted Action*
405 *"Evaluation of Fish Freshness"*, AIR3CT94-2283, ed. by Ólafsdóttir G, Luten J, Dalgaard P, Carreche
406 M, Verrez-Bagnis V, Martinsdóttir E and Heia K, Nantes (France), p 11 (1997).
- 407 17. Hyldig G, Martinsdóttir E, Sveinsdóttir K, Schelvis R and Bremner A, Quality index Methods,
408 in *Handbook of Seafood and Seafood Products Analysis*, ed. by Nollet LML and Toldrá F. CRC Press,
409 pp 463-480 (2010).
- 410 18. Gibson DM and Ogden ID, Total Viable Counts, in *Methods to Determine the Freshness of Fish*
411 *in Research and Industries, Proceedings of the Final Meeting of the Concerted Action "Evaluation of*
412 *Fish Freshness"*, AIR3CT94-2283, ed. by Olafsdottir G, Luten J, Dalgaard P, Carreche M, Verrez-
413 Bagnis V, Martinsdóttir E and Heia K, Nantes (France), pp 147-150 (1997).
- 414 19. Howgate P, A review of the kinetics of degradation of inosine monophosphate in some species
415 of fish during chilled storage. *International journal of food science & technology* **41**:341-353 (2006)
- 416 20. Lopez-Amaya C and Marangoni A, Phospholipases, in *Seafood Enzymes: Utilization and*
417 *Influence on Postharvest Seafood Quality*, ed. by Haard NF and Simpson BK. CRC Press, pp 91-120
418 (2000).
- 419 21. Gram L and Dalgaard P, Fish spoilage bacteria—problems and solutions. *Current opinion in*
420 *biotechnology* **13**:262-266 (2002)
- 421 22. Sotelo CG and Rehbein H, TMAO-Degrading Enzymes, in *Seafood Enzymes: Utilization and*
422 *Influence on Postharvest Seafood Quality*, ed. by Haard NF and Simpson BK. CRC Press, pp 167-190
423 (2000).
- 424 23. Dos Santos J-P, Iobbi-Nivol C, Couillault C, Giordano G and Méjean V, Molecular analysis of
425 the trimethylamine N-oxide (TMAO) reductase respiratory system from a *Shewanella* species. *Journal*
426 *of molecular biology* **284**:421-433 (1998)
- 427 24. Marzo A, Monti N, Ripamonti M, Muck S and Martelli EA, Determination of aliphatic amines
428 by gas and high-performance liquid chromatography. *Journal of Chromatography A* **507**:241-245 (1990)
- 429 25. Ólafsdóttir G and Jónsdóttir R, Volatile aroma compounds in Fish, in *Handbook of Seafood and*
430 *Seafood Products Analysis*, ed. by Nollet LML and Toldrá F. CRC Press, pp 97-117 (2010).
- 431 26. Timm M and Jørgensen BM, Simultaneous determination of ammonia, dimethylamine,
432 trimethylamine and trimethylamine-N-oxide in fish extracts by capillary electrophoresis with indirect
433 UV-detection. *Food chemistry* **76**:509-518 (2002)
- 434 27. Veciana-Nogues M, Albala-Hurtado M, Izquierdo-Pulido M and Vidal-Carou M, Validation of a
435 gas-chromatographic method for volatile amine determination in fish samples. *Food chemistry* **57**:569-
436 573 (1996)

- 437 28. Béné A, Hayman A, Reynard E, Luisier J and Villettaz J, A new method for the rapid
438 determination of volatile substances: the SPME-direct method: Part II. Determination of the freshness of
439 fish. *Sensors and Actuators B: Chemical* **72**:204-207 (2001)
- 440 29. Li X, Zeng Z, Zhou J, Gong S, Wang W and Chen Y, Novel fiber coated with amide bridged-
441 calix [4] arene used for solid-phase microextraction of aliphatic amines. *Journal of Chromatography A*
442 **1041**:1-9 (2004)
- 443 30. Chan ST, Yao MW, Wong Y, Wong T, Mok C and Sin DW, Evaluation of chemical indicators for
444 monitoring freshness of food and determination of volatile amines in fish by headspace solid-phase
445 microextraction and gas chromatography-mass spectrometry. *European Food Research and Technology*
446 **224**:67-74 (2006)
- 447 31. Chung S and Chan B, Trimethylamine oxide, dimethylamine, trimethylamine and formaldehyde
448 levels in main traded fish species in Hong Kong. *Food Additives and Contaminants: Part B* **2**:44-51
449 (2009)
- 450 32. Barbosa-Pereira L, Otero-Pazos P, De Quirós A, Sendon R, Vecino X, Cruz J, Romero-
451 Rodriguez M, Estevez N, Maroto J and Paseiro-Losada P, SPME-GC Method for the Determination of
452 Volatile Amines as indices of freshness in fish samples. *Italian Journal of Food Science* **24** (2012)
- 453 33. European Commission, Regulation (EC) No. 2074/2005 of 5 December 2005. *Official Journal*
454 *of European Union* **L338**:27-59 (2005)
- 455 34. Leduc F, Krzewinski F, Le Fur B, N'Guessan A, Malle P, Kol O and Duflos G, Differentiation
456 of fresh and frozen/thawed fish, European sea bass (*Dicentrarchus labrax*), gilthead seabream (*Sparus*
457 *aurata*), cod (*Gadus morhua*) and salmon (*Salmo salar*), using volatile compounds by SPME/GC/MS.
458 *Journal of the Science of Food and Agriculture* **92**:2560-2568 (2012)
- 459 35. Cardenas Bonilla A, Sveinsdottir K and Martinsdottir E, Development of Quality Index Method
460 (QIM) scheme for fresh cod (*Gadus morhua*) fillets and application in shelf life study. *Food control*
461 **18**:352-358 (2007)
- 462 36. Zwietering M, De Koos J, Hasenack B, De Witt J and Van't Riet K, Modeling of bacterial
463 growth as a function of temperature. *Applied and Environmental Microbiology* **57**:1094-1101 (1991)
- 464 37. Hall Jr H, Correlation of the base strengths of amines. *Journal of the American Chemical*
465 *Society* **79**:5441-5444 (1957)
- 466 38. Jia M, Zhang QH and Min DB, Optimization of solid-phase microextraction analysis for
467 headspace flavor compounds of orange juice. *Journal of Agricultural and Food Chemistry* **46**:2744-
468 2747 (1998)
- 469 39. Howgate P, A critical review of total volatile bases and trimethylamine as indices of freshness of
470 fish. Part 2. Formation of the bases, and application in quality assurance. *Electronic Journal of*
471 *Environmental, Agricultural and Food Chemistry* **9**:58-88 (2010)
- 472 40. Oehlenschläger J, Suitability of ammonia-N, dimethylamine-N, trimethylamine-N,
473 trimethylamine oxide-N and total volatile basic nitrogen as freshness indicators in seafoods, in *Methods*
474 *to Determine the Freshness of Fish in Research and Industries, Proceedings of the Final Meeting of the*
475 *Concerted Action "Evaluation of Fish Freshness"*, AIR3CT94-2283, ed. by Ólafsdóttir G, Luten J,
476 Dalgaard P, Carreche M, Verrez-Bagnis V, Martinsdóttir E and Heia K, Nantes (France), pp 92-99
477 (1997).
- 478

Table 1: ANOVA results obtained to assess the impact of three parameters on the shapes of both trimethylamine and dimethylamine peaks.

	Trimethylamine ^a		Dimethylamine ^b	
	FWHM ^c	TF ^d	FWHM ^c	TF ^d
TI: Temperature of injection	< 0.0001***	< 0.0001***	< 0.0001***	< 0.0001***
TC: Initial temperature of the column	0.0053**	0.74	< 0.0001***	< 0.0001***
S: Split value for injection	< 0.0001***	< 0.0001***	< 0.0001***	< 0.0001***
TI x TC	0.0036**	0.05*	< 0.0001***	< 0.0001***
TI x S	< 0.0001***	< 0.0001***	0.0007***	< 0.0001***
TC x S	0.07	0.94	0.0012**	< 0.0001***
TI x TC x S	0.77	0.0023**	0.0003***	< 0.0001***

ANOVA described here were achieved after normalization of the factors using the log (X+1) normalization method. ^a Data of ANOVA with trimethylamine pure solution, for each combination of split levels, injection and column temperatures, 5 replicated measurements were performed. ^b Data of ANOVA with dimethylamine pure solutions, for each combination of split levels, injection and column temperatures, 5 replicated measurements were performed. ^{c,d} Influence of parameters and combination of parameters on full width half maximum ^c and tailing factor ^d shape indicators; asterisks illustrate the significance level of the parameters impacts, in accordance with probabilities (*** p<0.001, ** p<0.01, * p<0.05).

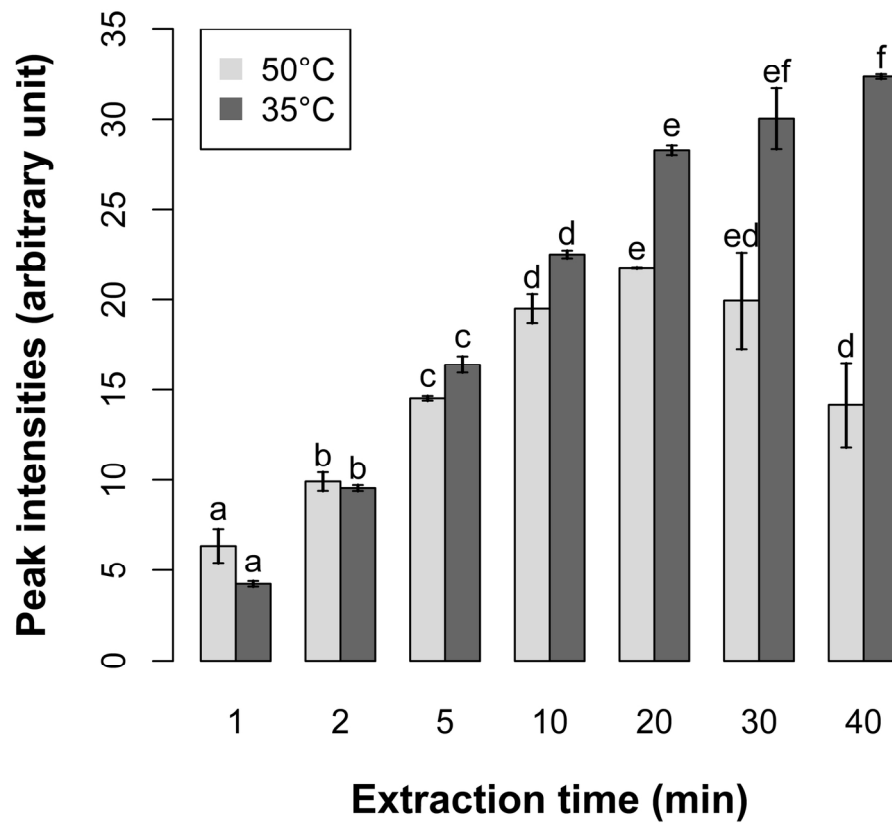


Figure 1: Bar plot of TMA peak intensities depending on extraction time and temperature of extraction from pure solutions. Error bars indicate the min/max peak intensities as measurements were carried out in duplicate. LSD tests were carried out in series (50°C and 35°C). Letters must be compared within a considered series.

79x79mm (600 x 600 DPI)

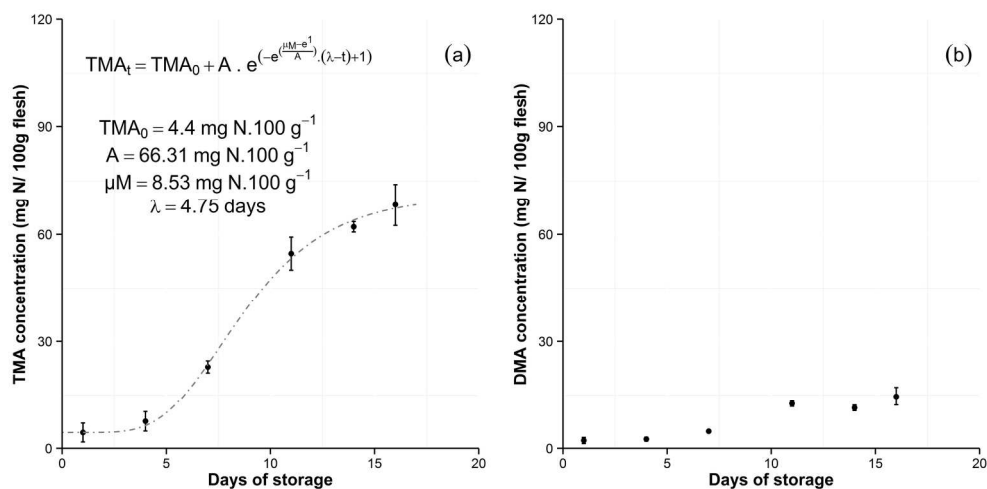


Figure 2: Evolution of mean TMA-N (a) and DMA-N (b) concentrations (mg N/100g) measured by SPME-GC-MS (n=9) throughout the storage period. Each dot reflects the mean value observed for three fillets analysed in triplicate. Concerning TMA-N production modelling, parameters, obtained after 20 iterative steps, correspond to maximum theoretical concentration (A), TMA-N concentration maximum daily increase during the exponential phase (μ), TMA-N concentration lag phase (λ) and initial TMA-N concentration (TMA₀).

95x47mm (600 x 600 DPI)

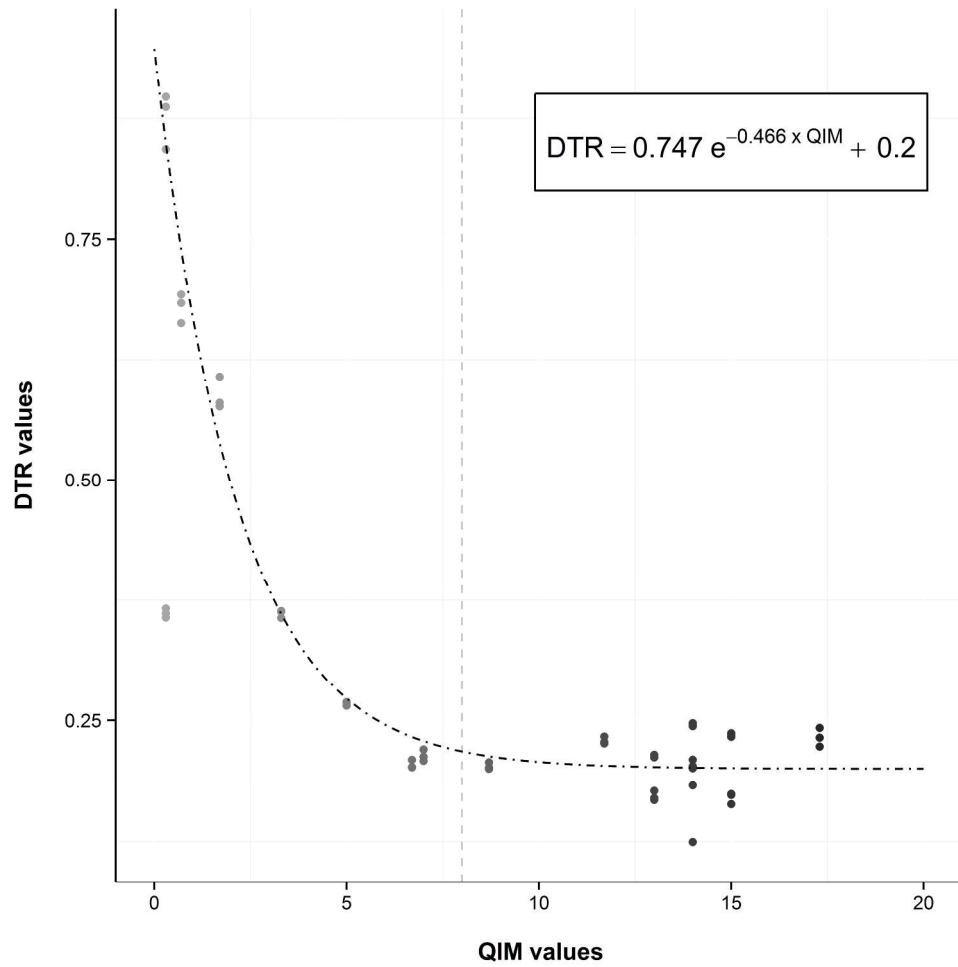


Figure 3: Dot plot showing the evolution of DMA-N/TMA-N ratio (DTR) values as a function of QIM values. Each dot corresponds to a single analysis repeat for a given fillet. The dark grey dot-dashed line illustrates the modelling of data using an exponential equation. The grey dashed line illustrates the estimated maximum QIM value regarding the work of Cardenas Bonilla et al. 35
133x133mm (600 x 600 DPI)

Development of SPME-GC-MS method for the specific quantification of dimethylamine and trimethylamine: use of a new ratio for the freshness monitoring of cod fillets.

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Algorithm S1: Modelling of TMA values

```

1
2
3
4
5 #Algorithm written by A.Dehaut
6 #adapted from the very interesting course of J.R Lobry available (in
7 #French) at http://pbil.univ-lyon1.fr/R/pdf/tdr46.pdf
8
9 #Assignment of the working directory in R_subfolder#
10
11 setwd('X:/R_folder/R_subfolder')
12
13 #Assignment of the file R_logistic, where header is text#
14
15 donnee<-read.csv2('R_logistic.csv',header=T)
16 attach(donnee)
17
18 #Function called "logistique" based on modified Gompertz model#
19
20 logistique <- function(t, param)
21 {
22     a <- param[1]
23     mu <- param[2]
24     l <- param[3]
25     b <- (mu*exp(1))/a
26     c <- (1-t)
27     return(est_TMA_min+a*exp(-exp(b*c+1)))
28 }
29
30 #####
31
32 #Definition of a new data frame with data on Days and TMA
33
34 da<-as.matrix(rbind(Days,TMA), dimnames = NULL)
35
36 #Definition of the function called "scelog" allowing finding the sum
37 #of squares of deviations #
38
39 scelog <- function(param, data)
40 {
41     tobs <- da[1,]
42     xobs <- da[2,]
43     xtheo <- sapply(tobs, logistique, param)
44     return(sum((xobs - xtheo)^2))
45 }
46
47 #####
48
49 #Use of the nlm function allowing finding the parameters that
50 #minimize the sum of squares of deviations #
51
52 nlm(f = scelog, p = c(a_est, mu_est, l_est), data = da)
53
54 #Recovery of the estimates of param a,mu and l.
55
56 est <- nlm(f = scelog, p = c(a_est, mu_est, l_est), data =
57 da)$estimate
58
59
60

```


Algorithm S2: Modelling of the DTR values

```

1
2
3
4
5
6 #Algorithm written by A.Dehaut
7 #adapted from the very interesting course of J.R Lobry available (in
8 #French) at http://pbil.univ-lyon1.fr/R/pdf/tdr46.pdf
9
10 #Assignment of the working directory in R_subfolder#
11
12 setwd('X:/R_folder/R_subfolder')
13
14 #Assignment of the file R_logistic, where header is text#
15
16 donnee<-read.csv2('R_logistic.csv',header=T)
17 attach(donnee)
18
19 #Function called "exponentielle" #
20
21 exponentielle <- function(t, param)
22 {
23     a <- param[1]
24     b <- param[2]
25     return(DTR_min+a*exp(-(b*t)))
26 }
27
28 #####
29
30 #Definition of a new data frame with data on Days and TMA
31
32 da<-as.matrix(rbind(RDT,QIM), dimnames = NULL)
33
34 #Definition of the function called "scelog" allowing finding the sum
35 #of squares of deviations #
36
37 scelog <- function(param, data)
38 {
39     tobs <- da[2,]
40     xobs <- da[1,]
41     xtheo <- sapply(tobs, exponentielle, param)
42     return(sum((xobs - xtheo)^2))
43 }
44
45 #####
46
47 #Use of the nlm function allowing finding the parameters that
48 #minimize the sum of squares of deviations #
49
50 nlm(f = scelog, p = c(a_est, b_est), data = da)
51
52 #Recovery of the estimates of param a,mu and l.
53
54 est <- nlm(f = scelog, p = c(a_est, b_est), data = da)$estimate
55
56
57
58
59
60

```

Outcomes from the nlm function after modelling of TMA_{Steam Distillation} TMA_{SPME} & DTR:

```

      TMASteam Distillation
$minimum
[1] 1009.049
$estimate
[1] 56.802525 7.802283 4.149119
$gradient
[1] -8.810839e-07 1.293319e-05 -
3.263012e-05
$code
[1] 1
$iterations
[1] 20

      TMASPME
$minimum
[1] 1394.435
$estimate
[1] 66.310266 8.533023 4.746776
$gradient
[1] 0.0000207965 0.0001402129 -
0.0002908528
$code
[1] 1
$iterations
[1] 20

DTR (atypical fillet having been
      excluded)
$minimum
[1] 0.05380738
$estimate
[1] 0.7468669 0.4660460
$gradient
[1] -3.146580e-07 -2.852996e-07
$code
[1] 1
$iterations
[1] 3
```

Figure S1: TMA and DMA calibration curves

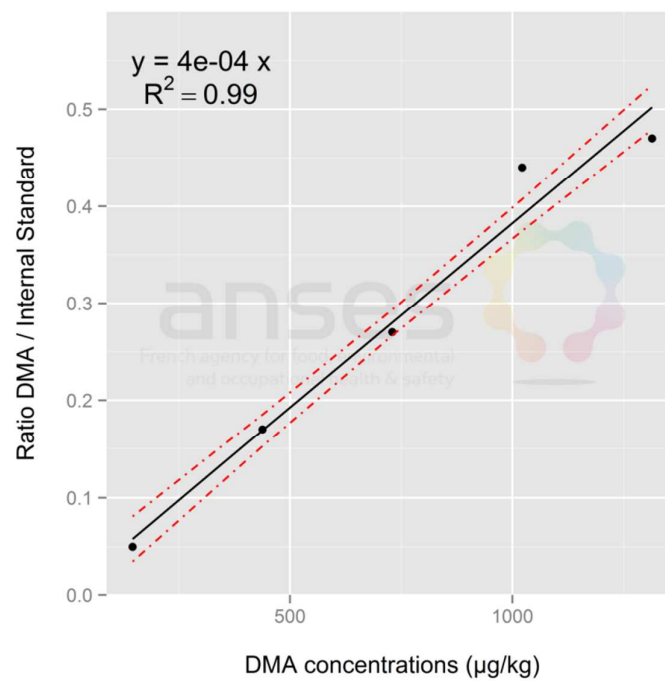
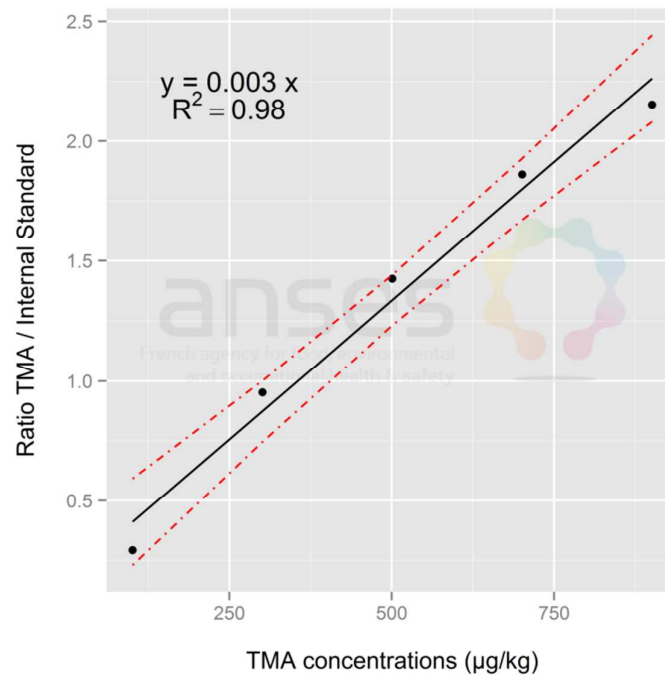
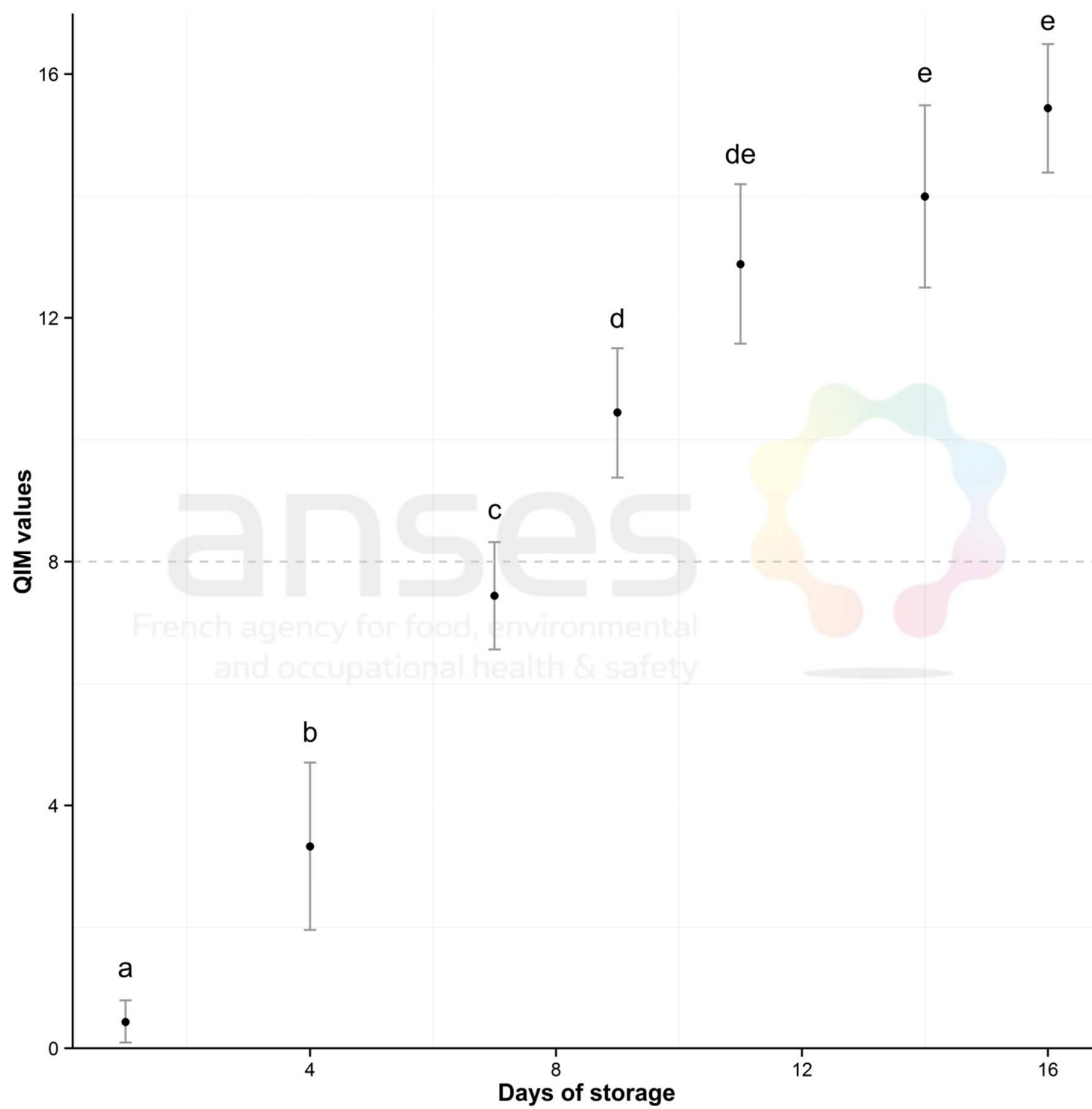
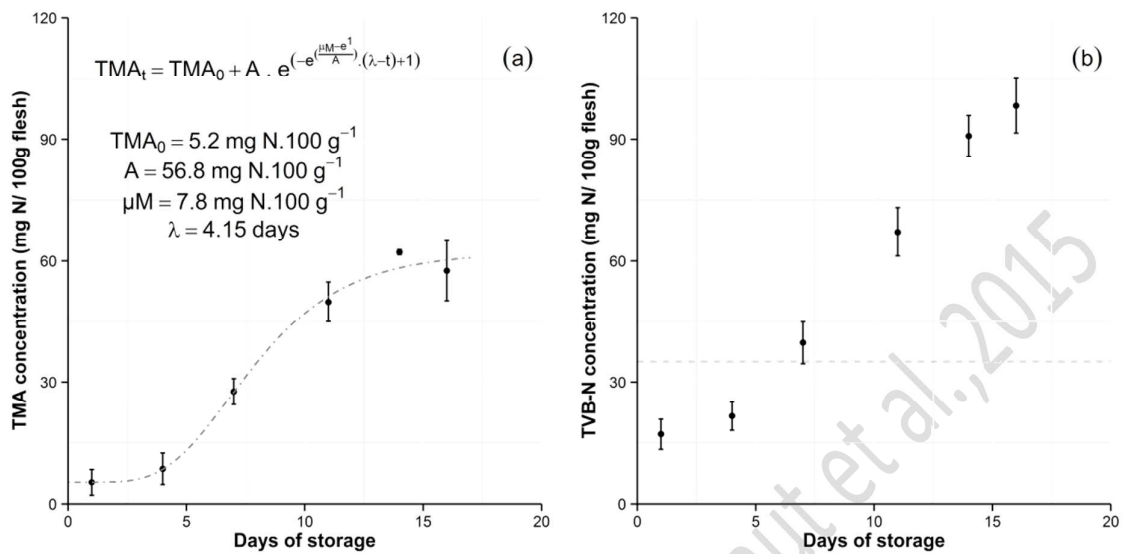


Figure S2: QIM score evolution throughout the storage



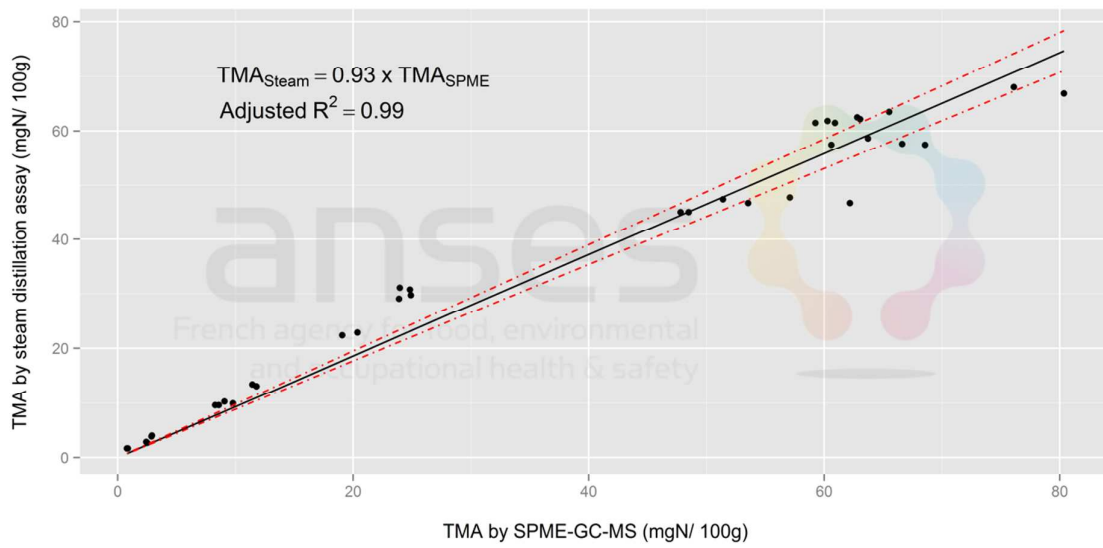
Dot plot showing the average QIM scores (n=9) of the three samples evaluated per storage day, by the panellists, throughout the storage. Error bars illustrate 95% confidence intervals of the mean (two standard errors). The grey dashed line illustrates the estimated maximum QIM value regarding the work of Cardenas Bonilla *et al.*³⁵ Letters over dots correspond to the results of LSD test.

Figure S3: Evolution of TMA-N and TVB-N evaluated by steam-distillation assays



Evolution of mean TMA (a), TVB-N (b) concentrations (mg N/100g) measured by steam-distillation assay (n=6); each dot reflects the mean value observed for three fillets analysed in duplicate. Concerning TVB-N, the grey dashed line refers to the limit value set by EC Regulation 2074/2005.

Figure S4: Relation between TMA_{steam distillation} and TMA_{SPME} values



Supporting informations Dehian
2020
Review