

Occurrence and effects of plastic additives on marine environments and organisms: a review

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- 1 Occurrence and effects of plastic additives on marine environments and
- 2 organisms: a review
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Abstract

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Plastics debris, especially microplastics, have been found worldwide in all marine compartments. Much research has been carried out on adsorbed pollutants on plastic pieces and hydrophobic organic compounds (HOC) associated with microplastics. However, only a few studies have focused on plastic additives. These chemicals are incorporated into plastics from which they can leach out as most of them are not chemically bound. As a consequence of plastic accumulation and fragmentation in oceans, plastic additives could represent an increasing ecotoxicological risk for marine organisms. The present work reviewed the main class of plastic additives identified in the literature, their occurrence in the marine environment, as well as their effects on and transfers to marine organisms. This work identified polybrominated diphenyl ethers (PBDE), phthalates, nonvlphenols (NP), bisphenol A (BPA) and antioxidants as the most common plastic additives found in marine environments. Moreover, transfer of these plastic additives to marine organisms has been demonstrated both in laboratory and field studies. Upcoming research focusing on the toxicity of microplastics should include these plastic additives as potential hazards for marine organisms, and a greater focus on the transport and fate of plastic additives is now required considering that these chemicals may easily leach out from plastics.

Keywords

33 Microplastics, Plastic additives, Bisphenol A, Phthalates, Brominated Flame retardant

1. Introduction

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Due to their numerous societal benefits, plastics hold an important place in human society (Andrady and Neal, 2009). Plastic, a man-made material, is inexpensive, strong, durable, lightweight and easy to produce (Thompson et al., 2009). As a consequence, plastic production has been increasing since the 1950s, and notably rose from 225 million tons in 2004 to 322 million tons in 2015, representing a 43% increase over the last decade (PlasticsEurope, 2016). However, this estimate does not take into account the proportion of synthetic fibers which are widely used in the textile and fishery industries (Dris et al., 2016) and there is an underestimation of 15% to 20% depending on the year (Industrievereinigung Chemiefaser, 2013). Low estimates predicted that floating marine plastic weigh between 70,000 and 270,000 tons (Cózar et al., 2014; Eriksen et al., 2014; Van Sebille et al., 2015). Small pieces of plastics called microplastics (MP) account for a total of 51 trillion plastic debris (Van Sebille et al., 2015). Microplastics have been defined as plastics particles smaller than 5 mm (Arthur et al., 2009). Growing attention has been accorded to microplastics during the last decade, since the publication by Thompson et al. (2004). Micro-sized plastic pieces originate from two distinct pathways: primary and secondary sources. Primary sources of MP correspond to (i) plastics that are directly manufactured at micrometric size, including plastic pellets (Barnes et al., 2009; Cole et al., 2011), (ii) MP from exfoliating cosmetics (Chang, 2015; Fendall and Sewell, 2009; Napper et al., 2015; Zitko and Hanlon, 1991) and (iii) clothing fibers found in wastewater treatment plants (Browne et al., 2011; Carr et al., 2016). Secondary MP results from the breakdown of larger pieces due to mechanical abrasion and photochemical oxidation in the environment (Andrady, 2011; Bouwmeester et al., 2015; Lambert and Wagner, 2016). MP can also degrade into smaller pieces called nanoplastics (Gigault et al., 2016; Koelmans et al., 2015; Lambert and Wagner, 2016).

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Due to their small size, MP can be ingested by a wide range of marine organisms such as zooplankton, bivalves and worms (De Witte et al., 2014; Devriese et al., 2015; Graham and Thompson, 2009; Rochman et al., 2015; Sussarellu et al., 2016; Van Cauwenberghe and Janssen, 2014; Van Cauwenberghe et al., 2015) and by organisms from higher trophic levels such as fish (Boerger et al., 2010; Carpenter et al., 1972; Dantas et al., 2012; Foekema et al., 2013; Lusher et al., 2013; Neves et al., 2015; Possatto et al., 2011; Rochman et al., 2015) and marine mammals (Eriksson and Burton, 2003; Lusher et al., 2015). This ingestion of MPs can result in physical damage such as obstruction or internal abrasions (Wright et al., 2013). In addition to these physical threats, MP can potentially transfer chemicals adsorbed on their surface (Mato et al., 2001; Teuten et al., 2007; Teuten et al., 2009) or plastic additives. However, less attention has been paid to the transfer of plastic additives to marine organisms in comparison with hydrophobic organic compounds (HOC), despite the fact that many additives have been recognized as hazardous (Lithner et al., 2011). Therefore, the transport and fate of plastic additives leaching out from plastic debris should definitely be carefully addressed in future field, laboratory and modelling works. Plastics are made by polymerizing monomers and other substances (Lithner et al., 2011) including plastic additives. Plastic additives are chemical compounds, like plasticizers, which provide required properties to a plastic polymer or are incorporated to facilitate the manufacturing process (OECD, 2004). Moreover, some plastic additives are used as monomers, for example bisphenol A is the monomer of polycarbonate (PC) but also a stabilizer in other polymers. Plastic additives are mainly used as plasticizers, flame retardants, stabilizers, antioxidants and pigments. Phthalates, BPA, nonylphenols, and brominated flame retardants (BFR) are the most common additives recovered from the environment (Bergé et al., 2012; David et al., 2009; de Boer et al., 1998; de los Ríos et al., 2012; Mackintosh et al., 2004; Net et al., 2015; Xie et al., 2005; Xie et al., 2007) and represent a hazard to the environment and

organisms (Lithner et al., 2011; Meeker et al., 2009; Oehlmann et al., 2009). These plastic additives are released into the marine environment by numerous pathways including industrial and municipal wastewater, atmospheric deposition, runoff and river transport resulting from application of sewage sludge in agriculture. In addition leaching of plastic additives from macro and microplastics is known to occur in the marine environment. Thus, the accumulation and degradation of plastic debris might represent another major input of these chemical compounds in oceans. As a consequence, more research is needed on the hazards of plastic additives associated with microplastics.

The aim of this review is to (i) list and describe the most predominant plastic additives used worldwide in the plastic industry, (ii) present an overview of the occurrence of plastics additives in the marine environment, and (iii) document the effects of plastic additives on marine organisms. Lastly, recommendations will be made in order to identify the polymer-additives pairs of major concern on which further research should focus.

2. Chemicals used as plastic additives

Multiple types and families of chemicals are mixed with polymers to produce plastics. The type of additive depends on the plastic polymer and the requirements of the final product (Table 1).

Table 1: List of the most commonly produced polymers and their associated plastic additives. Adapted from Hansen *et al.* (2013)

Polymer	Consumption in the EU27 (in million tons) in 2015 ¹	Additive types	Amount in polymers (% w/w)	Hazardous substances ²
DD	0	Antioxidant	0.05 – 3	Bisphenol A; Octylphenol; Nonylphenol
PP 9	Flame retardant (cable insulation and electronic applications)	12 – 18	Brominated flame retardant; Boric acid; Tris(2-chloroethyl)phosphate	
HDDE		Antioxidant	0.05 - 3	Bisphenol A; Octylphenol; Nonylphenol
HDPE 8	8	Flame retardant (cable insulation application)	12 -18	Brominated flame retardant; Boric acid; Tris(2-chloroethyl)phosphate
LDPE	6	Antioxidant	0.05 - 3	Bisphenol A; Octylphenol; Nonylphenol
LDPE	6	Flame retardant (cable insulation application)	12 – 18	Brominated flame retardant; Boric acid; Tris(2-chloroethyl)phosphate
DVC	5	Plasticizer	10 - 70	Phthalate
PVC	5	Stabilizer	0.5 - 3	Bisphenol A; Nonylphenol
PUR	3.5	Flame retardant	12 - 18	Brominated flame retardant; Boric acid; Tris(2-chloroethyl)phosphate

¹ According to PlasticsEurope (2016); PP: Polypropylene; HDPE: High Density Polyethylene; LDPE: Low Density Polyethylene; PVC: Polyvinyl Chloride; PUR: Polyurethane.

² Hazardous substances refer to chemicals that pose a risk to the environment and to human health as defined by the REACH regulation in the European Union according to the European Chemical Agency (2017).

The following section describes the most common additive types used in the manufacturing processes (Table 2) that have been reported in macro- and microplastic debris collected in environmental surveys: brominated flame retardants, phthalates used as plasticizers, nonylphenols, bisphenol A and antioxidants.

Table 2: List of common plastic additives and their associated functions and potential effects

Additives	Function	Effects
Brominated	Reduce flammability in plastic. Also	Potential endocrine disruptors
Flame	adsorbed on plastic from the surrounding	
Retardants	environment.	
(BFR)		
Phthalates	Plasticizers to soften plastic mainly in	Endocrine disruptors
	polyvinyl chloride.	
Nonylphenol	Antioxidant and plasticizer in some plastics	Endocrine disruptors
Bisphenol A	Monomer in polycarbonate and epoxy	Endocrine disruptors
(BPA)	resins.	Estrogen mimic
	Antioxidant in some plastics.	
Irganox®	Antioxidant in some plastics.	

The main plastic additives described are listed in Table 3 with their associated octanol-water partition coefficient (K_{ow}). K_{ow} has been used for predicting how a chemical will concentrate in marine organisms and an increase in log K_{ow} indicates an increase in the potential bioconcentration in organisms (Net *et al.*, 2015).

Table 3: Plastic additives and their associated octanol-water partition coefficients (Log K_{ow}). Data were extracted from the following reviews: Bergé *et al.* (2012), Espinosa *et al.* (2016), Net *et al.* (2015) and Oehlmann *et al.* (2008)

Full name	Abbreviation	$Log\;K_{\rm ow}$
butyl benzyl phthalate	BBP	4.70
di(2-ethylexyl) phthalate	DEHP	7.73
diethyl phthalate	DEP	2.54
diisobutyl phthalate	DiBP	4.27
diisodecyl phthalate	DiDP	9.46
diisononyl phthalate	DiNP	8.60
dimethyl phthalate	DMP	1.61
di-n-butyl phthalate	DnBP	4.27
di-n-octyl phthalate	DnOP	7.73
hexabromocyclododecane	HBCD	5.07 - 5.47
polybrominated diphenyl ether	PBDE	5.52 - 11.22
tetrabromobisphenol A	TBBPA	4.5
bisphenol A	BPA	3.40
nonylphenol	NP	4.48 - 4.80

2.1. Brominated flame retardants

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Brominated flame retardants (BFR) are a class of additives used in plastic products to reduce flammability. These BFR are used in a variety of consumer products ranging from electronic devices to insulation foams. BFRs include a wide range of chemicals, of which polybrominated diphenyl ethers (PBDE), hexabromocyclododecane (HBCD - Pubchem ID: 18529) and tetrabromobisphenol A (TBBPA – Pubchem ID: 6618) (Talsness et al., 2009) represent the main BFRs used in the plastic industry. These 3 classes (PBDE, HBCD and TBBPA) are described in details below. Lately, attention has been given to other emerging BFRs such as 1,2-bis(2,4,6-tribromophenoxy)ethane Pubchem ID: 37840), (BTBPE decabromodiphenylethane (DBDPE - Pubchem ID: 10985889) and hexabromobenzene (HBB - Pubchem ID: 6905) as these have been identified in many environmental compartments, organisms, food and humans (European Food Safety Authority, 2012). As they are not chemically bound to the polymer matrix, they can leach into the surrounding environment (Engler, 2012; Meeker et al., 2009) with an exception for TBBPA which is chemically bound

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131 to the polymer (Morris et al., 2004). 132 PBDEs are hydrophobic substances that include numerous formulations used in plastics as 133 flame retardants. Indeed, there are three main commercial formulations called penta-, octa- and 134 deca-BDEs (Chua et al., 2014). These additives are ubiquitous, toxic, persistent and 135 bioaccumulate in the environment and are of great concern for human health (Engler, 2012). 136 As a result, penta- and octa-BDEs have been banned by the European Union since 2004 137 (European Directive, 2003), while deca-BDE was banned only in 2009 from electronic and 138 electrical applications in the European Union (European Council Decision, 2009). These 139 formulations can no longer be used in mixtures or products with a concentration higher than 140 0.1% by mass. In addition, tetra- to hepta-BDEs are listed for elimination in the Annex A of the 141 Stockholm Convention on persistent organic pollutant (POP) (Stockholm Convention, 2016). 142 Moreover, in Canada the use of tetra- to deca- BDE has been restricted under the SOR/2008-143 218 Regulations (Consolidated Regulation, 2008). Since 2006, penta- to octa- BDE have been 144 subjected to a 90 day notification before importation or production in the US. Finally, deca-145 BDE importation and production have been entirely stopped (US Environmental Protection 146 Agency, 2006, 2012) since 2013. 147 HBCD has three dominant stereoisomers: α-, β-, and γ-HBCD (European Food Safety 148 Authority, 2011a). These BFRs are listed as POPs in the Stockholm Convention (Stockholm 149 Convention, 2016) and the three isomers are subject to a request for authorization in the 150 European Union (European Council Regulation, 2006). HBCDs are found in expanded PS 151 (EPS) and extruded PS (XPS) up to 4-7% by weight (Al-Odaini et al., 2015). Its use has been 152 subjected to authorization in the European Union since 2006 in the annex XIV of the 153 Registration, Evaluation, Authorisation and Restriction of Chemicals (REACH) regulation 154 (European Food Safety Authority, 2011a), Moreover, in 2013 HBCD was listed for elimination in the Annex A of the Stockholm Convention with specific exemption for use and production in EPS and XPS (Cruz *et al.*, 2015; Stockholm Convention, 2016). In the US, Environmental Protection Agency conducted a risk assessment for HBCD according to the 2010 "Toxic Substances Control Act" action plan (US Environmental Protection Agency, 2010b).

TBBPA is produced by brominating bisphenol A (European Food Safety Authority, 2011b). According to the European Food Safety Authority (2011b), TBBPA is the most commonly produced BFR in the world and represents 60% of the BFR market. This BFR is used in acrylonitrile butadiene styrene (ABS) and in other plastic such as high impact PS and phenolic resin (Cruz *et al.*, 2015). Until now, no legislation concerning TBBPA has been applied in the European Union (Vandermeersch *et al.*, 2015).

2.2. Phthalates

Phthalic acid esters (PAE) or phthalates are a family of plastic additives used as plasticizers, mainly in PVC production (Arbeitsgemeinschaft PVC und Umwelt e.V, 2006). As a result, PVC can contain 10% to 60% phthalates by weight (Net *et al.*, 2015). As phthalates are not chemically bound to the polymer matrix, they can easily leach into the environment during manufacturing, use and disposal (Net *et al.*, 2015). PAEs have been found in a wide range of environments (as reviewed by Net *et al.* (2015)) and this is of concern, since some phthalates have been defined as endocrine disruptors, even at low concentrations (Oehlmann *et al.*, 2009).

In 2015, 8.4 million tons of plasticizers were used around the world, and di(2-ethylexyl) phthalate (DEHP – Pubchem ID: 8343) was the most commonly used plasticizer, representing 37.1% of the global plasticizer market (ECPI, 2016). Europe accounted for 1.3 million tons of the global plasticizer market in 2015 (ECPI, 2016), but DEHP was not the most commonly used plasticizer in Europe, as suggested by its 20% decrease in consumption between 1999 and 2004. DEHP has gradually been replaced by diisononyl phthalate (DiNP – Pubchem ID: 590836),

diisodecyl phthalate (DiDP – Pubchem ID: 33599) and di(2-Propyl Heptyl) phthalate (DPHP – Pubchem ID: 92344), which represented 57% of plasticizer consumption in Europe in 2015 (Arbeitsgemeinschaft PVC und Umwelt e.V, 2006; ECPI, 2016).

2.3. Bisphenol A

BPA (Pubchem ID: 6623) is the most representative chemical of the bisphenol group and is one of the most commonly produced chemicals worldwide, with over three million tons produced annually (Laing *et al.*, 2016). BPA is mainly used as a monomer for polycarbonate (PC) plastics (65% of volume used) and epoxy resins (30% of volume used), which are for instance the main component of the lining layer of aluminum cans (Crain *et al.*, 2007; ICIS, 2003). BPA can also be used as an antioxidant or as a plasticizer in other polymers (PP, PE and PVC) (Rani *et al.*, 2015). Leaching of BPA can occur (Sajiki and Yonekubo, 2003), leading to the release of this additive from food and drink packaging, which is considered as a source of exposure for human beings (Vandermeersch *et al.*, 2015). Despite its potential to leach from food packaging and the fact that it has been identified as a significant endocrine disruptor (Oehlmann *et al.*, 2009), BPA is still allowed in the European Union for use in food contact material (European Council Regulation, 2011). Other bisphenol analogs, such as bisphenol B, bisphenol F and bisphenol S are used in plastics and may represent a threat to the environment even though their toxicity is still unknown (Chen *et al.*, 2016).

2.4. Nonylphenols

Nonylphenols (NP) are intermediate products of the degradation of a widely used class of surfactants and antioxidants: nonylphenol ethoxylates (NPE) (Engler, 2012). NP and NPE are organic chemicals used for many applications such as paints, pesticides, detergents and personal care products (US Environmental Protection Agency, 2010a). They can also be used as antioxidants and plasticizers for the production of plastics (Rani *et al.*, 2015; US Environmental

Protection Agency, 2010a). Furthermore, NP have been found to leach out from plastic bottles to their water content (Loyo-Rosales *et al.*, 2004). Moreover, effluents from wastewater treatment plants are the major source of NP and NPE in the environment (Soares *et al.*, 2008). NP are considered as endocrine disruptors and their use is prohibited in the European Union due to their effects on the environment and human health (Rani *et al.*, 2015).

2.5. Antioxidants

Antioxidants are used as additives in many synthetic polymers including polyolefins (mainly PE and PP) which represent 60% of global demand for antioxidant additives (Höfer, 2012). Antioxidants are used to prevent the ageing of plastics and to delay oxidation (Lau and Wong, 2000). However, as with other plastic additives, antioxidants can leach out of the plastic and can migrate to food from plastic packaging and pose a threat in terms of food safety (Lau and Wong, 2000). Antioxidants from the Irganox® series are commonly used in plastics and they include Octadecyl 3-(3,5-di-*tert*-butyl-4-hydroxyphenyl)propionate (Irganox® 1076 – Pubchem: 16386), Pentaerythrityl-tetrakis-3-(3,5-di-*tert*-butyl-4-hydroxyphenyl)propionate (Irganox® 1010 – Pubchem ID: 64819) and 2,4-di-*tert*-butylphenol (Irgafos® 168 – Pubchem ID: 91601) (Lau and Wong, 2000).

Owing to the variety of plastic additives (BFR, phthalates, BPA, NP and antioxidants) used for plastic products conception and their detection in macro- and microplastic debris collected in environmental surveys, their occurrence in environmental matrices (water, sediment, biota) is expected and may pose major environmental concern as described below.

3. Plastic additives in the environment

3.1. Marine waters

Marine waters are affected by anthropogenic pressures as this natural compartment is the final receptacle of all discharge waters. Consequently, chemical pollutants including plastics additives have been detected in worldwide marine waters (Tables 4, 5 and 6) (Bergé *et al.*, 2013; Net *et al.*, 2015). Of all BFR, PBDE are the most commonly studied molecules in marine environments. PBDE have been widely found and multiple congeners have been monitored (Table 4). Concentrations varied from a few ng L⁻¹ to more than 10 ng L⁻¹ and congeners varied among the studied sites (Table 4).

Table 4: Concentrations of polybrominated diphenyl ether (PBDE) in seawater in ng L-1

Location	ΣΡΒDΕ	Range	BDE congeners	Dominant	References
	(ng L ⁻¹)	(ng L ⁻¹)	detected	congener	
Port sea,	23.2	4.2 – 19	BDE-28, -47	BDE-28	Sánchez-Avila
Mediterranean Sea,					et al. (2012)
Spain					
Surface microlayer,	0.33	0.004 -	BDE-28, -47, -	BDE-156	Wurl et al.
China Sea, Hong-		0.056	99, -100, -156, -		(2006)
Kong			183		
Subsurface water,	0.1	0.002 -	BDE-28, -47, -	BDE-47	Wurl et al.
China Sea, Hong-		0.082	99, -100, -183		(2006)
Kong					

Many studies on the contamination of the marine environment by phthalates showed concentrations ranging from a few pg L⁻¹ to around 10 µg L⁻¹, with DEHP being the most concentrated phthalate found in marine waters (Table 5).

Table 5: Concentrations of phthalates in seawater in $\mu g \ L^{\text{-}1}$

Location	DMP	DEP	DnBP	DiBP	BBP	DEHP	DnOP	Reference
Tees Bay, UK	$< 1 \times 10^{-3}$	0.025 - 0.5	0.47 - 0.55	0.66 - 1.1		0.98 - 2.2		Law et al. (1991)
North Sea,	0.2×10^{-3}	0.67×10^{-2}	1.7×10^{-3}		0.05×10^{-3}	2.2×10^{-3}		Xie et al. (2005)
Germany								
Surface waters,	0.004 -	0.07 - 2.3	<0.066 – 3.1		0.001 - 1.8	0.9 - 5	0.002 -	Vethaak <i>et al.</i> (2005)
the Netherlands	0.49						0.078	
Bay of Biscay,	(7.5 ± 0.4)	$(33 \pm 3) x$	$(83 \pm 7) \times 10^{-3}$		$(8 \pm 1) \times 10^{-1}$	$(64 \pm 4) x$	(3.6 ± 0.4)	Prieto et al. (2007)
Spain	x 10 ⁻³	10 ⁻³	3		3	10^{-3}	x 10 ⁻³	
Coastal seawater,	0.003 -	0.024 -			0.001 -	0.03 - 0.62		Sánchez-Avila et al.
Mediterranean	0.14	0.48			0.10			(2012)
Sea, Spain								
Port sea,	0.004 -	0.024 -			0.003 -	0.06 - 5.97		Sánchez-Avila et al.
Mediterranean	0.012	0.87			0.80			(2012)
Sea, Spain								
River – sea	0.005	0.07 - 0.16			0.003 -	0.02 - 0.21		Sánchez-Avila et al.
interface,					0.07			(2012)

Mediterranean							
Sea, Spain							
Liguarian Sea,						$18.38 \pm$	Fossi et al. (2012)
Mediterranean						44.39	
Sea, Italy ¹							
Sardinian Sea,						23.42 ±	Fossi et al. (2012)
Mediterranean						32.46	
Sea, Italy ¹							
Barkley Sound,			0.18 - 3.0			0.01 - 0.95	Keil et al. (2011)
Canada							
Puget Sound, USA						0.06 - 0.64	Keil et al. (2011)
Klang River						3.10 - 64.3	Tan (1995)
estuary, Australia							
Caspian Sea, Iran	0.49	0.52					Hadjmohammadi et
							al. (2011)
Arctic	40 x 10 ⁻⁶	138 x 10 ⁻⁶	51 x 10 ⁻⁶	22 x 10 ⁻⁶	8 x 10 ⁻⁶	448 x 10 ⁻⁶	Xie et al. (2007)

Nonylphenol was detected in marine waters of Europe, Asia and North America (Bergé *et al.*, 2012; David *et al.*, 2009) (Table 6) and concentrations ranged from $0.2 \times 10^{-5} \,\mu g \, L^{-1}$ in the Sea of Japan to $4.6 \,\mu g \, L^{-1}$ in the Mediterranean Sea (Table 6). BPA, as for other additives, has been globally quantified in marine waters all around the world and concentrations ranged from $ng \, L^{-1}$ in China to $\mu g \, L^{-1}$ in coastal waters of Singapore (Table 6).

Table 6: Ranges of concentrations of nonylphenol and BPA in seawater in $\mu g \; L^{\text{-}1}$

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Chemicals	Location	Concentrations (µg L ⁻¹)	References
	German Bight, North Sea,	0.006 - 0.033	Bester <i>et al.</i> (2001)
	Germany	$9 \times 10^{-5} - 0.0014$	Xie et al. (2006)
	Estuaries, the Netherlands	0.031 - 0.934	Jonkers <i>et al</i> . (2003)
	Estuaries, UK	0.1 - 2.6	Blackburn <i>et al</i> . (1999)
Nonylphenol	Mediterranean Sea, Spain	0.3 – 4.1	Petrovic <i>et al</i> . (2002)
	Venetian Lagoon, Italy	0.004 - 0.211	Pojana <i>et al.</i> (2007)
	Jamaica Bay, US	0.077 - 0.416	Ferguson <i>et al.</i> (2001)
	Masan Bay, South Korea	0.0097 - 0.928	Li et al. (2008)
	C f. I I	$0.2 \times 10^{-5} - 9.3 \times$	Kannan et al.
	Sea of Japan, Japan	10 ⁻⁵	(1998)
	Surface waters, the	0.009 - 1	Vethaak <i>et al.</i> (2005)
BPA	Netherlands	< 0.012 - 0.33	Belfroid <i>et al</i> . (2002)
DrA	Venetian Lagoon, Italy	< 0.001 - 0.145	Pojana et al. (2007)
	Jiaozhou Bay, China	0.001 - 0.092	Fu et al. (2007)
	Estuaries, Japan	0.036 - 0.058	Kawahata <i>et al</i> . (2004)

Coastal waters, Singapore

0.01 - 2.47

Basheer *et al*. (2004)

Overall, plastic additives have been detected worldwide in estuarine and marine waters at concentrations ranging from pg/L to μ g/L with PBDE and DEHP being the most commonly reported congeners among BFR and phthalates, respectively. In addition to BPA and NP are also frequently detected in seawater. As most of the plastic additives exhibit high K_{ow} , higher concentrations are expected in sediment and marine organisms.

3.2. Sediment

As for marine waters, sediments are also affected by anthropogenic discharges and chemicals including plastic additives. Regarding BFRs, multiple BDE congeners have been found in marine sediments with BDE-209 being the major PBDE quantified in most studies at concentrations ranging from ng/kg to mg/kg (Table 7). In the Netherlands, HBCD were also found in sediments from the North Sea and Scheldt Estuary, respectively, at levels of 0.76 to 6.9 µg kg⁻¹ dry weight (dw) and 30 to 71 µg kg⁻¹ dw (Klamer *et al.*, 2005; Verslycke *et al.*, 2005).

Table 7: Concentrations of polybrominated diphenyl ether (PBDE) in marine sediments in µg kg⁻¹ dry weight

	ΣΡΒDΕ	Range				
Location	$(\mu g kg^{-1})$	$(\mu g kg^{-1})$	BDE congeners detected	Most abundant	References	
Location	dry	dry	BDE congeners detected	congener	References	
	weight)	weight)				
North Sea, the	126.3	0.4 - 32	BDE-28, -47, -66, -71, -75, -77, -85, -99, -100, -119, -	BDE-209	Klamer <i>et al.</i> (2005)	
Netherlands	120.3	0.4 - 32	138, -153, -190, -209	BDE-209	Kiaiiici ei al. (2003)	
Scheldt		0.2	DDF 20 47 (/ 05 00 100 120 152 154		Manalanda (1	
Estuary, the	2198	0.2 –	BDE-28, -47, -66, -85, -99, -100, -138, -153, -154, -	BDE-209	Verslycke <i>et al</i> .	
Netherlands		1650	209		(2005)	
Coastal waters,	27.0	0.0037 -	BDE-3, -7, -15, -28, -47, -49, -66, -71, -77, -85, -99, -	DDE 200	M (20071)	
South Korea	27.8	27.4	100, -119, -126, -138, -153, -154, -183, -209	BDE-209	Moon <i>et al.</i> (2007b)	
Industrialized		0.0012	DDE 2 7 15 29 47 40 66 71 77 95 00			
bays, South	357.8	0.0012 -	BDE-3, -7, -15, -28, -47, -49, -66, -71, -77, -85, -99, -	BDE-209	Moon et al. (2007a)	
Korea		283	100, -119, -126, -138, -153, -154, -183, -209			

TBBPA was also found in the Scheldt Estuary at levels below 0.1 μg kg⁻¹ dw (Verslycke *et al.*, 2005). In their study, Klamer *et al.* (2005) also reported the presence of phthalates in North Sea sediments, namely dimethyl phthalate (DMP – Pubchem ID: 8554), diethyl phthalate (DEP – Pubchem ID: 6781), DBP, BBP, DEHP and DOP with DEHP being the most concentrated phthalate with 170 to 3,390 μg kg⁻¹. Phthalates in marine sediments from the Gulf of Mexico were detected on average at 7.6 and 6.6 μg kg⁻¹ dw for di-*n*-butyl phthalate (DnBP – Pubchem ID: 3026) and DEHP respectively (Giam *et al.*, 1978). In Singapore Bay, phthalates reached 890 to 2,790 μg kg⁻¹ dw for DEHP (Chee *et al.*, 1996). For nonylphenol (Bergé *et al.*, 2012; David *et al.*, 2009), concentrations ranged from less than 1 μg kg⁻¹ dw in estuaries in the Netherlands to more than 20,000 μg kg⁻¹ dw in the sediments of Tokyo Bay (Table 8). Like BFRs, NP and phthalates, BPA has also been found worldwide in sediments (Table 8). Indeed, BPA concentrations ranged from a few μg kg⁻¹ dw in Japan and China to hundreds of μg kg⁻¹ dw in the Venetian Lagoon (Table 8).

Whether the plastic additives detected in marine sediments come from diffuse sources (wastewater, atmospheric deposition, sewage sludge, etc.) or leachate from plastic debris is unclear even though an increasing amount of evidence (Al-Odaini *et al.*, 2015) suggests that microplastic and plastic debris in general likely constitute sources of plastic additives in the marine environment.

Table 8: Ranges of concentrations of nonylphenol and BPA in marine sediments in µg kg⁻¹ dry weight, nd: not detected.

Localization	Nonylphenol range (µg kg -1 dry weight)	BPA range (μg kg -1 dry weight)	Reference
North Sea, Germany	13 – 55		Bester <i>et al.</i> (2001)
Estuaries, the Netherlands	0.9 - 1080		Jonkers et al. (2003)
Mediterranean Sea, Spain	18 - 590		Petrovic et al. (2002)
Venetian Lagoon, Italy	47 - 192		Pojana et al. (2007)
Jamaica Bay, US	7 - 13,700		Ferguson et al. (2001)
Southern California Bight, US	130 – 3200		Schlenk et al. (2005)
Masan Bay, South Korea	92 - 557		Li et al. (2008)
Talvus Day, Jaman	142 - 20,700		Kurihara et al. (2007)
Tokyo Bay, Japan	120 - 640		Isobe et al. (2001)
The Netherlands		<1.1 - 43	Vethaak et al. (2005)
Venetian Lagoon, Italy		< 2.0 - 118	Pojana et al. (2007)
Jiaozhou Bay, China		0.7 - 17	Fu et al. (2007)
Estuaries, Japan		nd – 2.7	Kawahata <i>et al</i> . (2004)

3.3. Microplastics

To date, only a few studies have focused on the detection of plastic additives from MP collected in marine environments (Faure *et al.*, 2015; Fries *et al.*, 2013; Hirai *et al.*, 2011; Mato *et al.*, 2001; Rani *et al.*, 2015; Rochman *et al.*, 2014). Mato *et al.* (2001) detected nonylphenols in PP pellets deployed in Tokyo Bay and suggested that these compounds came from plastic additives found in the PP pellets themselves. In another study, Hirai *et al.* (2011) measured high concentrations of PBDEs, BPA and nonylphenols in PE and PP fragments collected on remote or urban beaches and in the open ocean. It was stated that they originated from plastic additives used for the manufacture of PP and PE. A wide range of plastic additives were also identified using Pyrolysis-GC/MS with thermal desorption in MP collected from sediment of Norderney Island (Fries *et al.*, 2013). MP particles were identified as PE, PP, PS and polyamide-6 (PA-6).

They were associated with DEHP, DnBP, diisobutyl phthalate (DiBP – Pubchem ID: 6782), and 2,4-di-*tert*-butylphenol (2,4-DTBP – Pubchem ID: 7311), used here as antioxidant additives for PE and PP, DnBP, DiBP, DEP and DMP for PS, and DEHP, and DiBP and DEP for PA-6 (Fries et al., 2013). Moreover, Rani *et al.* (2015) detected multiple plastic additives in plastic marine debris found on a beach in Geoje, South Korea. Indeed, the authors found BPA and phthalates in PP and PE plastic marine debris as well as antioxidants including Irganox 76 and 2,4-DTBP in PP and PE plastic marine debris. In a study dealing with plastic debris in the Atlantic Ocean, BPA, PBDEs and 4-nonylphenol were detected in plastic samples found at sea and the authors suggested that this chemical came mainly from plastic additives (Rochman *et al.*, 2014). Moreover, some plastic additives were detected at concentrations up to 6 orders of magnitude higher than the concentrations measured in the surrounding water (Rochman *et al.*, 2014). In a more recent study, Faure *et al.* (2015) quantified MP pollution in Swiss lakes and detected MP associated with plastic additives including PBDEs, NPs, BPA and phthalates at concentrations comparable to those reported in marine studies (from 10-1 to 106 ng g-1) (Faure *et al.*, 2015).

These six studies demonstrated that plastic additives, some of which are known to be potential endocrine disruptors, are quantifiable in MPs found in sediments or in marine waters, suggesting that leaching of additives occurs in the environment. This is clearly of great concern as microplastics exhibit a high propensity to enter all trophic levels due to their small size and ubiquity in marine environments, and given the fact that leaching may also occur in the digestive conditions of organisms upon MP ingestion.

4. Transfer and toxicity of plastics additives for marine organisms

4.1. Contamination of marine organisms by plastic additives

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PBDE have been detected in tissues of numerous marine organisms such as bivalves $(\Sigma_{13}BDE \text{ ranged from 6.6 to 440 } \mu g \text{ kg}^{-1} \text{ lipid weight)}$ (Bellas et al., 2014; Johansson et al., 2006; Ramu et al., 2007), fish (Σ₇BDE ranged from 30.6 to 281 μg kg⁻¹ lipid weight) (Peng et al., 2007) and mammals (around 100 µg kg⁻¹ wet weight (ww) in sperm whale (Physeter microcephalus) blubber) (de Boer et al., 1998), suggesting that transfer from seawater, food or plastics to organisms occurs. In their work on the contamination of the Scheldt Estuary, Verslycke et al. (2005) found PBDE in sediment and in mysid shrimp (Neomysis integer) living in this estuary ($\Sigma_{10}BDE$: 2095 to 3562 ng g⁻¹ lipid weight), and they highlighted that bioaccumulation was highest for BDE-47, -99 and -100 and lowest for BDE-209 because (i) highest brominated accumulate slower than lowest brominated congeners in the marine environment and (ii) they can be debrominated photolytically or biologically (Verslycke et al., 2005). Phthalates (DMP, DEP, DiBP, DnBP, BBP, DEHP, DnOP, DnNP) were found in a wide range of organisms, including 18 different species ranging from primary producers (plankton and macroalgae) to picked dogfish (Squalus acanthias), but no biomagnification of the studied phthalates was observed through the food web (Mackintosh et al., 2004). Recently, Cheng et al., (2013) also detected phthalates (DMP, DEP, dipropyl phthalate (DPRP – PubChem ID: 8559), DiBP, DnBP, 2-Methoxyethyl phthalate (DMEP – PubChem ID: 8344), DHP, BBP, DEHP, DOHP, DnOP, DNP+DiDP) in fish at concentrations ranging from 0.2 to 1.23 µg g⁻¹ ww (Cheng et al., 2013). NP has been detected in many organisms commonly consumed as seafood products including oysters (Crassostrea gigas) (Cheng et al., 2006), mussels (Perna perna) (Isobe et al., 2007) and fishes (Ferrara et al., 2008). For instance, Basheer et al. (2004) found NP and BPA in multiple fresh seafood products, including prawns (*Penaeus monodon*),

crabs (*Portunus pelagicus*), blood cockles (*Anadara granosa*), white clams (*Meretrix meretrix*), squid (*Loligo* sp.) and fish (*Decapterus russelli*), from a supermarket in Singapore.

Overall, these results suggest that contamination of marine organisms by plastic additives may occur *via* natural pathways (*i.e.* waterborne or foodborne exposure) or *via* ingestion of plastic debris including MP.

4.2. Plastics additive transfer to marine organisms

4.2.1. Evidence from laboratory experiments

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To investigate the potential leaching of additives from MP in environments characterized by different conditions (pH, temperature, salinity, etc), several laboratory studies have been conducted over the last years. First, the influence of gut surfactant was tested on the desorption of adsorbed chemicals, including perfluorooctanoic acid (PFOA – Pubchem ID: 9554) and DEHP, from PVC and PE in a study undertaken by Bakir et al. (2014). Desorption was higher in gut surfactant at 38°C (i.e. warm blooded animals) than in gut surfactant at 18°C (i.e. cold blooded animals) and in seawater at 18°C for DEHP. PFOA exhibited low affinity for PVC or PE regardless of the surfactant (Bakir et al., 2014). The same authors suggested that the passage of plastic through the gut could enhance desorption of pollutants and act as a transfer route for accumulation of these pollutants. However, in a more recent study, Bakir et al. (2016) demonstrated, using a one-compartment model, that MP do not provide an additional pathway for the transfer of adsorbed chemicals, including DEHP and PFOA, from seawater to marine organisms even if MP transits through the gut. Some laboratory studies have used MP in the presence of additives to determine if these chemicals can transfer to organisms. For instance, Chua et al. (2014) exposed the marine amphipod Allorchestes compressa to PBDE in the presence or absence of microbeads with PBDEs adsorbed on microbeads. Both microbead ingestion and PBDE transfer via the microbeads were demonstrated at the end of the exposure.

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However, concentrations of PBDEs were lower in amphipods exposed to PBDE adsorbed on microbeads than in amphipods exposed to PBDEs without microbeads (Chua et al., 2014) suggesting that transfer of PBDE adsorbed on MP can occur but at a lesser extent than the transfer via water. Similarly, Wardrop et al. (2016) exposed rainbow fish (Melanotaenia fluviatilis) to microbeads spiked with PBDEs (BDE-28, -47,-100, -99, -153, -154, -183 and -209) and compared them to control fish and fish exposed to microbeads alone. Here, PBDEs were analyzed in fish tissues excluding the stomach, liver, gall bladder and gonads to exclude spiked microbeads from the PBDEs analyses. During exposure, fish exposed to microbeads spiked with PBDEs showed higher concentrations than the two controls, and lower brominated congeners were better transferred in fish tissues than higher brominated congeners. On the other hand control fish and fish exposed to PBDE-free microbeads showed the same low levels of PBDEs concentration in their tissues suggesting that MP do not reduce contaminant body burden as it was previously hypothesized (Koelmans et al., 2013a, b; Koelmans et al., 2016). More realistic experiments have been performed using plastics incubated or collected in natural environments. For instance Rochman et al. (2013) used low-density polyethylene (LDPE) pellets deployed in seawater for two months and showed that the LDPE pellets adsorbed chemicals from the surrounding environment. Exposure of Japanese medaka (*Oryzias latipes*) to these pellets resulted in the accumulation of significant amounts of PBDEs and was associated with liver toxicity and pathology including glycogen depletion and cell necrosis for example (Rochman et al., 2013). Bioaccumulation of PBDEs was also demonstrated in a terrestrial invertebrate, the house cricket (Acheta domesticus), as a result of PUR foam ingestion (Gaylor et al., 2012). Another laboratory study demonstrated that the transfer of nonylphenol, triclosan and PBDE-47 can occur via MP ingestion in the lugworm (Arenicola marina) with possible effects on lugworm behavior (Browne et al., 2013).

Overall, these laboratory experiments demonstrated transfer of plastic additives upon MP ingestion, sometimes in association with toxicity or behavior change.

4.2.2. Evidence from field studies

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Levels of accumulated plastic additives in the environment or organisms have often been considered as a proxy indicator of plastic exposure in the marine environment as a consequence of the release of additives from plastic debris. For instance, a study on *Puffinus tenuirostris* showed that this bird ingested plastics at sea and that these plastics transferred flame retardant additives (PBDE) including BDE-209, which is specific to plastic (Tanaka et al., 2013). In another study, the authors demonstrated that the transfer of PBDE may occur mainly by plastic ingestion through exposure by prey ingestion (Tanaka et al., 2015). In another study, Rochman et al. (2014) examined the possible relationship between plastic densities at sea and levels of chemicals in fish inhabiting those areas. The results showed that PBDEs, especially the highest brominated congeners (BDE-209), may be an indicator of plastic pollution as previously suggested (Tanaka et al., 2013). In the North Pacific Gyre, vellowtail (Seriola lalandi) were sampled to evaluate levels of plastic in the stomach and concentrations of pollutants and additives in their tissues (Gassel et al., 2013). Ten percent of the yellowtail had ingested plastics, and PBDE and nonylphenol were concomitantly found in the fish tissues (Gassel et al., 2013). Gassel et al. (2013) suggested that contamination of fish by nonylphenol and PBDE-209 could originate from the ingested plastic as mentioned above (Hirai et al., 2011; Rochman et al., 2014; Tanaka et al., 2013; Teuten et al., 2009). Other chemicals are also used as proxies for MP contamination such as DEHP (Fossi et al., 2012; Fossi et al., 2014; Fossi et al., 2016). More recently, a study showed a higher accumulation of HBCDs in mussels (Mytilus galloprovincialis) inhabiting styrofoam debris (EPS) in comparison with mussels living on other plastic debris or rocks (Jang et al., 2016). The authors also suggested that the isomeric

profiles of detected HBCDs support the transfer of this flame retardant from the styrofoam debris to mussels through ingestion of EPS particles.

Field surveys showed that MP ingestion may constitute another route of transfer of plastic additives in marine organisms, leading to the use of plastic additives tissue content (mainly BDE-209 and DEHP) as a proxy for plastic exposure or ingestion.

4.3. Toxicity of plastic additives demonstrated by leaching experiments

Evidence for plastic toxicity has been rising in the last years. While direct toxicity can occur due to the physical impacts of plastic ingestion (for a review, see Wright *et al.* (2013)), indirect toxicity may be observed in relation to the release of hazardous chemicals from plastics. As most plastic additives are not chemically but physically bound to the plastic, they can be released into the environment and become available to organisms. Recent studies demonstrated, using leaching experiments, that various plastics are toxic to a wide range of organisms (Table 9).

Table 9: list of aquatic species, plastic polymer types, exposure times and endpoints used in various leaching experiments

Species	Plastic type	Exposure time	Exposure level	Endpoints	Reference
	PC, PVC, PU, PE,				
Danhuia maana	LDPE, PMMA, PET,	24 and 48 hours	70 100 1-1	Montality	1.41
Daphnia magna	HDPE, PTFE, ABS, PP,	24 and 48 nours	$70 - 100 \text{ g L}^{-1}$	Mortality	Lithner et al. (2009)
	MDPE				
Darle	PP, HDPE, PVC, ABS,	24 1 40 1	11-4-250 - 1-1	Mantaltan	Lithner et al. (2012)
Daphnia magna	Epoxy resin	24 and 48 hours	Up to 250 g L ⁻¹	Mortality	
	PP, PVC, PS, PET,				
Nitroca sinipes	PUR, LDPE, HDPE,	96 hours	100 g L^{-1}	Mortality	Bejgarn et al. (2015)
	ABS, PLA, Unknown				
Amphibalanus	PET, HDPE, PVC,	24, 48, 72 and 96	0.1 0.5 21-1	G all	1' (2016)
amphitrite	LDPE, PP, PS, PC	hours	$0.1 - 0.5 \text{ m}^2 \text{ L}^{-1}$	Settlement	Li <i>et al</i> . (2016)
n.	Virgin (PP) and beached	40.1	250/ C 11 / / /		Gandara e Silva <i>et al</i> .
Perna perna	pellets	48 hours	25% of pellets (v/v)	Embryo development	(2016)
Pseudochromis	PE (two different	40.1		M 1'.	II 1' (2015)
fridmani	origins)	48 hours	-	Mortality	Hamlin <i>et al.</i> (2015)

ABS: Acrylonitrile butadiene styrene; PC: Polycarbonate; PE: Polyethylene; LDPE: Low-Density Polyethylene; MDPE: Medium-Density Polyethylene; HDPE: High-Density Polyethylene; PET: Polyethylene terephthalate; PLA: Poly Lactic Acid; PMMA: Polymethyl Methacrylate; PP: Polypropylene; PTFE: Polytetrafluoroethylene; PU: Polyurethane; PVC: Polyvinyl Chloride

426 Li et al. (2016) used the seven categories of recyclable plastics (High Density PE (HDPE), 427 LDPE, PP, PVC, Polycarbonate (PC), PET and PS) to quantify the impact of their leachate on 428 the survival and settlement of barnacle Amphibalanus amphitrite larvae. Leachates were 429 prepared by placing 1 x 1 cm pieces of each plastic in 20 mL of filtered seawater for 24h at 430 28°C (Li et al., 2016). Survival was significantly lowered at the highest leachate concentration 431 (0.10 and 0.50 m² of plastic material L⁻¹) for all plastics and PVC was the most toxic plastic for 432 A. Amphitrite larvae. Additionally, settlement was also inhibited with all plastics leachates (Li 433 et al., 2016). Similarly, Bejgarn et al. (2015) exposed the copepod Nitocra sinipes to the 434 leachate of weathered or non-weathered plastics. Here, leaching experiments were performed with leachates prepared with 10 g of each plastic placed in 100 mL of brackish seawater from 435 436 the Baltic rotating at 6-21 rpm for 72h in the dark (Bejgarn et al., 2015). Of the twenty-one 437 plastics tested, eight (DVD-case (PP), biodegradable bag, costume- (PVC), flyswatter packaging (PVC), computer housing (unknown), garden hose (PVC), car dashboard (unknown) 438 439 and phone cover (PUR)) demonstrated toxicity (mortality after 96h) to N. sinipes, and after 440 weathering, toxicity either increased or decreased depending on the plastics (Bejgarn et al., 441 2015). Two leaching studies were carried out on the copepod Daphnia magna (Lithner et al., 442 2012; Lithner et al., 2009), a common organism used in ecotoxicological studies. Lithner et al. 443 (2009) prepared their leachates by placing plastic pieces in deionized water to obtain a liquid 444 to solid ratio of 10 L kg⁻¹ which was horizontally shaken at 60 rpm for 24h (16h of fluorescent 445 light and 8h of darkness) at 20°C. Out of the thirty-two plastics tested only nine, including five 446 composed of PVC, showed acute toxicity (immobility after 24 and 48h; EC₅₀ ranging from 5 to 80 g plastic material L⁻¹) to D. magna and it has been suggested that the toxicity of PVC was 447 448 due to the phthalate content (Lithner et al., 2009). In a second study, Lithner et al. (2012) used 449 PP, PE, PVC, acrylonitrile-butadiene-styrene (ABS) and epoxy resin, and they prepared their 450 leachates by adding 250 g of plastic in 1 L of deionized water shaken at 90 rpm at 50°C for 3

451 days. As previously demonstrated, PVC caused acute toxicity (immobility after 24 and 48h; 452 EC₅₀ ranging from 2-235 g plastic material L⁻¹) probably in relation to its phthalate leachates, 453 however the acute toxicity observed with the epoxy resin was not attributed to a specific 454 chemical compound (Lithner et al., 2012). A more recent study evaluated the toxicity of virgin 455 and beached pellets on the embryo development of brown mussels (Perna perna) (Gandara e 456 Silva et al., 2016). Here, the authors exposed the brown mussel embryo to 2 mL of virgin (PP) 457 or beached (42% PE and 58% unknown composition) pellets in 8 mL of seawater, and toxicity 458 was assessed by determining the percentage of dead or abnormal embryos (Gandara e Silva et 459 al., 2016). The leaching experiment led to 23.5% and 100% dead or abnormal embryos for 460 virgin and beached pellets, respectively. It has been suggested that the difference in toxicity 461 was mainly due to the difference in the chemical load of the pellets used (Gandara e Silva et 462 al., 2016). Beached pellets were exposed to in situ contamination leading to adsorption of 463 pollutants and to additives already found inside the polymeric matrix. These leaching 464 experiments showed that plastic leachates and especially PVC leachates (i.e. phthalates) can 465 lead to adverse effects on organisms. However, the toxicity highlighted in these five 466 experiments was not attributed to specific chemical compounds (Bejgarn et al., 2015; Gandara 467 e Silva et al., 2016; Li et al., 2016; Lithner et al., 2012; Lithner et al., 2009). As suggested by 468 Li et al. (2016), chemical identification should be undertaken during leaching experiments with 469 a focus on plastic additives in order to identify the compound or its degradation products 470 responsible for the observed toxicity. For instance, a more recent leaching study focused on the 471 effects of nonylphenol on the coral reef fish *Pseudochromis fridmani* by exposing single fish to 472 the leachate of plastic bags made of two PE (PE1 and PE2) from different manufacturers for 473 48h (Hamlin et al., 2015). Nonylphenol leached in the water at higher concentrations for PE2 than for PE1; with respectively 41.0 ± 5.5 and $2.5 \pm 0.2 \,\mu g \, L^{-1}$, and 60% and 11% of the fish 474 475 died after 48 hours of exposure to leachates from PE2 and PE1, respectively. However, Hamlin

et al. (2015) only focused their work on nonylphenol and did not study PE1 and PE2 compositions in terms of other additive contents. This study demonstrated that exposing fish to two identical plastic polymers (PE) may result in drastically different outcomes, as the plastic additives incorporated in each plastics are dependent on the plastic manufacturer and most of the time, their exact compositions remain unknown (Hamlin et al., 2015). Studies are required to explore potential differences between plastics from different manufacturers and toxicity related to the diversity of chemicals used in the plastic industry.

Exposure experiments based on leaching processes conducted on a wide range of polymers and target organisms confirmed toxicity of plastics additives, which highlights the need for non-target screening analysis covering a broad range of chemicals in order to better identify the main compound(s) responsible for the toxicity.

4.4. Relative importance of HOC in comparison with plastic additives: case of modelling studies

The high affinity of plastic polymers for HOC has been demonstrated in numerous laboratory experiments (Bakir *et al.*, 2014; Teuten *et al.*, 2007), and an increasing number of studies have focused on the role of MP as a vector for HOC in marine organisms (Besseling *et al.*, 2013; Rochman *et al.*, 2013). However, recent studies have suggested that given (i) baseline contamination levels of seawater and marine organisms, and (ii) the low abundance of plastic particles relative to other suspended particles found in oceans (such as organic matter, plankton, detritus etc), exposure to HOC *via* plastic may be negligible compared to natural pathways (Bakir *et al.*, 2016; Beckingham and Ghosh, 2017; Koelmans *et al.*, 2013a, b; Koelmans *et al.*, 2016; Paul-Pont *et al.*, 2016). Moreover, Koelmans *et al.* (2016) suggested that MPs ingestion by marine biota does not increase their exposure to HOCs and could have a cleaning effect while concerns have arisen regarding risk due to plastic additives.

So far most modelling studies have focused their work on adsorbed HOC (Bakir *et al.*, 2016; Koelmans *et al.*, 2016). However, no model is yet available on the transport and fate of plastic additives leaching from plastic debris although (i) plastic additives can be added in very high concentration depending on the application; and (ii) transfer of plastic additives to marine organism upon plastic ingestion has been demonstrated both in laboratory experiments and in field studies. It highlights the need to include these chemicals in future modelling work in order to better clarify their potential for transfer.

5. Conclusion

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Plastic additives associated with MP have received less attention than HOC adsorbed on MPs and the present review highlighted the need for upcoming studies to better characterize plastic additives associated with microplastics found at sea as well as their potential release in environmental matrices. As PE and PP are the main plastic debris found at sea, these two polymers should be investigated alongside with PVC due to its particularly high concentration in hazardous additives. Non-target screening analysis is required to identify the broad range of plastic additives leaching from these polymers and to better identify the main compound(s) responsible for toxicity. In addition, special attention should definitely be paid to hazardous plastic additives known to be major endocrine disruptor, namely bisphenol A and phthalates. Experimental and modelling studies are required to better characterize (i) the transfer of plastic additives upon MP ingestion relative to waterborne and foodborne exposure, and (ii) the effects of plastic additives on marine organisms. Such experiments should be realized using standardized "laboratory-made" MP, in which plastic additives are well characterized and introduced in controlled amounts reflecting industrial processes. The impacts of ageing plastic under realistic conditions on the transfer of plastic additives also need to be evaluated to investigate more environmentally relevant scenarios.

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