

Health risk assessment related to pinnatoxins in French shellfish

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1	Type of paper: Full-Length Research Papers
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3	Health risk assessment related to pinnatoxins in French shellfish
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31 ABSTRACT

32 Pinnatoxins (PnTXs) are a group of emerging marine biotoxins produced by the benthic dinoflagellate 33 Vulcanodinium rugosum, currently not regulated in Europe or in any other country in the world. In 34 France, PnTXs were detected for the first time in 2011, in mussels from the Ingril lagoon (South of 35 France, Mediterranean coast). Since then, analyses carried out in mussels from this lagoon have 36 shown high concentrations of PnTXs for several months each year. PnTXs have also been detected, 37 to a lesser extent, in mussels from other Mediterranean lagoons and on the Atlantic and Corsican 38 coasts. In the French data, the main analog is PnTX G (low levels of PnTX A are also present in some 39 samples). No cases of PnTXs poisoning in humans have been reported so far in France or anywhere 40 else in the world. In mice, PnTXs induce acute neurotoxic effects, within a few minutes after oral 41 administration. Clinical signs of toxicity include decreased mobility, paralysis of the hind legs, tremors, 42 jumps and breathing difficulties leading to death by respiratory arrest at high doses. The French 43 agency for food safety (ANSES) recently conducted a review of the state of knowledge related to 44 PnTXs and V. rugosum. Based on (i) the clinical signs of toxicity in mice, (ii) the mode of action of 45 PnTXs as nicotinic acetylcholine receptor competitive antagonists and (iii) knowledge on drugs and 46 natural toxins with PnTX-related pharmacology, potential human symptoms have been extrapolated 47 and proposed. In this work, a provisional acute benchmark value for PnTX G of 0.13 µg/kg bw per day 48 has been derived from an oral acute toxicity study in mice. Based on this value and a large shellfish 49 meat portion size of 400g, a concentration lower than 23 µg PnTX G/kg shellfish meat is not expected 50 to result in adverse effects in humans. ANSES recommends taking into account PnTXs in the French 51 official monitoring program for shellfish production and identified data gaps to refine health risk 52 assessment.

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54 KEYWORDS: Pinnatoxins, shellfish, emerging marine biotoxins, risk assessment

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61 **1. Introduction**

62 Pinnatoxins (PnTXs) belong to the group of cyclic imines considered as emerging marine biotoxins 63 (Efsa, 2010a), which, to date, includes 40 compounds, without considering acyl esters produced by 64 shellfish metabolism. This group includes different families determined by their structural 65 characteristics: prorocentrolides, spiroprorocentrimine, gymnodimines (GYMs), spirolides (SPXs), pinnatoxins (PnTXs), pteriatoxins (PtTXs) and portimine. To date, 8 PnTXs (named A to H) have been 66 identified (reviewed by Molgó et al., 2017). PnTXs are soluble in solvents such as acetone, 67 68 isopropanol and methanol (Zendong et al. 2014). They are amphoteric compounds, i.e. with both 69 acidic and basic properties, which explains their relative water solubility. Due to their lipophilic 70 properties, they can be detected during the mouse bioassay used to screen for lipophilic marine 71 biotoxins.

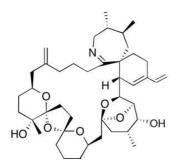
72 Vulcanodinium rugosum is the producer of PnTXs identified by Nézan and Chomerat (2011), based on 73 water samples from a Mediterranean lagoon (Ingril, France). V. rugosum also produce portimine 74 (Selwood et al., 2013; Abadie et al., 2016). This benthic dinoflagellate was a new species belonging to 75 a new genus. No other producer is reported in the literature. The identification of this dinoflagellate in 76 France originates from an atypical situation that occurred in 2006, as part of official monitoring of 77 shellfish production areas. The mouse bioassay used to screen for lipophilic marine biotoxins had 78 revealed unusual neurotoxic effects after the injection of extracts from mussels of the Ingril lagoon. In 79 fact, the symptoms observed for lipophilic toxins are gastrointestinal (diarrhea), and not neurotoxic. 80 Such neurotoxicity could not be related to the presence of regulated marine biotoxins (Amnesic, Paralytic, Diarrhetic toxins) screened for by chemical analysis. In addition, observations of water 81 82 samples did not conclude to the presence of any microalgae species known to produce neurotoxins. 83 Vulcanodinium rugosum has also been reported in New Zealand (Rhodes et al., 2010), Australia 84 (Rhodes et al., 2011; Munday et al., 2012), Japan (Smith et al., 2011), China Sea (Zeng et al., 2012), 85 Mexico (Hernandez-Becerril et al., 2013), Arabian Gulf (Al Muftah et al., 2016) and Cuba (Moreira-86 Gonzalez et al., 2018).

PnTXs in shellfish are mainly analysed by physico-chemical methods using liquid chromatography coupled with tandem mass spectrometry (LC-MS/MS). Biological (mouse bioassay) or biochemical (functional tests) methods can also be used. None of the methods has yet undergone inter-laboratory validation or standardisation. In addition, there are very few standards (reference substances)
available for PnTXs, which limits and complicates their detection and quantification. Indeed, to date,
only PnTX A and G are marketed as calibration solutions and only PnTX G (Figure 1) has been
certified.

Since the first identification of *V. rugosum* in France, high concentrations of PnTXs have been measured in mussels from the Ingril lagoon for several months each year. In 2012, the French Research Institute for Exploitation of the Sea (Ifremer) reported that PnTXs concentrations varied greatly depending on the years (2010, 2011 and 2012), with a maximum of 1244 µg of PnTX G per kg of shellfish (wet weight) in 2010 (Ifremer, 2012). However, *V. rugosum* is rarely detected in water samples collected for the phytoplankton surveillance (in the water column), which can be explained by the benthic nature of this dinoflagellate.

101 The aims of this work were 1) to review the toxicity data of PnTXs to derive an acute oral health-based 102 guidance value, 2) estimate exposure of shellfish consumers to PnTXs, 3) identify if there might be a 103 public health concern regarding the levels of contamination reported in certain French shellfish 104 production areas, and 4) provide recommendations for the monitoring PnTXs in the marine 105 environment.

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108 Figure 1: Structure of PnTX G (modified from Araoz et al., 2011).

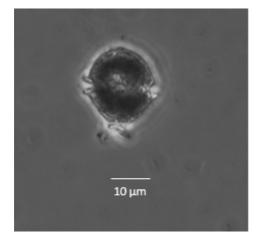
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110 2. Vulcanodinium rugosum

As almost all dinoflagellates, *V. rugosum* (Figure 2) is assumed to have two distinct life phases: a vegetative propagation phase (asexual reproduction) and a sexual phase (Zeng et al., 2012). The asexual phase probably corresponds to the pelagic phase, when the cells are present in the water 114 column. The sexual reproduction corresponds to a benthic phase, which could result in the formation 115 of a planozygote that produces a resistance cyst. The pelagic and benthic phases are closely related, 116 but the environmental and/or physiological factors controlling the transition from one phase to the 117 other are not yet known.

118 The formation of resistance cysts, which can occur at the end of sexual reproduction, is a crucial step 119 for survival in adverse environmental conditions due to chromosomal mixing, which increases intraspecies genetic diversity. Cysts are also one of the ways in which the species spreads, through the 120 121 transfer of sediments or shellfish from one area to another (cysts are present in the gastrointestinal 122 tract and in intervalvular liquid), as well as via ship ballast water (Garrett et al., 2014). Therefore, 123 identification of the resistance forms and knowledge of their distribution area are important for 124 preventing expansion and contamination. Similarly, determination of the key factors for growth and 125 toxin production by V. rugosum is essential for better understanding the risk associated with this new 126 species.

127 Temperature is the most important factor for the growth of *V. rugosum*. The data suggest that *V. rugosum* is a thermophilic species, consistent with its development in the Ingril lagoon from June to 129 September and the highest concentrations of PnTX G found in mussels during this period (Abadie et 130 al., 2016).



- 131
- 132 Figure 2: Micrograph of a cell of *Vulcanodinium rugosum*
- 133
- 134 **3. Hazard characterisation**
- 135 **3.1 Acute** *in vivo* toxicity with purified PnTXs

136 *In vivo* acute toxicity studies with purified toxins were reviewed using an analysis grid and assessed 137 for quality using ToxRTool (Toxicological data Reliability assessment Tool, https://eurl-138 ecvam.jrc.ec.europa.eu/about-ecvam/archivepublications/toxrtool), a tool that ranks the studies 139 according to the Klimisch rating (Klimish et al., 1997).

Data on the acute toxicity of purified PnTXs are very limited. Indeed, the available studies were carried out in only one species (mice), one sex (females, which are considered to be more sensitive than males by OECD guidelines) and with very few animals tested per dose (1 to 8 mice, when the number is specified). The results are presented in table 1.

Orally, only three studies are available and focused on PnTX E, F, G and H (Munday *et al.*, 2012;
Selwood *et al.*, 2014; ANSES-University of Trieste-CNRS report, 2014 also published in Sosa *et al.*,
2020). We did not find any information on the toxicity by the oral route for PtTXs (metabolites of certain
PnTXs) and portimine (a toxin also produced by *V. rugosum*). Regarding the intraperitoneal (IP) route,
four studies have been published on PnTX E, F, G and H and portimine (Munday *et al.*, 2012;
Selwood *et al.*, 2010, 2013, 2014). We identified additional unpublished information regarding PnTX A
(personal communication from J. Molgó), but did not find any robust information regarding PtTXs.

151 Despite the limited number of studies for each PnTX and the low number of mice tested per dose, a 152 consistent set of information enables outlining the main characteristics of the acute toxicity of this toxin 153 group. Firstly, the toxicity of the PnTXs is rapid, with symptoms appearing within minutes of 154 administration (whether oral or IP). This fact was already known, since PnTXs (like other cyclic imines) 155 belong to the fast-acting toxins group (Efsa, 2010a). The second point relies on neurotoxic symptoms, 156 guickly leading to the mouse's death by respiratory arrest. Clinical signs of toxicity, regardless of the 157 route of administration and the PnTX analogue, include decreased mobility (sometimes preceded by 158 an initial phase of hyperactivity immediately following administration), paralysis of the hind legs and 159 breathing difficulties (Munday et al., 2012), with tremors and jumps also being reported (ANSES-160 University of Trieste-CNRS Report, 2014; Sosa et al., 2020). In the study by Munday et al. (2012), the 161 authors reported that at sublethal doses (without specifying which ones), some mice recovered 162 completely after exhibiting symptoms.

LD50 values vary according to the PnTX analogue and route of administration (Table 1). Orally, LD50
values range from 25 to 2,800 μg/kg bw (for PnTX F and PnTX E respectively). The analogues tested
can be ranked as follows in decreasing order of toxicity: PnTX F > PnTX G ~ PnTX H >> PnTX E. By

- 166 IP route, LD50 range from 13 to 115 μ g/kg bw (for PnTX F and PnTX A respectively) and the 167 analogues can be ranked as follows: PnTX F > PnTX G > PnTX E > PnTX H > PnTX A.
- 168 LD99 values for some PnTXs and PtTXs have been reported in the literature, but these data were 169 considered not sufficiently reliable due to a lack of information on the protocol used (Uemura *et al.*,
- 170 1995; Chou *et al.*, 1996; McCauley *et al.*, 1998; Takada *et al.*, 2001).
- Portimine has lower acute IP toxicity than that of PnTXs, with an estimated LD50 of 1,570 µg/kg bw.
 No effect was observed at 500 and 700 µg/kg bw (Selwood *et al.*, 2013). The authors indicated that
- 173 the signs of toxicity prior to death in mice appeared less rapidly after IP administration compared to
- 174 PnTXs (without however mentioning which signs of toxicity were observed). Toxicity of portimine by
- 175 oral administration is unknown.
- 176

177 Table 1: Acute *in vivo* toxicity in mice of PnTXs and portimine

178 **1a. Oral administration**

Toxin (purity)	Route of administration and number of mice	LD50 (µg/kg bw)	MTD (μg/kg bw)	References	Study quality
PnTX E*	Gavage Number of mice not specified (OECD GL 425) Fed (non-fasted) mice	2800 Cl95: 2380-3000	600	Munday <i>et</i> <i>al.,</i> 2012	ToxRTool: 13 Klimisch: 2
PnTX F*	Gavage Number of mice not specified (OECD GL 425) Fed (non-fasted) mice	25 Cl95: 19.1-35.1	9.9	Munday <i>et</i> <i>al.</i> , 2012	ToxRTool: 13 Klimisch: 2
PnTX F*	16h fasted mice	29.9 Cl95: 25-32	Not determined		
	In cream cheese Number of animals not specified (OECD GL 425) Fed (non-fasted) mice	50 Cl95: 39.4-62.8	16.0	Munday <i>et</i> <i>al.,</i> 2012	ToxRTool: 13 Klimisch: 2
	In peanut butter Number of mice not specified (OECD GL 425) Fed (non-fasted) mice	50 Cl95: 37.9-71.5	Not determined		
	In dry mouse food Number of mice not	50 Cl95: 37.9-71.5	Not determined		

Toxin (purity)	Route of administration and number of mice	LD50 (µg/kg bw)	MTD (μg/kg bw)	References	Study quality
	specified (OECD GL 425) 16h fasted mice				
	In cream cheese, 16h fasted mice	77 Cl95: not calculated	Not determined		
	In peanut butter, 16h fasted mice	50 Cl95: 39.4-62.8	Not determined		
PnTX G*	Gavage Number of mice not specified (OECD GL 425) Fed (non-fasted) mice	150 Cl95: 105-100	75	Munday et al., 2012	ToxRTool: 13 Klimisch: 2
PnTX G (declared purity 100%)	Gavage Groups of 3 to 8 mice (8 for the control group; 3 for the doses 8, 20, 50, 120 µg/kg bw; 5 for the doses 220, 300, 370, 400 µg/kg bw) Mice fasted for 3 hours before administration	208 Cl95: 155-281	120	ANSES- University of Trieste-CNRS report, 2014; Sosa <i>et al.</i> , 2020	ToxRTool: 17 Klimisch: 2
PnTX G*	In cream cheese Number of mice not specified (OECD GL 425) Fed (non-fasted) mice	400 Cl95: 380-470	153	Munday <i>et</i> <i>al.,</i> 2012	ToxRTool: 13 Klimisch: 2
PnTX H*	Gavage Number of animals not specified Fasting not specified	163 Cl95: 139-175		Selwood <i>et</i> <i>al.</i> , 2014	ToxRTool: 4 This low score is due to the fact that a lot of details are missing in the article, but this team has already described the protocol for other PnTXs

* Purity verified by NMR according to the authors but percentage not mentioned in the publication

Cl95: 95% confidence interval MTD (Maximum Tolerated Dose): dose at which no mortality or clinical signs are observed

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1b. Intraperitoneal (IP) administration 183

Toxin (purity)	Route of administration and number of mice	LD50 (µg/kg bw)	MTD (μg/kg bw)	References	Study quality
Synthetic PnTX A (purity > 97%)	IP n=18 mice, 9 doses tested (1 to 3 mice/dose)	114.8		J. Molgó (personal communication)	ToxRTool: 16

Toxin (purity)	Route of administration and number of mice	LD50 (µg/kg bw)	MTD (µg/kg bw)	References	Study quality
	Fed (non-fasted) mice				
PnTX E*	IP Number of animals not specified (OECD GL 425)	57 (fed) Cl95: 39.7-75.3 48 (16h fasted) Cl95: 33.5-63.5	22 Not determined	Munday <i>et al.,</i> 2012	ToxRTool: 13
	IP (OECD GL 425) n=2 at 36 μg/kg, n=3 at 45 μg/kg, n=1 at 54 μg/kg, n=1 at 60 μg/kg.	45 (fed) Cl95: 32-58		Selwood <i>et al.</i> , 2010	ToxRTool: 15
PnTX F*	IP Number of animals not specified (OECD GL 425)	12.7 (fed) CI95: 9.5-14.6 14.9 (16h fasted) CI95: 12.6-15.8	3.2 Not determined	Munday <i>et al.,</i> 2012	ToxRTool: 13
	IP (OECD GL 425) n=1 at 10.1 μg/kg, n=1 at 12.7 μg/kg, n=3 at 16.0 μg/kg, n=2 at 20.1 μg/kg.	16 (fed) Cl95: 12-23		Selwood <i>et al.,</i> 2010	ToxRTool: 15
PnTX G*	IP Number of animals not specified (OECD GL 425)	48 (fed) Cl95: 36.3-68.1 42.7 (16h fasted) Cl95: 40-50	18.8 Not determined	Munday <i>et al.,</i> 2012	ToxRTool: 13
PnTX G*	IP (OECD GL 425) n=2 at 40 μg/kg, n=3 at 50 μg/kg, n=1 at 60 μg/kg.	50 (fed) Cl95: 35-66		Selwood <i>et al.</i> , 2010	ToxRTool: 15
Synthetic PnTX G (purity > 97%)	IP Number of animals not specified	65.8		J. Molgó (personal communication)	ToxRTool: 16
PnTX H*	IP Number of animals not specified Fasting not specified	67 CI95: 63-79		Selwood <i>et al.,</i> 2014	ToxRTool: 4 This low score is due to the fact that a lot of details are missing in the article, but this team has already described the protocol for other PnTXs
Portimine*	IP Number of animals not	1570 Cl95: 1269-3080	No effect at 500 and 700	Selwood <i>et al.,</i> 2013	ToxRTool: 4 This low score is due to the fact

Toxin (purity)	Route of administration and number of mice	LD50 (µg/kg bw)	MTD (μg/kg bw)	References	Study quality
	specified Fasting not specified				that a lot of details are missing in the article, but this team has already described the protocol for other PnTXs

184 * Purity verified by NMR according to the authors but percentage not mentioned in the publication

185 CI95: 95% confidence interval

186 MTD (Maximum Tolerated Dose): dose at which no mortality or clinical signs are observed

187

188 **3.2 Subchronic and chronic toxicity**

189 No repeated dose toxicity studies were identified in the literature.

190

191 **3.3 Mode of action and extrapolation to humans**

192 We reviewed the data available on the mode of action of PnTXs (Delcourt et al., 2019). PnTXs are

193 high-affinity competitive antagonists of nicotinic acetylcholine receptors (nAChRs) (Araoz et al., 2011;

194 Hellyer et al., 2015). Their lethal effects in mice studies are consistent with the inhibition of muscle

195 nAChRs, inducing respiratory distress and paralysis (Delcourt et al., 2019). PnTX E, F , G and A block

196 nerve stimulation-induced skeletal muscle contraction, but do not alter muscle contraction caused by

197 direct stimulation of isolated adult rodent muscles (Hellyer *et al.*, 2013; Benoit et al., 2019).

198 The clinical effects observed in humans after exposure to compounds (drugs, natural toxins) whose

199 pharmacology is comparable to that of PnTXs have been described (Delcourt et al., 2019). Possible

200 symptoms in humans due to PnTXs could include muscle weakness (myasthenic-like syndrome),

201 dyspnea, anticholinergic syndrome, dysautonomia, pyramidal syndrome, and seizures.

202

203 3.4 Development of a health value

The methodology set in ANSES's guide for the development of toxicological reference values (2017)was followed.

206

207 3.4.1 Choice of the key study

Two acute oral toxicity studies in mice with purified PnTX G were identified (Munday et al., 2012; ANSES-University of Trieste-CNRS report, 2014 also published in Sosa *et al.*, 2020). The key study selected was the one carried out by the Department of Life Sciences at the University of Trieste with funding from the DGAL and DGS (ANSES-University of Trieste-CNRS report, 2014; Sosa et al., 2020), which obtained the highest score (17 out of 21 with ToxRTool, corresponding to a Klimisch score of 2 reliable with restriction).

In this study, groups of three to five 4 weeks-old female SD-1 mice received a single administration of purified PnTX G by gavage at a dose of 8, 20, 50, 120, 220, 300, 370 or 450 µg/kg bw. A control group of eight mice was included. Mice were fasted for 3 hours before gavage; food was given back again *ad libitum* 2 hours after administration during the 24-hour observation period. Lethality, clinical signs of toxicity, histological analysis of several organs and blood biochemistry were recorded.

The results regarding lethality and the observed symptoms are shown in Table 2. Administration of PnTX G resulted in mouse lethality from 220 μ g PnTX G/kg bw (3/5 mice, in 22 min). No lethality was observed at the doses of 8, 20, 50 and 120 μ g/kg bw. All mice (5/5) died at the dose of 370 μ g/kg bw (survival time between 13 and 18 min). The LD50 for PnTX G was calculated at 208 μ g/kg bw (95% confidence interval = 155-281 μ g/kg bw).

Before death, the main symptoms were neurotoxic, i.e. prostration, tremors, jumps, abdominal breathing, hypothermia, hind leg paralysis and cyanosis. No macroscopic organ changes were observed during necropsy of the mice treated with PnTX G. Histological analysis of major organs and tissues revealed only minor changes in the small intestine of mice given PnTX G at doses equal to or greater than 300 µg/kg bw (moderate mucosal degeneration, villous atrophy). No differences in biochemical blood parameters were found between treated and control mice (Sosa *et al.*, 2020).

230

Table 2: Lethality and clinical signs of toxicity in mice after single administration by gavage of

232 purified PnTX G (ANSES-University of Trieste-CNRS Report, 2014; Sosa et al., 2020)

Purified PnTX G dose	Lethality	Survival time (h:min)	Clinical signs of toxicity
Control	0/8	-	-
8 μg/kg bw	0/3	-	-
20 µg/kg bw	0/3	-	-
50 μg/kg bw	0/3	-	-
120 µg/kg bw	0/3	-	-
220 µg/kg bw	3/5	00:20; 00:22; 00:22	prostration, tremors, jumps, abdominal

			breathing, hypothermia, hind leg paralysis, cyanosis
300 µg/kg bw	4/5	00:12; 00:13; 00:17; 00:23	prostration, tremors, jumps, abdominal breathing, hypothermia, hind leg paralysis, cyanosis
370 μg/kg bw	5/5	00:13; 00:15; 00:16; 00:17; 00:18	prostration, tremors, jumps, abdominal breathing, hypothermia, hind leg paralysis, cyanosis
450 μg/kg bw	5/5	00:12; 00:12; 00:15; 00:16; 00:29	prostration, tremors, jumps, abdominal breathing, hypothermia, hind leg paralysis, cyanosis

234 3.4.2 Choice of the critical dose

Based on the data from the key study, two approaches were explored:

First approach: the 120 μg PnTX G/kg bw dose was selected as the maximum tolerated dose
 (MTD), i.e. the dose at which no effect is observed in the studied parameters over a 24-hour
 period after treatment.

- Second approach: calculation of a benchmark dose¹ from a response level of 10% mortality
 (PROAST Web software, RIVM, v. 65.2). The model that best fitted the experimental data
 according to the Akaike Information Criterion (AIC) was the probit model, for which the BMDL_{95%}
 was 69.1 µg PnTX G/kg bw of mouse.
- 243 It is unusual to use data on mouse mortality from single oral administration to derive a health reference
 244 value. This is justified by the fact that the mouse mortality occurs rapidly and there is a very small
 245 difference between a dose with no signs of toxicity and a lethal dose.
- A response level of 10% (for lethality) is considered high but was exceptionally selected in this expert
- appraisal because the results obtained with a response level of 1% were regarded as too uncertain
- 248 due to the small number of mice tested in the study.
- The results obtained in the other acute oral toxicity study of purified PnTX G with non-fasted mice (Munday *et al.*, 2012) were consistent with those of the key study selected conducted by the Department of Life Sciences at the University of Trieste:
- 252 i. The LD50 was 150 μg PnTX G/kg bw (95% CI = 105-199 μg/kg bw);

¹ The benchmark dose is a dose producing a measurable effect corresponding to a given response level compared to a control group. The lower limit of its 95% or 90% confidence interval ($BMDL_{95\%}$ or $BMDL_{90\%}$) is most often used. This approach is based on modelling of the experimental data taking into account the entire dose-response curve (ANSES 2017a).

- 253 ii. The dose tested without lethality and with no apparent sign of neurotoxicity was 75 µg PnTX G/kg
- 254

bw.

- 255 256 3.4.3 Choice of uncertainty factors 257 Uncertainty factors were identified for both approaches explored when establishing the critical dose: 258 First approach: the dose of 120 µg PnTX G/kg bw was selected as the maximum tolerated dose 259 (MTD). The associated overall uncertainty factor was 900: 260 \circ inter-species uncertainty factor: UF_A = 10 261 \circ inter-individual uncertainty factor: UF_H = 10 262 \circ other factors: UF_D = 3 (insufficient data) and 3 (to take into account the severity and 263 pattern of the dose-response curve) Second approach: the BMDL of 69.1 µg PnTX G/kg bw was chosen as the point of departure. 264 265 The associated overall uncertainty factor was 525: 266 o allometric adjustment factor: 7 for the mouse 267 \circ UF_A = 2.5 for the toxicodynamic component о UF_H = 10 268 \circ UF_D = 3 (insufficient data). 269 270 271 3.4.4 Proposed health-based guidance value 272 Due to insufficient data for the hazard characterisation of PnTX G, the guidance value derived is 273 considered as a provisional acute benchmark value. The provisional acute benchmark value is 0.13 µg PnTX G/kg bw, (Table 3) regardless of the 274 approach used² (MTD or BMDL). 275 276
- 277 Table 3: Table summarising development of the provisional acute benchmark value for PnTX G

Critical effect Key study	Critical dose for the mouse	UF	Provisional acute benchmark value for humans	Confidence level
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² MTD approach: (120 μg PnTX G/kg bw) / (10 x 10 x 3 x 3) = 120/900 = 0.1333 μg PnTX G/kg bw BMDL approach: 69.1 μg PnTX G/kg bw) / (7 x 2.5 x 10 x 3) = 69.1/525 = 0.1316 μg PnTX G/kg bw

First approach	<i>Critical effect:</i> absence of mortality and symptoms in mice 24 hours after a single oral administration by gavage of purified PnTX G	Maximum tolerated dose of 120 μg PnTX G/kg bw	Total factor: 900 $UF_A = 10$ $UF_H = 10$ $UF_D = 9$ (3 for insufficient data and 3 for the severity and pattern of the dose-response curve).	0.13 μg PnTX G/kg bw	Moderate
Second approach	ANSES-University of Trieste-CNRS report, 2014; Sosa et al., 2020	BMDL of 69.1 μg PnTX G/kg bw	Total factor: 525 Allometric adjustment: 7 for the mouse $UF_A = 2.5$ $UF_H = 10$ $UF_D = 3$ for insufficient data	0.13 μg PnTX G/kg bw	Moderate

279 **3.4.5 Proposed maximum tolerable concentration of PnTX G in shellfish**

Based on the provisional acute benchmark value of 0.13 µg PnTX G/kg bw, a default serving size of 400 g of shellfish (EFSA 2010b) and a default body weight of 70 kg, the concentration not to be exceeded in shellfish would be **23 µg PnTX G/kg of total meat**.

283

284 3.4.6 Confidence level

An overall confidence level **moderate** was assigned to the provisional acute benchmark value based on the following criteria:

287 - Level of confidence in the type and quality of the body of data: Low

The literature review revealed that there were few data on the acute oral toxicity of purified PnTXs and none on repeated administration. In fact, only two acute oral toxicity studies conducted on a single species (mice) and sex (females, which are considered to be more sensitive than males) have been published. The lack of data in another rodent species (rats) and of knowledge on the toxicokinetics and toxicodynamics of PnTXs is underlined.

293 - Level of confidence in the choice of the critical effect and the mode of action: Moderate

The key study is an acute toxicity study with a single oral administration. Its objective was to study mouse mortality and define a median lethal dose (LD50). Signs of neurological toxicity were also observed and are consistent with the known mode of action of PnTXs.

We stress that it is unusual to use data on mouse mortality from single oral administration to derive a health reference value. This is justified by the fact that the mouse mortality occurs rapidly and there is a very small difference between a dose with no signs of toxicity and a lethal dose.

300 - Level of confidence in the choice of the key study: Moderate/High

301 The key study was analysed using ToxRTool and obtained a total score of 17 (out of 21), 302 corresponding to a score of 2 using the Klimisch method (reliable study with restriction).

303 - Level of confidence in the choice of the critical dose: High

Two approaches were explored for selecting the critical dose from the key study data. The first approach retained a maximum tolerated dose (MTD), at which there is no mortality or signs of toxicity in mice during the 24-hour observation period (which is considered sufficient because PnTXs are fastacting toxins, with neurotoxic signs in mice occurring within 30 minutes of administration). The second approach modelled the dose-response relationship (lethality) to calculate a BMDL.

This overall confidence level may be reassessed when new acute oral toxicity data become available
for PnTX G or other toxins produced by *V. rugosum*.

311

312 4. Occurrence of PnTXs in shellfish

313 4.1 Ingril lagoon

As soon as PnTX G was identified in France (2011), a retrospective analysis of the samples from 2009 to 2012 was conducted by lfremer, which revealed the kinetics of shellfish contamination as a function of time. The maximum annual concentrations were 261, 1244, 568 and 652 µg/kg of total mussel meat for 2009, 2010, 2011 and 2012 respectively (Table 4). Concentrations in mussels were higher than those observed in clams, when shellfish were sampled simultaneously (Table 5).

To track this phenomenon and maintain monitoring for PnTXs, sampling of Ingril mussels was carried out monthly between 2013 and 2017 by Ifremer. The results show that PnTX G peaks were observed 321 between June and September, but the maximum values varied according to the year (887 in 2013,

322 918 in 2014, 1143 in 2015, 600 in 2016 and 640 in 2017, expressed in µg/kg of total meat).

323

- Table 4: Data on concentrations of PnTX G in mussels from Ingril lagoon from 2010 to 2017
- 325 (µg/kg total meat)

Year	2010	2011	2012	2013	2014	2015	2016	2017
Number of samples	30	32	24	12	27	2	13	10
Min	45.4	36.6	11.0	49.5	22.6	572	44.0	52.9
Max	1244	568	652	887	918	1143	600	640
Mean	223	227	209	293	247	858	278	294

326

327 Table 5: Data on concentrations of PnTX G in mussels and clams from Ingril lagoon from 2010

328 to 2012 (µg/kg total meat)

	Mussels	Clams
Number of samples	86	20
Min	11.0	13.6
Max	1244	95.3
Mean	217	28.1

329

330 4.2 Other Mediterranean lagoons

In addition to the monitoring carried out in the Ingril lagoon, screening for PnTX G in wild mussel samples took place the same week in four other Mediterranean lagoons in 2013 (Table 6). Maximum levels of PnTX G increased in the following order: Parc Leucate (11 μ g/kg of total meat) > Thau (15 μ g/kg) > Le Prevost (54 μ g/kg) > Vic (89 μ g/kg) > Ingril (887 μ g/kg).

335

336 Table 6: Data on concentrations of PnTX G in mussels from several Mediterranean lagoons in

337 2013 (µg/kg total meat)

	Parc Leucate	Thau	Le Prévost	Vic	Ingril
Number of samples	12	11	12	11	12
Min	0.68	1.4	11.8	12.3	49.5

Max	10.7	14.7	53.9	89.0	887
Mean	3.4	10.0	26.2	48.4	293

339 4.3 Atlantic and Corsican coasts

In France, the EMERGTOX scheme for monitoring the emergence of marine biotoxins in shellfish was set up by the DGAL to complement national surveillance schemes for regulated toxins (REPHYTOX, the DGAL's surveillance plan). Its purpose is to bring to light any possible hazard associated with the presence in shellfish of known regulated and unregulated lipophilic toxins, either identified in France or that could be introduced into France via ballast water or commercial trade between countries.

345 Since 2018, this scheme enables the targeted analysis by chemical methods of PnTXs in mussels and 346 oysters from eleven areas located all along the French metropolitain coasts. During this first year of 347 monitoring, three of these areas (Ingril, Le Scoré in Brittany and the Diana lagoon in Corsica) were 348 affected by the presence of PnTXs, mainly PnTX G and to a lesser degree PnTX A (only in Ingril). Of 349 the three affected areas, the Ingril lagoon remains the most heavily contaminated area (with concentrations detected every month, varying from 40 to 2614 µg PnTX G/kg of digestive gland). The 350 351 levels found at Le Scoré and Diana were low (maximum around 10 µg PnTX G/kg of digestive gland). 352 The concentrations of PnTX A detected at Ingril ranged from 6 to 32 µg/kg of digestive gland.

We believe it is important to point out that the presence of PnTXs in France is not limited to Mediterranean lagoons. Given that global warming is making ecophysiological conditions more favourable to the development of *V. rugosum,* vigilance should be maintained along the Atlantic coast, since the detection of PnTXs in mussels from Brittany points out the presence of the dinoflagellate producer also in these waters.

358

359 4.4 Outside France

The literature review concluded that very limited data on PnTX levels in shellfish from Northern and Southern Europe, Canada and New Zealand have been published. Among the eight studies reviewed (MacKenzie et al., 2011; Rundberget, et al., 2011; McCarron et al., 2012; McNabb et al., 2012; Garcia-Altares et al., 2014; Rambla Alegre et al., 2018; Lamas et al., 2019; Otero et al., 2019), the highest reported levels of PnTX G in mussels were 115 µg/kg in Norway (Rundberget *et al.*, 2011), 83 µg/kg in Canada (McCarron *et al.*, 2012) and 59 µg/kg in Spain (Garcia-Altares *et al.*, 2014). These levels are lower than the annual maximum levels reported in France in the Ingril lagoon (between 261 and 1244 µg/kg, depending on the year). It should be noted that the study by Rambla-Alegre *et al.*(2018) reported the persistence of PnTX G in samples of canned mussels (up to 12 µg/kg).

Lastly, ANSES sent a questionnaire to the European Food Safety Authority (EFSA) network of focal points on 15 February 2018, to which 16 Member States replied. At the time of the request, none of the responding health agencies had conducted a health risk assessment for PnTXs. Some respondents provided contamination data in shellfish from their country. Most of the data were already published in a scientific journal and had been identified in our literature search. Some unpolished data were also provided but without information about the performance of the analytical method used.

In conclusion, the concentrations of PnTX G measured in mussels from Ingril are the highest reportedin the world to date.

377

378 5 Dietary exposure

379

380 **5.1. Food consumption data**

A consumption survey on seafood products in France (CONSOMER) was conducted in 2016-2017 as part of a research agreement between ANSES and CREDOC (2015-CRD-25). The aim of the survey was to assess seafood consumption by an adult population (over 18 years of age) living in coastal areas and with access to local sources of supply. The database includes answers from 2481 adults. Only data for the Mediterranean population were used in the present work, including 821 adults (Table 7).

387

Table 7: Consumption data of mussels and clams from CONSOMER survey (Mediterranean
 population only), daily portion size.

	Number of consumers	Mean amount in g/day	P95 in g/day
Mussels	582	91.2	200
Clams	140	19.8	40

390

391 **5.2 Contamination data**

392 Data on concentrations of PnTX G in mussels and clams from several Mediterranean lagoons from
393 2010 to 2017 were provided to ANSES by Ifremer (Table 8). Three scenarios were selected,
394 depending on the site where the shellfish were sampled:

395 - "All lagoons" scenario: contamination data of all mussels on the one hand and all clams on the

396 other (from all the lagoons studied);

397 - "Ingril" scenario: contamination data ONLY from the Ingril lagoon, for mussels on the one hand and

398 clams on the other;

- 399 "Except Ingril" scenario: contamination data from all lagoons (Vic, le Prévost, Parc Leucate and
- 400 Thau) except "Ingril", for mussels only (no clams sampled in other lagoons).
- 401
- 402
- 403
- 404
- 405

406 Table 8: Concentrations of PnTX G in mussels and clams from several Mediterranean lagoons

407 from 2010 to 2017

	Number of samples	Concentration of PnTX G (µg/kg total meat)			
	of mussels	Mean	P95		
All lagoons	196	196	633		
Ingril	150	249	712		
Except Ingril*	46	21.7	60.3		
	Number of samples	Concentration of PnTX G (µg/kg total meat)			
	of clams	Mean	P95		
All lagoons	20	28.1	57		
Ingril	20	28.1	57		
Except Ingril	0	No data	No data		

408 * Number of samples of mussels in other lagoons than Ingril are described in Table 6.

409

413

410 **5.3. Method to calculate exposure**

411 From the individual consumption data and the contamination data, exposure was calculated according

412 to the following equation:

$$E_i = \sum_{k=1}^n \frac{C_{i,k} \times L_k}{BW_i}$$

- 414 where:
- 415 *E_i* is the total daily exposure of individual i associated with the consumption of food k (µg/kg of body
- 416 weight/day);
- 417 *C*_{*i,k*} is the consumption of food k by individual i (kg/d) (serving size for acute exposure);

- 418 L_k is the PnTX G contamination of food k (μ g/kg of fresh weight) (95th percentile for acute exposure);
- 419 BW_i is the body weight of individual i (kg).

421 6 Risk characterisation

422 6.1 Results

A comparison of acute exposure estimates was made with the provisional acute benchmark value of
0.13 μg PnTX G/kg of body weight. A percentage of the population exceeding this value was
calculated, firstly in the total population and secondly for consumers of mussels and clams only (Table
9).

- 427
- 428

429 Table 9: Acute exposure associated with consumption of mussels and clams (CONSOMER,

430 Mediterranean area) for adults (in µg PnTX G/kg bw)

	Total population (Mediterranean area)			Consumers only (Mediterranean area)			Provisional acute benchmark value (PABV) exceeded		
	n Ave_pop P95_pop			n_cons	Ave_cons	P95_cons	% > PABV	% > PABV_cons	
All lagoons	821	0.590	1.572	591	0.816	1.733	71	100	
Ingril	ngril 821 0.663 1.767		591 0.917 1.946		71 100				
Except Ingril (mussels only*)	821	0.056	0.149	582	0.079	0.167	7.3	10.23	

- 431 * analyses of clams were only carried out for the Ingril lagoon
- 432 n: number of adult individuals in the population monitored during the survey (this population includes consumers
- 433 and non-consumers of mussels and clams from the first column)
- 434 Ave_pop: average exposure in the population (whether consumers or not) (µg/kg bw)
- 435 P95_pop: exposure to the 95th percentile calculated in the population (µg/kg bw/d)
- 436 n_cons: number of adult individuals consuming mussels and clams from the first column (consumers only)
- 437 Ave_cons: average exposure in the consumer population only (µg/kg bw/day)
- 438 P95_cons: exposure to the 95th percentile calculated in the consumer population only (µg/kg bw/day)
- 439 % > PABV: percentage of the general population exceeding the provisional acute benchmark value of 0.13 μg/kg
- 440 bw
- 441 % > PABV_cons: percentage of consumer population only exceeding the provisional acute benchmark value of
- 442 0.13 μg/kg bw

443

These results show that in the tested scenarios including the contamination data from the Ingril lagoon the provisional acute benchmark value would be exceeded in 71% of adults according to the consumption data from the CONSOMER study for the Mediterranean area (in the scenario including non-consumers of mussels and clams). If only individuals consuming mussels and clams are considered, this proportion increases to 100%. In the tested scenario excluding the contamination data from the Ingril lagoon, the provisional acute benchmark value would be exceeded in 7% of the total Mediterranean population and in 10% of the population of consumers.

451 6.2 Conclusion in terms of health concern

452 Considering the percentage of the French population for which the provisional acute benchmark value 453 would be exceeded in the "Except Ingril" scenario, there may be a health concern related to the 454 consumption of shellfish contaminated with PnTXs from these Mediterranean lagoons. Estimates were 455 not made for the other French sites where PnTXs were detected in mussels (Le Scoré, Diana) 456 because the contamination was measured only in the digestive glands, and the distribution of PnTXs 457 between the digestive gland and the total meat is not known.

458 **7. Recommendations for surveillance**

459 7.1 Environmental surveillance

Several studies on the ecology of the dinoflagellate *V. rugosum* have been carried out *in vitro* and *in situ.* However, its origin and the determinism of its blooms in natural environments remain to be elucidated. While the contamination of molluscs by PnTXs in some areas is undeniable, it remains difficult to establish the relationship with *V. rugosum* blooms as the observation of the pelagic phase in the water column is challenging. Official surveillance of shellfish production areas based on the identification and counting of toxic phytoplanktonic species in the water column is therefore ill-suited to this species.

467 Several monitoring options can be proposed:

Regarding dinoflagellate surveillance, monitoring of the benthic population of this organism in
risk areas is recommended. It is essential to sample the macroalgae present in the area, in
order to collect *V. rugosum* cells. The protocol set up for "*Ostreopsis*" monitoring and tested in
the study by Abadie *et al.* (2018) shows that such methodology is adequate.

Alongside the monitoring of *V. rugosum*, more systematic surveillance of the presence of
PnTX A and G toxins in molluscs should be considered. The extraction protocol for PnTXs is
identical to the one used for official monitoring of lipophilic toxins in the French shellfish
production areas (REPHYTOX). Although an additional step for LC-MS/MS analysis is
required, the cost remains moderate. A systematic screening for PnTXs in total meat could be
added to the REPHYTOX analyses of lipophilic toxins, and not only on samples taken for the
EMERGTOX scheme, that focuses only on the digestive gland.

479 Such proposal will enable estimating and monitoring the presence of PnTXs in bivalve480 molluscs in national production areas.

481

482 **7.2 Health monitoring, reporting and procedures**

To carry on with this issue, ANSES is working on raising awareness among health professionals of the potential neurological symptoms that could be associated with PnTXs poisoning in humans (Delcourt at al., 2019). An epidemiological study is currently underway with the objective to identify potential human cases related to shellfish consumption and PnTX exposure in France.

487

488 8. Conclusion

This work points out that, in case of high consumption and/or high contamination, the provisional acute benchmark value for PnTX G could be exceeded, suggesting a health concern related to the consumption of shellfish contaminated with PnTXs from Mediterranean lagoons, and particularly from the Ingril area. There is currently no shellfish production intended for sale from Ingril area. Nevertheless, ANSES highlighted the need to avoid all consumption of shellfish from this area. ANSES recommends that PnTXs should be taken into account in the French official monitoring of shellfish production areas.

The risk assessment underlying this conclusion is based on a worst-case scenario for estimating exposure to PnTX G, using the 95th percentile of shellfish contamination. A more realistic risk assessment could be conducted using a probabilistic approach (taking into account the distribution of all the data, on both contamination and consumption), which would require the acquisition of more contamination data in shellfish, particularly oysters (Anses, 2019).

502 9. Data gaps

503 ANSES recommends undertaking research to estimate more accurately the exposure of shellfish 504 consumers. More data on shellfish contamination by PnTXs are needed, including other species than 505 mussels and clams. More research is also needed to better characterize the oral toxicity of toxins 506 produced by V. rugosum. As portimine is produced in large quantities by Mediterranean strains of 507 Vulcanodinium rugosum, this toxin should be also considered. This toxin only seems to accumulate at 508 very low levels in shellfish. Additional data are required to confirm its low accumulation in shellfish. 509 Although less toxic than PnTXs in mice by the intraperitoneal route, the toxicological data for portimine 510 are limited and investigation should be carried on. Three of the identified PtTXs (A to C) are structural 511 analogues belonging to the PnTX group but, due to the lack of data available in the literature, they 512 could not be taken into account in this risk assessment. More toxicity and contamination studies are 513 needed.

An extensive acute oral toxicity study on PnTX G in rodents is needed (including description of clinical symptoms, assessment of biological, hematological and anatomopathological parameters as in the repeated dose studies, for a range of doses close to the maximum tolerated dose in the 2014 ANSES-University of Trieste-CNRS report study/Sosa et al., 2020), with a 14-day observation period.

A 28-day repeated oral administration study (OECD guideline 407) or even a 90-day study (OECD guideline 408) should be conducted. Given the data currently available, the effects on the central and peripheral nervous system as well as on the cardiovascular system should be investigated. Conducting a neurotoxicity study according to OECD guideline 424 is recommended.Preliminary results on *in ovo* exposure of chickens (Couesnon et al., 2014; unpublished results) suggest that PnTXs could affect embryonal growth. Therefore, the effects of PnTXs on offspring should be investigated, according to the OECD guideline 422.

525

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528

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531

532 References

- 533 Abadie, E., A. Muguet, T. Berteaux, N. Chomerat, P. Hess, E. Roque D'OrbCastel, E. Masseret, M.
- 534 Laabir, 2016. Toxin and Growth Responses of the Neurotoxic Dinoflagellate Vulcanodinium

rugosum to Varying Temperature and Salinity. Toxins 8(5). doi: 10.3390/toxins8050136.

536

Abadie, E., C. Chiantella, A. Crottier, L. Rhodes, E. Masseret, T. Berteaux, M. Laabir, 2018. What are
the main environmental factors driving the development of the neurotoxic dinoflagellate Vulcanodinium
rugosum in a Mediterranean ecosystem (Ingril lagoon, France)? Harmful Algae 75:75-86.

540 doi:10.1016/j.hal.2018.03.012.

541

542 Al Muftah, A., A.I. Selwood, A.J. Foss, H.M. Al-Jabri, M. Potts, M. Yilmaz, 2016. Algal toxins and 543 producers in the marine waters of Qatar, Arabian Gulf. Toxicon 122:54-66.

544

Anses (Agence nationale de sécurité sanitaire de l'alimentation, de l'environnement et du travail),
2017.

547 Valeurs toxicologiques de référence. Guide d'élaboration de l'Anses (only in French).

548 https://www.anses.fr/fr/system/files/SUBSTANCES2017SA0016Ra.pdf

549

Anses (Agence nationale de sécurité sanitaire de l'alimentation, de l'environnement et du travail),
2019. Risques liés aux pinnatoxines dans les coquillages. Avis de l'Anses et Rapport d'expertise
collective, Mars 2019 - Édition scientifique, 136 p,
https://www.anses.fr/fr/system/files/ERCA2016SA0013Ra.pdf (full report only in French).

554

Anses-Université de Trieste-CNRS, 2014. Pinnatoxines en lien avec l'espèce Vulcanodinium rugosum
(II) Partie 2 : Etudes sur la toxicité des pinnatoxines et d'extraits de Vulcanodinium rugosum, 72p.

557 https://archimer.ifremer.fr/doc/00285/39635/38127.pdf

558

- Araoz, R., D. Servent, J. Molgó, B.I. lorga, C. Fruchart-Gaillard, E. Benoit, Z. Gu, C. Stivala. A.
 Zakarian. 2011, Total synthesis of pinnatoxins A and G and revision of the mode of action of
 pinnatoxin A. J. Am. Chem. Soc. 133:10499-10511.
- 562
- Benoit, E., A. Couesnon, J. Lindovsky, B.I. lorga, R. Araoz, D. Servent, .A. Zakarian, J. Molgó. 2019.
 Synthetic pinnatoxins A and G reversibly block mouse skeletal neuromuscular transmission In Vivo
 and In Vitro. Mar. Drugs 17(5). pii: E306. doi: 10.3390/md17050306.
- 566 Couesnon, A., J. Lindovsky, A. Zakarian, S. Creuzet, J. Molgó, 2014. Pinnatoxins block skeletal 567 neuromuscular junction activity and affect embryo development. Toxicon 91:175-176. 568 doi:10.1016/j.toxicon.2014.08.036.
- 569
- 570 Chou, T., T. Haino, M. Kuramoto, D. Uemura, 1996. Isolation and structure of pinnatoxin D, a new 571 shellfish poison from the Okinawan bivalve Pinna muricata. Tetrahedron Letters 37(23): 4027-4030.
- 572
- Delcourt, N., Lagrange, E., Abadie, E., Fessard, V., Frémy, J.-M., Vernoux, J.-P., Peyrat, M.-B.,
 Maignien, T., Arnich, N., Molgó, J., Mattei, C., 2019. Pinnatoxins' deleterious effects on cholinergic
 networks: From experimental models to human health. Marine Drugs 17, Issue 7, Article number 425.
- 577 EFSA (European Food Safety Authority). 2010a. Scientific Opinion on marine biotoxins in shellfish –
 578 Cyclic imines (spirolides, gymnodimines, pinnatoxins and pteriatoxins). EFSA Journal 8 (6):1628. doi:
 579 doi:10.2903/j.efsa.2010.1628.
- 580

EFSA (European Food Safety Authority). 2010b. Statement on further elaboration of the consumption
figure of 400 g shellfish meat on the basis of new consumption data. EFSA Journal 8 (8):1706.
doi:10.2903/j.efsa.2010.1706.

584

Garcia-Altares, M., A. Casanova, V. Bane, J. Diogene, A. Furey, P. de la Iglesia, 2014. Confirmation of
pinnatoxins and spirolides in shellfish and passive samplers from Catalonia (Spain) by liquid
chromatography coupled with triple quadrupole and high-resolution hybrid tandem mass spectrometry.
Mar Drugs 12 (6):3706-32. doi: 10.3390/md12063706.

- 589
- Garrett, M. J., C. Puchulutegui, A. I. Selwood, J. L. Wolny. 2014. Identification of the harmful
 dinoflagellate Vulcanodinium rugosum recovered from a ballast tank of a globally traveled ship in Port
 Tampa Bay, Florida, USA." Harmful Algae 39:202-209. doi: 10.1016/j.hal.2014.07.014.
- 593
- Hellyer, S. D., A. I. Selwood, L. Rhodes, D. S. Kerr, 2013. Neuromuscular blocking activity of pinnatoxins
- 596 E, F and G. Toxicon 76:214-20. doi: 10.1016/j.toxicon.2013.10.009.
- 597
- Hellyer, S.D., D. Indurthi, T. Balle, V. Runder-Varga, A.I. Selwood, J.D.Tyndall, M. Chebib, L. Rhodes,
- D.S. Kerr. 2015. Pinnatoxins E, F and G target multiple nicotinic receptor subtypes. J. Neurochem.
 135:479-91. doi: 10.1111/jnc.13245.
- 601
- Hernandez-Becerril, D. U., M. C. Rodriguez-Palacio, C. Lozano-Ramirez, 2013. Morphology and life
 stages of the potentially pinnatoxin-producing thecate dinoflagellate Vulcanodinium rugosum from the
 tropical Mexican Pacific. Botanica Marina 56 (5-6):535-540. doi: 10.1515/bot-2013-0079.
- 605
- Ifremer (Institut français de recherche pour l'exploitation de la mer). 2012. P. Hess, F. Hervé, E.
 Abadie, V. Séchet, J. Molgó, Z. Amzil, V. Fessard. Pinnatoxines en lien avec l'espèce Vulcanodinium
 rugosum. Référence Ifremer: 11/1219401/NYF, 09-2012 R.RBE/EMP/PHYC 12-05, 144 p.
 https://archimer.ifremer.fr/doc/00094/20518/18190.pdf
- 610
- Klimisch, H. J., M. Andreae, U. Tillmann, 1997. A systematic approach for evaluating the quality of
 experimental toxicological and ecotoxicological data." Regul Toxicol Pharmacol 25 (1):1-5.
 doi:10.1006/rtph.1996.1076.
- 614
- Lamas, J.P., Arévalo, F., Moroño, Á., Correa, J., Muñíz, S., Blanco, J. 2019. Detection and spatiotemporal distribution of pinnatoxins in shellfish from the Atlantic and Cantabrian coasts of Spain.
 Toxins 11(6). pii: E340. doi: 10.3390/toxins11060340.
- 618

- MacKenzie, L. A., A. I. Selwood, P. McNabb, L. Rhodes, 2011. Benthic dinoflagellate toxins in two
 warmtemperate estuaries: Rangaunu and Parengarenga Harbours, Northland, New Zealand. Harmful
 Algae 10 (6):559-566. doi: 10.1016/j.hal.2011.02.007.
- 622
- 623 McCarron, P., W. A. Rourke, W. Hardstaff, B. Pooley, M. A. Quilliam, 2012. Identification of 624 pinnatoxins
- and discovery of their fatty acid ester metabolites in mussels (Mytilus edulis) from eastern Canada. J
 Agric Food Chem 60 (6):1437-46. doi: 10.1021/jf204824s.
- 627
- McCauley, J.A., K. Nagasawa, P.A., S.G. Mischke, M.A. Semones, Y. Kishi, 1998. Total synthesis of
 pinnatoxin A. J. Am. Chem. Soc. 120(30):7647-7648.
- 630
- 631 McNabb, P. S., D. J. McCoubrey, L. Rhodes, K. Smith, A. I. Selwood, R. van Ginkel, A. L. MacKenzie, R. Munday, P. T. Holland, 2012. New perspectives on biotoxin detection in Rangaunu Harbour, New 632 633 Zealand discovery Harmful arising from the of pinnatoxins. Algae 13:34-39. 634 doi:10.1016/j.hal.2011.09.013.
- 635
- Molgó, J., P. Marchot, R. Aráoz, E. Benoit, B.I. Iorga, A. Zakarian, P. Taylor, Y. Bourne, D. Servent.
 2017. Cyclic imine toxins from dinoflagellates: a growing family of potent antagonists of the nicotinic
 acetylcholine receptors. J. Neurochem., 142 S2:41-51. doi: 10.1111/jnc.13995.
- 639
- Moreira-Gonzalez, A.R., A. Comas-Gonzalez, A. Valle-Pombrol, M. Seisdedo-Losa, C. Alonsohernadez, O. Hernandez-Leyva, L.F. Ferandes, N. Chomerat, G. Bilien, F. Herve, G.A. Rovillon, P.
 Hess, L.L. Mafra, 2018. Summer bloom of Vulcanodinium rugosum in Cienfuegos Bay (Cuba)
 associated to dermatitis in swimmers. 18th edition of the International Conference on Harmful Algae
 (ICHA), 21- 26 October 2018, Nantes (France). Poster 238. Abstract book
 https://www.icha2018.com/medias/content/files/ICHA 2018 Abstract Book v8.pdf

Munday, R., A. I. Selwood, L. Rhodes, 2012. Acute toxicity of pinnatoxins E, F and G to mice. Toxicon
60 (6):995-999.

Nézan, E., Chomérat, N., 2011. Vulcanodinium rugosum gen. nov., sp. nov. (dinophyceae): A new
marine dinoflagellate from the French Mediterranean coast. Cryptogam. Algol. 32, 3–18.

652

- Otero, P., Miguéns, N., Rodríguez, I., Botana, L.M. 2019. LC-MS/MS Analysis of the Emerging Toxin
 Pinnatoxin-G and High Levels of Esterified OA Group Toxins in Galician Commercial Mussels. Toxins
 11(7), 394. doi: 10.3390/toxins11070394.
- 656
- Rambla-Alegre, M., C. O. Miles, P. de la Iglesia, M. Fernandez-Tejedor, S. Jacobs, I. Sioen, W.
 Verbeke, I. A. Samdal, M. Sandvik, V. Barbosa, A. Tediosi, E. Madorran, K. Granby, M. Kotterman, T.
 Calis, J. Diogene, 2018. Occurrence of cyclic imines in European commercial seafood and consumers
- 660 risk
- 661 assessment. Environ Res 161:392-398. doi: 10.1016/j.envres.2017.11.028.
- 662

Rhodes, L., K. Smith, A. Selwood, P. McNabb, R. van Ginkel, P. Holland, R. Munday, 2010.
Production of pinnatoxins by a peridinoid dinoflagellate isolated from Northland, New
Zealand."Harmful Algae 9(4):384-389. doi: 10.1016/j.hal.2010.01.008.

- 666
- Rhodes, L., K. Smith, A. Selwood, P. McNabb, S. Molenaar, R. Munday, C. Wilkinson, G. Hallegraeff,
 2011. Production of pinnatoxins E, F and G by scrippsielloid dinoflagellates isolated from Franklin
 Harbour, South Australia. New Zealand Journal of Marine and Freshwater Research 45 (4):703-709.
 doi: 10.1080/00288330.2011.586041.
- 671
- Rundberget, T., J. A. Aasen, A. I. Selwood, C. O. Miles, 2011. Pinnatoxins and spirolides in Norwegian
 blue mussels and seawater. Toxicon 58 (8):700-11. doi: 10.1016/j.toxicon.2011.08.008.

674

Selwood, A. I., C. O. Miles, A. L. Wilkins, R. van Ginkel, R. Munday, F. Rise, P. McNabb, 2010.
Isolation, structural determination and acute toxicity of pinnatoxins E, F and G. J Agric Food Chem
58(10):6532-42. doi: 10.1021/jf100267a.

678

- Selwood, A. I., A. L. Wilkins, R. Munday, F. Shi, L. L. Rhodes, P. T. Holland, 2013. Portimine: A
 bioactive metabolite from the benthic dinoflagellate Vulcanodinium rugosum. Tetrahedron Letters
 54(35):4705-4707. doi: 10.1016/j.tetlet.2013.06.098.
- 682
- Selwood, A. I., A. L. Wilkins, R. Munday, H. Gu, K. F. Smith, L. L. Rhodes, F. Rise, 2014. Pinnatoxin
 H:
- A new pinnatoxin analogue from a South China Sea Vulcanodinium rugosum isolate. Tetrahedron
 Letters 55 (40):5508-5510. doi: 10.1016/j.tetlet.2014.08.056.
- 687
- Smith, K. F., L. L. Rhodes, S. Suda, A. I. Selwood, 2011. A dinoflagellate producer of pinnatoxin G,
 isolated from sub-tropical Japanese waters." Harmful Algae 10 (6):702-705.
 doi:10.1016/j.hal.2011.05.006.
- 691
- Sosa, S., Pelin, M., Cavion, F., Hervé, F., Hess, P., Tubaro, A. 2020. Acute oral toxicity of pinnatoxin
 G in mice. Toxins (Basel). 2020 Jan 28;12(2). pii: E87. doi: 10.3390/toxins12020087.
- 694
- Takada, N., N. Umemura, K. Suenaga, D. Uemura, 2001. Structural determination of pteriatoxins A, B
 and C, extremely potent toxins from the bivalve Pteria penguin. Tetrahedron Letters 42 (20):34953497. doi: 10.1016/S0040-4039(01)00478-6.
- 698
- Uemura, D., T. Chou, T. Haino, A. Nagatsu, S. Fukuzawa, S. Z. Zheng, H. S. Chen, 1995. Pinnatoxina -a toxic amphoteric macrocycle from the Okinawan bivalve Pinna muricata. J Am Chem Soc
 117(3):1155-1156. doi: 10.1021/ja00108a043.
- 702
- Zendong, Z., C. Herrenknecht, E. Abadie, C. Brissard, C. Tixier, F. Mondeguer, V. Sechet, Z. Amzil, P.
 Hess. 2014. Extended evaluation of polymeric and lipophilic sorbents for passive sampling of marine
 toxins. Toxicon 91:57-68. doi: 10.1016/j.toxicon.2014.03.010.
- 706
- Zeng, N., H. Gu, K. F. Smith, L. L. Rhodes, A. I. Selwood, W. Yang, 2012. The first report of
 Vulcanodinium rugosum (Dinophyceae) from the South China Sea with a focus on the life cycle. New

709	Zealand	Journal	of	Marine	and	Freshwater	Research	46	(4):511-521.
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710 doi:10.1080/00288330.2012.719911.