

Combining seroprevalence and capture-mark-recapture data to estimate the force of infection of brucellosis in a managed population of Alpine ibex

Sébastien Lambert, Emmanuelle Gilot-Fromont, Carole Toïgo, Pascal Marchand, Elodie Petit, Sophie Rossi, Anne Thébault

▶ To cite this version:

Sébastien Lambert, Emmanuelle Gilot-Fromont, Carole Toïgo, Pascal Marchand, Elodie Petit, et al.. Combining seroprevalence and capture-mark-recapture data to estimate the force of infection of brucellosis in a managed population of Alpine ibex. Epidemics, 2022, 38, pp.100542. 10.1016/j.epidem.2022.100542. anses-03597873

HAL Id: anses-03597873 https://anses.hal.science/anses-03597873v1

Submitted on 22 Jul2024

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NonCommercial 4.0 International License

Version of Record: https://www.sciencedirect.com/science/article/pii/S1755436522000056 Manuscript_79c9ee206bff931bb4482d6ba9cacc29

- 1 Title: Combining seroprevalence and capture-mark-recapture data to estimate the force
- 2 of infection of brucellosis in a managed population of Alpine ibex
- 3

4 Author names and affiliations:

- 5 Sébastien Lambert ^{a*}, Emmanuelle Gilot-Fromont ^b, Carole Toïgo ^c, Pascal Marchand ^d,
- 6 Elodie Petit^{b,e}, Sophie Rossi^{f**}, Anne Thébault^g
- 7
- ⁸ ^a Université de Lyon, Université Lyon 1, CNRS, Laboratoire de Biométrie et Biologie
- 9 Évolutive UMR 5558, Villeurbanne, France
- 10 ^b Université de Lyon, VetAgro Sup, CNRS, Laboratoire de Biométrie et Biologie Évolutive
- 11 UMR 5558, Villeurbanne, France
- 12 ^c Office Français de la Biodiversité (OFB), Unité Ongulés Sauvages, Gières, France
- ¹³ ^d Office Français de la Biodiversité (OFB), Unité Ongulés Sauvages, Juvignac, France
- ^e Office Français de la Biodiversité (OFB), Unité Sanitaire de la Faune, Sévrier, France
- ¹⁵ ^f Office Français de la Biodiversité (OFB), Unité Sanitaire de la Faune, Gap, France
- ^g Agence nationale de sécurité sanitaire, de l'alimentation, de l'environnement et du travail
- 17 (Anses), Direction de l'Évaluation des Risques, Maisons-Alfort, France
- ^{*} Present address: Centre for Emerging, Endemic and Exotic Diseases, Department of
- 19 Pathobiology and Population Sciences, Royal Veterinary College, University of London,
- 20 Herts, UK
- 21 ** The authors wish to dedicate this work to the beloved memory of Dr Sophie Rossi, who
- sadly passed away during the preparation of this manuscript. She dedicated her career to
- 23 further the understanding of wildlife diseases and improve wildlife health, and her passionate
- 24 work will remain an inspiration to us all.
- 25

- 26 Corresponding author: Dr Sébastien Lambert, Department of Pathobiology and Population
- 27 Sciences, Royal Veterinary College, University of London, Hertfordshire, AL9 7TA, United
- 28 Kingdom, selambert@rvc.ac.uk

30 Abstract: In wildlife, epidemiological data are often collected using cross-sectional surveys 31 and antibody tests, and seroprevalence is the most common measure used to monitor the 32 transmission dynamics of infectious diseases. On the contrary, the force of infection, a 33 measure of transmission intensity that can help understand epidemiological dynamics and 34 monitor management interventions, remains rarely used. The force of infection can be 35 derived from age-stratified cross-sectional serological data, or from longitudinal data 36 (although less frequently available in wildlife populations). Here, we combined 37 seroprevalence and capture-mark-recapture data to estimate the force of infection of 38 brucellosis in an Alpine ibex (Capra ibex) population followed from 2012 to 2018. Because 39 the seroprevalence of brucellosis was 38% in this population in 2012, managers conducted 40 two culling operations in 2013 and 2015, as well as captures every year since 2012, where 41 seronegative individuals were marked and released, and seropositive individuals were 42 removed. We obtained two estimates of the force of infection and its changes across time, by 43 fitting (i) a catalytic model to age-seroprevalence data obtained from unmarked animals 44 (cross-sectional), and (*ii*) a survival model to event time data obtained from recaptures of 45 marked animals (longitudinal). Using both types of data allowed us to make robust inference 46 about the temporal dynamics of the force of infection: indeed, there was evidence for a decrease in the force of infection between mid-2014 and late 2015 in both datasets. The force 47 of infection was estimated to be reduced from 0.115 year⁻¹ [0.074–0.160] to 0.016 year⁻¹ 48 49 [0.001–0.057]. These results confirm that transmission intensity decreased during the study 50 period, probably due to management interventions and natural changes in infection dynamics. 51 Estimating the force of infection could therefore be a valuable complement to classical 52 seroprevalence analyses to monitor the dynamics of wildlife diseases, especially in the 53 context of ongoing disease management interventions.

- 55 Keywords (5): wildlife disease management; force of infection; seroprevalence; catalytic
- 56 model; survival analysis

57 INTRODUCTION

58 Recent outbreaks of wildlife diseases in Europe such as Chronic Wasting Disease or African 59 Swine Fever have brought to light the need for effective, science-based disease management 60 strategies in the wild (Mysterud and Rolandsen, 2018; Vicente et al., 2019). Depending on 61 the host species and infectious disease considered, interventions such as culling, fencing, test-62 and-remove or vaccination have been implemented with various success in wild populations 63 (le Roex et al., 2015; McDonald et al., 2008; Müller et al., 2015; Mysterud and Rolandsen, 64 2019). The first step in the choice of an appropriate management intervention is a sound 65 evaluation of the epidemiological situation, in order to optimize strategies before any action 66 is implemented (Artois et al., 2009). Then, ongoing management interventions should be 67 accompanied by a close monitoring of both the infection and the population dynamics to 68 assess their effectiveness (Joseph et al., 2013; Vicente et al., 2019). However, the 69 management of wildlife diseases is often hampered by our limited knowledge of population 70 and infection dynamics, and management actions are often implemented before a full picture 71 of the situation has been drawn. It would therefore be desirable to extract the most 72 information out of the available data to inform management decisions. 73 Incidence, i.e. the number of new infections in a population at risk over a given period of 74 time, and prevalence, the proportion of infected individuals at a given point in time or over a 75 certain period of time, are two common measures used to describe epidemiological dynamics 76 of infections (Keiding, 1991). In wildlife populations, the collection of epidemiological data 77 is often performed using cross-sectional surveys and serological (antibody) tests (Gilbert et 78 al., 2013). Therefore, the prevalence of individuals positive to antibody tests (called 79 seroprevalence) is often the only measure used to monitor the dynamics of infections in 80 managed populations (Gilbert et al., 2013). However, the variation of seroprevalence over 81 time provides only limited information on transmission dynamics, as the presence of

82 antibodies does not necessarily reflect active infections or inform on the timing of infection, 83 especially in long-lived species. Unfortunately, incidence is rarely available for wildlife 84 populations, because it requires longitudinal samplings of individual animals which are often 85 logistically difficult (Gilbert et al., 2013). The force of infection (FOI), on the other hand, is a 86 measure of transmission intensity related to incidence that can be obtained from age-87 seroprevalence data collected during cross-sectional surveys (Heisey et al., 2006) and that 88 can help planning and optimizing management interventions (Lewis et al., 2014). 89 The FOI is the rate at which susceptible individuals become infected, and combines the 90 contact rate, the probability of transmission given contact, and the number of infectious 91 individuals in the population (Begon et al., 2002). Classical methods for estimating the FOI 92 are based on the fact that the proportion of individuals who have ever been infected increases 93 with age as a result of increased time of exposure to the infection (Heisey et al., 2006; Hens 94 et al., 2010). These methods, also described as catalytic models (Muench, 1934), have been 95 widely used for life-long immune infections in humans (Farrington, 1990; Keiding, 1991; 96 Papoz et al., 1986). A common assumption in these models is that the FOI is constant over 97 time, or at least constant on average (Hens et al., 2010), which is often not the case for 98 diseases that are emerging or under management. Nonetheless, several studies were able to 99 estimate time-varying FOI in humans (Katzelnick et al., 2018) and animals (Courtejoie et al., 100 2018). Estimating the FOI from age-seroprevalence data also has the additional advantage of 101 assessing for infection-related mortality (Heisey et al., 2006; Reynolds et al., 2019). 102 The aim of this study was to estimate the FOI of brucellosis (Brucella melitensis) in Alpine 103 ibex (*Capra ibex*) in the Bargy Massif (French Alps). This is the first reported example of 104 self-sustained brucellosis infection in European wild-living ungulates, with a seroprevalence 105 level as high as 38% at the beginning of monitoring in 2012-2013 (Garin-Bastuji et al., 2014; 106 Hars et al., 2013). As France has been declared officially free of *Brucella melitensis* and

107 Brucella abortus in domestic ruminants since 2005 (Perrin et al., 2016a, 2016b), the 108 persistence of *B. melitensis* in a wildlife population raises serious public health and economic 109 concerns. This led the French authorities to undertake several management interventions 110 since the discovery of the outbreak in 2012 (Lambert et al., 2018; Marchand et al., 2017). 111 Among these, two massive culling operations were carried out in autumn 2013-spring 2014 112 (*n*=251 individuals culled, estimated population size before culling: 567, 95% CI [487-660]) 113 and in autumn 2015 (*n*=70 individuals culled, estimated population size before culling: 344, 114 95% CI [290-421]). In parallel, the population was monitored by Capture-Mark-Recapture 115 (CMR) to collect demographic and epidemiological data every year. During captures (n=387 116 between 2012 and 2018), a test-and-remove program was implemented where seropositive 117 individuals were removed (n=119), while seronegative individuals were marked and released. 118 The 2015 test-and-remove campaign was particularly intense with 38 individuals removed 119 among the 125 captured. 120 As a consequence, cross-sectional data are available from unmarked animals, whereas the 121 follow up of marked animals can be seen as data of a cohort study of seronegative animals. Contrary to the seroprevalence (Calenge et al., 2021; Marchand et al., 2017), the FOI has 122 123 never been studied in the population before, although Calenge et al., 2021 estimated the FOI

124 exerted on susceptible marked animals only, and assuming that it was proportional to the

125 proportion of the total ibex population being seropositive, unmarked and actively infected.

126 Therefore, we were interested in estimating the FOI from the available datasets to get a

127 complementary picture of the epidemiological dynamics in the study population.

128 In ruminants, brucellosis is considered to produce lifelong infections, in particular in goats

129 (European Commission, 2001; Schlafer and Miller, 2007), a species closely related to ibex.

130 Brucella infection induces both humoral and cellular immune responses (Carvalho Neta et

131 al., 2010; European Commission, 2001), which remains detectable over long periods of time

132 (Godfroid et al., 2010). The characteristics of brucellosis are therefore compatible with the133 use of catalytic models.

134 The presence of antibodies detected by serologic tests is used to detect previous exposure to 135 Brucella (Godfroid et al., 2010). However, a potential issue with using antibodies as a proxy 136 of past exposure is that infected individuals could be missed if antibody levels wane over 137 time and decrease below detectable levels (Benavides et al., 2017). In elk (Cervus 138 *canadensis*) infected with *B. abortus*, evidence suggest lifelong protective immunity but a 139 progressive antibody loss after some years, which explains the decline in seroprevalence in 140 older individuals (Benavides et al., 2017). In bison (Bison bison), a similar decline in 141 seroprevalence in older individuals is observed (Treanor et al., 2011). This could be 142 explained by the finding that active infections, which can be reactivated on favourable 143 occasions such as oestrus or pregnancy (European Commission, 2001; Schlafer and Miller, 144 2007) and are associated with increased antibody levels (Durán-Ferrer et al., 2004), decreases 145 with age in seropositive bison (Treanor et al., 2011). 146 In ibex, similar patterns have been found with a decline of seroprevalence in older individuals 147 (Marchand et al., 2017) and a decrease of the probability of active infection with age 148 (Lambert et al., 2018). Therefore, a partial antibody loss cannot be excluded; however, in the 149 absence of relevant data to estimate this process, we assumed life-long infection and 150 seropositivity in our study. 151 Infection-related mortality is another mechanism that could explain a decrease of 152 seroprevalence with age (Benavides et al., 2017; Heisey et al., 2006). Therefore, one aim of the present study was to look for evidence of infection-related mortality, which can be 153 154 estimated at the same time as the FOI (Heisey et al., 2006; Reynolds et al., 2019). Although 155 there is currently no evidence for brucellosis-related mortality in the Alpine ibex population 156 of the Bargy massif, it remains a possibility, e.g. because of brucellosis-related lesions such

157 as arthritis (Freycon et al., 2017). In other wild species, evidence for brucellosis-related 158 mortality were found in moose (Alces alces) and African buffalo (Syncerus caffer) (Forbes et 159 al., 1996; Gorsich et al., 2015), but not in bison or elk (Benavides et al., 2017; Fuller et al., 160 2007; Joly and Messier, 2005). 161 Our study also aimed at evaluating temporal changes in the FOI between October 2012 and 162 June 2018. The seroprevalence did not change between 2012 and 2015 (Marchand et al., 163 2017), when management interventions were dominated by massive culling operations. 164 However, a recent study demonstrated that it then decreased between 2015 and 2016, before 165 getting roughly stable from 2016 to 2018 (Calenge et al., 2021). Management interventions in 166 this second period were mainly based on test-and-remove during captures, especially in 2015 167 when 125 captures were achieved, followed by around 30-50 captures each following year. 168 Given this information, we hypothesized that the FOI also decreased between 2015 and 2016, 169 at the same time as the seroprevalence and following changes in the management 170 interventions. Alternatively, if individuals remain immune lifelong after infection, test-and-171 remove of seropositive individuals could also increase the FOI by specifically removing 172 immune individuals (Ebinger et al., 2011). We also hypothesized that the FOI could change 173 in 2012, as there was no management in the study population before brucellosis was detected. 174 We therefore expected three periods, one before 2012, and two others between 2012 and 175 2018.

176

177 MATERIAL AND METHODS

178 **Population management and monitoring**

179 The study area, ibex population and discovery of *B. melitensis* infection have been described

180 previously (ANSES, 2015; Garin-Bastuji et al., 2014; Hars et al., 2013; Mailles et al., 2012;

181 Marchand et al., 2017; Mick et al., 2014).

182 The estimates of the population size were: 567 individuals (95% Confidence Interval: [487-183 660]) in 2013, 310 [275-352] in 2014, 347 [290-421] in 2015, 272 [241-312] in 2016, 291 184 [262-327] in 2017 and 374 [326-435] in 2018 (Calenge et al., 2021; Marchand et al., 2017). Between the beginning of the monitoring by CMR in October 2012 and summer 2018 (see 185 186 Figure 1 for the timing of captures and recaptures), a total of 387 captures and recaptures 187 have been performed (2012: 24, 2013: 57, 2014: 71, 2015: 125, 2016: 35, 2017: 27 and 2018: 188 48 – Lambert et al., 2020, 2018; Marchand et al., 2017). For each individual, information on 189 age (in years, by counting horn growth annuli, Michallet et al., 1988), sex, date of sampling 190 and spatial location of capture were systematically recorded. Captured animals were blood-191 sampled by trained technical staff and serological tests were performed on serum samples. 192 The serological antibody tests were performed according to requirements of the European 193 Union for diagnosis of brucellosis in small ruminants and following standards of the World 194 Organization of Animal Health (OIE). Four serological tests were performed in parallel in 195 laboratory conditions: the Rose Bengal Test, the Complement Fixation Test, and the indirect 196 and the competitive Enzyme Linked ImmunoSorbent Assays. Individuals were considered as 197 seropositive when at least two of the tests were positive. Sensitivities and specificities of 198 these four tests for the diagnosis of *B. melitensis* in small domestic ruminants are generally 199 high: 75-100% and 97.6-100%, respectively (Blasco et al., 1994b, 1994a; Ferreira et al., 200 2003; Nielsen et al., 2004). Although they remain unknown in ibex, the sensitivity and 201 specificity of the diagnosis protocol in ibex are considered high because of the use of 202 multiple tests in parallel and few cases of inconsistent results (ANSES, 2015). As part of a 203 test-and-remove program, captured individuals were either euthanized when they were 204 seropositive, or marked and released if they were seronegative at the time of sampling. Since 205 2015, the decision to euthanize ibex was based on the results of a rapid Lateral Flow Immune-chromatographic Assay (LFIA), available for direct use in the field before 206

207 confirmation by the other tests in the laboratory. The LFIA had been validated on ibex samples
208 by the EU/OIE/FAO and National reference laboratory for animal brucellosis, showing very good
209 correlation with the four laboratory tests (ANSES, 2014; Corde et al., 2014).

210 In addition to CMR monitoring and test-and-remove operations, culling operations were also 211 implemented as part of the disease management strategy conducted by the French Authorities 212 (Figure 1). In spring 2013, four individuals showing clinical signs (e.g., lameness or presence 213 of visible gross lesions of the joints or the testes) were culled. In autumn 2013 and spring 214 2014, 233 and 18 individuals estimated to be older than 5 years were culled, based on the 215 observation that seroprevalence was significantly higher for this age class (Hars et al., 2013). 216 Non-selective culling of individuals that were never captured was also performed in autumn 217 2015 (n=70), autumn 2017 (n=5) and spring 2018 (n=5). The same antibody tests as in 218 captured animals were performed on culled ones whenever blood samples of good quality 219 were available. Among the 42 culled individuals for which results of serological tests were 220 available, the serological status of culled animals was based on a single LFIA test in a few 221 cases (n=12). This was also the case for one captured animal in 2015, and two marked 222 animals recaptured (all three with negative test results).

223

224 Cross-sectional and longitudinal datasets

In total, cross-sectional serological data were available from 363 unmarked individuals tested
between 2012 and 2018 (320 captured individuals, one found-dead individual and 42 culled
individuals- 129 seropositive and 234 seronegative results; Table S1). The first dataset we
used was therefore age-seroprevalence data from these unmarked individuals (Figure 2A,
Table S1 and Figure S1).

Among the seronegative individuals that were marked and released, 50 were recaptured at

231 least once during the study period and were tested again for antibodies against brucellosis.

232 The second dataset we used was therefore the follow up time (in days) between the first 233 capture and the last recapture of marked animals. Because these animals were seronegative at 234 the first capture, they either remained seronegative (n=46) or became seropositive in the 235 interval (n=4 – Figure 2B and Figure S1). This type of data is also called "event time" data, 236 where the event here would be the seroconversion from negative to positive. Individuals that 237 seroconverted are "interval censored", i.e., the event of interest occurred between the last 238 negative test and the positive test, but the exact time is not known (Kleinbaum and Klein, 239 2005). Conversely, individuals that remained seronegative at the last recapture are "right 240 censored", i.e., the event of interest did not occur yet and may or may not occur in the future 241 (Kleinbaum and Klein, 2005).

242 The longitudinal event time dataset covered the period 2012-2018, from first captures to last 243 recaptures (Figure 2B), while the cross-sectional age-seroprevalence dataset covered the 244 period from 1998 to 2018 (see below and Figure S1). The sample size for the longitudinal 245 dataset (n=50) was also much smaller than for the cross-sectional dataset (n=363), because in 246 the study population, most captures targeted unmarked individuals, in an attempt to test new 247 individuals and maximise chances to remove infected individuals (Calenge et al., 2021). 248 Indeed, marked ibex have more chances to be seronegative, as they already were during their 249 last capture in a more or less recent past (Calenge et al., 2021).

250

251 Estimating the FOI using cross-sectional age-seroprevalence data

252 The FOI $\lambda(t)$ represents the rate at which susceptible individuals become infected per unit of 253 time, at time t (Vynnycky and White, 2010), and can be calculated using the equation: $\lambda(t) =$ 254 $cv \frac{I(t)}{N(t)}$, where *c* is the contact rate, *v* is the probability that the contact leads to infection, and 255 $\frac{I(t)}{N(t)}$ is the proportion of infectious individuals in the population at time *t* (Begon et al., 2002). The FOI changes over time, although its average value remains unchanged over time inendemic situations.

If we were to follow over time a cohort of individuals born the same year, we would observe that, as the cohort ages, the proportion of individuals who have ever been infected increases (in our case, the proportion of seropositive individuals as we are using seropositivity as a proxy of past infection). A cross-sectional sample would produce a similar pattern, as animals would be of different ages and therefore would have experienced various exposure times (Heisey et al., 2006). This has led to the development of the 'catalytic model' (Muench, 1934), that can be written as:

265 $z(a) = 1 - e^{-\lambda a}$

where z(a) is the proportion ever infected at age a (in years), and λ is the average FOI (in years⁻¹). The main assumptions are that the average FOI λ is independent of age and year, that infection is lifelong or confers lifelong immunity, that all individuals are born without infection, and that susceptible and infected animals have the same mortality rate (i.e., no infection-related mortality experienced by infected animals).

271 When antibodies remain detectable lifelong, this model can be fitted to observed age-

seroprevalence data to estimate the average FOI. However, the limitations related to the

273 model assumptions led to new developments over the years, dealing for instance with age-

274 dependent FOI (see Hens et al., 2010 for a review).

In this study, we used the general FOI model developed by Heisey et al. (2006) that allows for the existence of infection-related mortality in infected animals and the influence of covariates $X_1, ..., X_n$ on the FOI λ , leading to the new equation:

278
$$z(a) = \frac{1 - e^{-(\lambda - \mu)a}}{1 - \frac{\mu}{\lambda}e^{-(\lambda - \mu)a}}$$

where z(a) is the proportion of seropositive at age a, μ is the infection-related mortality (infection being brucellosis in our case) and λ is the average FOI with $\lambda = \lambda_0 e^{\beta_1 X_1 + \dots + \beta_n X_n}$, where λ_0 is the baseline FOI which is constant over age and time. Note that when $\mu = 0$, the equation simplifies to Muench's model: $z(a) = 1 - e^{-\lambda a}$. The model still assumes that all animals are born without infection, but can immediately leave this state and become positive (Heisey et al., 2006).

285 We tested for the existence of infection-related mortality by fitting this general FOI model to 286 our age-seroprevalence data using a maximum likelihood approach and the R function 287 currentstatus.1 provided in Heisey et al. (2006). We took into account the effects of sex and 288 socio-spatial unit, which were demonstrated to play a major role in explaining variation in 289 brucellosis seroprevalence in the study ibex population (Lambert et al., 2020; Marchand et 290 al., 2017). Indeed, it has been highlighted that females transmit the infection in ~90% of 291 cases (Lambert et al., 2020) and that they are spatially structured in five subpopulations with 292 contrasted seroprevalence levels, the units at the centre of the massif ("core area") having the 293 highest seroprevalence levels (Marchand et al., 2017).

We also tested the assumption that λ is independent of age. First, we added age (as a

295 quantitative variable) among the covariates (Table 1), even though, in this case, the function

296 for z(a) would be incorrect, as it was obtained under the hypothesis that λ_0 was independent

of age. However, this step is useful as a diagnostic tool, as if age or a function of age as

298 covariate improves the model fit, it means that this assumption should be revised and

functions other than constant with age should be explored for the baseline FOI (Heisey et al.,

300 2006). Second, we used the piecewise constant hazards approach developed by Heisey et al.

301 (2006) to enable the use of age-varying models for the baseline FOI λ_0 such as the Weibull,

302 the Gompertz or the log-logistic models (Table S2).

303 Similarly, we added year of sampling (as a categorical variable) among the variables 304 (Table 1), to test if the time-independence assumption holds (Heisey et al., 2006). 305 For each type of model (constant FOI, diagnostic model with covariate year – YEAR, 306 diagnostic model with covariate age - AGE, diagnostic model with covariate quadratic age -307 AGE², diagnostic model with both AGE or AGE² and YEAR, Weibull, Log-logistic, Pareto, Gompertz), we fitted submodels including infection-related mortality (μ) or not, taking into 308 309 account socio-spatial unit (UNIT) and sex (SEX) (Table 1 and Table S2). For each model, the 310 number of parameters (degrees of freedom), the log-likelihood, the Akaike Information 311 Criterion corrected for small sample sizes (AICc), and the difference between the AICc and 312 the lowest AICc value (Δ AICc) were computed. The Akaike Weights, which are relative 313 model likelihoods normalized over the likelihoods of all possible submodels, were computed 314 using the function aictabCustom of the AICcmodavg R package (Mazerolle, 2019). The 315 Akaike weight can be regarded as the conditional probability given the data of being the best 316 model among the set of possible submodels (Wagenmakers and Farrell, 2004). We selected a 317 set of models for which the \triangle AICc was below two (Burnham et al., 2011; Burnham and 318 Anderson, 2002). 319 Given that the selected models included the covariate year, we further explored the existence 320 of temporal variation in the annual FOI by assuming a constant FOI with age but estimating 321 different baseline FOI λ_0 for one, two or three periods of time to test our hypothesis. We 322 therefore assumed that the FOI could be considered as time-independent on average for 323 several years. This remains an approximation, but it allowed us to estimate temporal changes 324 in the FOI while staying in the framework of the general FOI model. 325 Because births in ibex take place between the end of May and the beginning of July, we considered FOI of year n to start from July 1^{st} year n-1 up to July 1^{st} year n. Based on age and 326 327 years of sampling, individuals were born between 1998 and 2017. Because the monitoring

and management interventions started in 2012, the years 1998-2012 were always included in
the first period. We compared all possible models with two or three periods, using all
possible threshold years or couple of threshold years between 2012 and 2018 to obtain
different periods.

332 For each explanatory variable of the final model, results were expressed as Hazard Ratios (HR). The hazard function is expressed as $\lambda = \lambda_0 e^{\beta_1 X_1 + \dots + \beta_n X_n}$. For a single qualitative 333 covariate X that is either present (X = 1) or absent (X = 0), the hazard (FOI) is $\lambda = \lambda_0 e^{\beta}$ for 334 X = 1, and $\lambda = \lambda_0$ for X = 0. Therefore, the hazard ratio is $HR = \lambda_0 e^{\beta} / \lambda_0 = e^{\beta}$. Similarly, 335 336 hazard ratios can be expressed for qualitative covariates with multiple categories, and for an 337 increase in one unit of a quantitative covariate. Hazard ratios have a similar interpretation as 338 odds ratios (Dohoo et al., 2009). Profile likelihood 95% confidence interval around each 339 estimated coefficient were computed (Therneau and Grambsch, 2000).

340

341 Estimating the FOI using longitudinal event time data

As we were interested in the time until the occurrence of infection ("survival time"), we analysed our longitudinal event time dataset using survival models. We note *T* the random variable representing the time between the first capture and the infection event, in days. The survival function is the probability that infection has not yet occurred by time *t* (follow up time): $Surv(t) = P(T \ge t)$. Note that in our data, animals were captured for the first time at different dates, meaning that the follow up time had different origins and therefore was not equivalent to the calendar time (Figure S1).

349 The survival function can be related to the hazard function $\lambda(t)$, i.e., the rate at which the

350 event occurs per unit time, at time *t*, using the following equation (Kleinbaum and Klein,

351 2005):

352 $Surv(t) = e^{-\int_0^t \lambda(u) \, du}$

353 In our case, the event of interest is infection, and therefore the hazard function is the FOI. 354 Note that if we assume a constant hazard risk (FOI) with follow up time, the hazard function is $\lambda(t) = \lambda$, and the corresponding survival function is $Surv(t) = e^{-\lambda t}$ (exponential model). 355 356 We tested the assumption that the hazard function is independent of age and calendar time, 357 using the exponential model from the function flexsurvreg of the *flexsurv* R package 358 (Jackson, 2016). We tested for the effects of age at last recapture (as a continuous variable, 359 calculated as the age at first capture incremented by the number of years between first capture 360 and last recapture) and year of recapture (as a categorical variable). Because of small sample 361 sizes, we had to group years 2014 and 2015 on the one hand and 2016 to 2018 on the other 362 hand. We were not able to take into account the effects of sex and socio-spatial unit because 363 of small sample sizes.

Given the results of the exponential model, we also explored the existence of variations with calendar time, fitting models assuming two periods between 2012 and 2018 characterised by two different FOI (independent of age and follow up time – Figure 2B). We fitted models considering two periods, using all possible monthly threshold dates between November 2012 and June 2018, and compared them to the model considering a constant FOI.

369

370 **RESULTS**

371 Force of infection estimated from cross-sectional age-seroprevalence data

372 Among the twelve diagnostic models taking into account the effect of SEX and UNIT, all

373 models with $\Delta AICc < 2$ included the variable year, which suggests that the assumption of a

time-independent FOI is not appropriate (Table 1; see also parameter estimates in Table S3).

375 Models 9, 11 and 12 also included an age effect (Table 1). However, the effect sizes of linear

376 or quadratic age were small, and adding these effects to model 3 including the variable year

did not significantly improve model fit (Likelihood Ratio Tests: p=0.10 and p=0.09,

378 respectively). For completeness, we fitted several models of age-varying FOI using a

379 piecewise constant hazards approach (models 13-20, Table S2). None of these models had a

380 better AICc than model 3, which does not support variation of the FOI with age.

381 Two models (4 and 12) with $\Delta AICc \le 2$ included infection-related mortality (Table 1). The

382 effect sizes of the infection-related mortality in models 4 and 12 were small (Table S3), and

adding infection-related mortality μ to model 3 or to model 11 did not significantly improve

model fit (Likelihood Ratio Test: *p*=0.13 and *p*=0.52, respectively).

Given that all models with $\Delta AICc < 2$ included the variable year, we further explored the

386 existence of temporal variation in the annual FOI by assuming a constant FOI with age and

387 no infection-related mortality, but estimating different baseline FOI λ_0 for one (as a reference

388 for no temporal variation), two or three periods of time.

389 The model with the lowest AICc (374.96; model 10 in Table S4) was obtained when

390 considering three periods, with 2012 and 2015 as threshold years (Figure 3A and Table S4),

and improved model fit compared to the model without temporal variation (AICc = 33.09,

 $\Delta AICc = 8.13$ with model 10). Baseline FOI estimates and hazard ratios for the effects of

393 SEX and UNIT are provided in Table S5: the FOI was highest in the second period, and

394 lowest in the third period; it was higher for females than males, and for the socio-spatial units

395 at the centre of the massif, where the seroprevalence levels are the highest. The average FOI

396 over the whole population (from the same model with three periods but without effects of

397 SEX and UNIT) was estimated to be 0.069 year^{-1} [0.046–0.097] for the period 1998-2012,

 0.115 year^{-1} [0.074–0.160] for the period 2012-2015 and 0.016 year⁻¹ [0.001–0.057] for the

399 period 2015-2018 (Figure 3B). A very similar model was obtained when considering three

400 periods but with 2012 and 2014 as threshold years (Tables S4 and S5, Figure S2).

401

402 Force of infection estimated from longitudinal event time data

403 Using the exponential model (constant baseline hazard) to fit our event time data collected on 404 marked animals, we found no significant effect of the variable AGE (HR_{per year} = 0.81 [0.54-405 1.20], p=0.28), but a significant effect of the variable YEAR (HR_{2014-2015 vs 2016-2018} = 0.095 406 [0.009-0.92], p=0.043).

407 These results did not support variation of the FOI with age. However, given the significance 408 of the variable YEAR, we further explored the existence of temporal variations in the FOI by 409 fitting a model with two different periods on the calendar time, where individuals were 410 submitted to different constant FOI before and after threshold dates. Among models 411 considering all possible monthly threshold dates between November 2012 and June 2018, the 412 best models ($\Delta AICc < 2$) were obtained when considering two periods with a threshold date 413 between July 2014 and December 2015 (Figure 3C). The model with the lowest AICc was 414 obtained when considering a threshold date in March 2015 (AICc = 30.72 - Figure 3C) and 415 improved the model fit compared to the simple exponential model without temporal 416 variations (AICc = 34.30, Δ AICc = 3.58). The FOI was estimated to be higher for the first period $(0.086 \text{ year}^{-1} [0.015 - 0.193])$ than for the second period (0 [0 - 0.030] - Figure 3D). 417

418

419 **DISCUSSION**

420 The aim of this study was to estimate the FOI of brucellosis in the wild Alpine ibex

421 population of the Bargy massif (France) between 2012 and 2018. This measure of

422 transmission intensity could help assess changes in transmission dynamics after management

423 interventions started in 2012, as a complement to existing seroprevalence studies (Calenge et

424 al., 2021; Marchand et al., 2017). Two massive culling operations were implemented in the

- 425 population in 2013 and 2015. In addition, test-and-remove operations were performed during
- 426 captures every year since 2012, with particularly high number of captures in 2015. We used
- 427 two different datasets: age-seroprevalence data obtained from culling or first captures

428 operations (n=363), and event time data, derived from recaptures of marked animals (n=50). 429 In both datasets, the model selection process supported an effect of time on the FOI, and 430 returned estimates supporting a potential decrease of the FOI between mid-2014 and late 431 2015. The force of infection was estimated to be reduced from 0.115 year⁻¹ [0.074–0.160] to 432 0.016 year⁻¹ [0.001–0.057] in the age-seroprevalence data.

433 Longitudinal samplings are difficult to obtain in wild populations and are therefore often not 434 available (Gilbert et al., 2013). In our case, data from recaptured animals were available, but 435 the sample size remained relatively low (only four seropositive among 50 recaptured 436 animals), leading to large confidence intervals in the estimates of the FOI (e.g. [0.015–0.193] 437 year⁻¹ for 2012-2015). Although a higher number of recaptures may have provided a more 438 complete dataset, it would have had negative impact in terms of disease management. The 439 total number of captures each year being constrained for logistical reasons, focusing captures 440 on already marked animals would have led to less animals being captured for the first time, 441 and thus less seropositive animals being removed from the population. When management 442 actions and disease surveillance both rely on the same field actions, a compromise has to be 443 found between both objectives that may lead to gather less relevant surveillance data. In such 444 cases where longitudinal data are absent or scarce because of practical or management 445 reasons, the derivation of the FOI from cross-sectional serological data, more frequently 446 collected in wildlife, is a useful complementary approach to gain insights into the 447 epidemiological dynamics as we illustrated in our study.

448

449 Model assumptions and infection-related mortality

In both FOI models of cross-sectional and longitudinal data, the main assumption is the
independence of the FOI with age. We did not find any support for age-dependency in our
datasets (Table 1).

Estimating the FOI from age-prevalence data also had the advantage of bringing insights on
brucellosis-related mortality, for which we did not find strong evidence (Table 1). Our results
did not detect brucellosis-induced mortality in ibex, similar to studies in bison (Fuller et al.,

456 2007; Joly and Messier, 2005) or elk (Benavides et al., 2017).

457

use of multiple serological assays, previous studies estimated the sensitivity of diagnosis
protocol to be around 95%, with perfect specificity (expert assumption in ANSES, 2015;
Lambert et al., 2020). Test failure thus does not appear to be a high source of bias, however,

We also postulated that the serological test has perfect sensitivity and specificity: due to the

461 in the case of lower quality tests, other modelling approaches could be investigated (Bollaerts

462 et al., 2012; Buzdugan et al., 2017). A few individuals in our datasets only had results from

the rapid LFIA test used in the field (see Material and Methods). The analyses produced

qualitatively and quantitatively similar results when using reduced datasets without theseindividuals (not shown).

466 In a former study, seroprevalence was found to decrease in old individuals, especially in 467 males (Marchand et al., 2017). It was hypothesized that this phenomenon could be explained 468 either by a lower survival rates or by lower probabilities to test positive in older individuals 469 infected by brucellosis (Marchand et al., 2017). We did not find strong evidence for 470 infection-related mortality, but we cannot exclude a decrease of antibody levels under 471 detectable levels as the animals age, which is difficult to reveal using seroprevalence data 472 (Heisey et al., 2006) (but see Benavides et al., 2017). Indeed, antibody titres measured by the 473 Complement Fixation Test appeared to decrease with age (Figure S3) (Lambert et al., 2018). 474 However, as high Complement Fixation Test titres are associated with active infections 475 (Durán-Ferrer et al., 2004), which are less often observed in old ibex (Lambert et al., 2018), 476 this may not reflect the antibody levels measured by the other serological tests used in the 477 study population. Unfortunately, the data for these other tests was not readily available.

Therefore, further studies investigating antibodies persistence with age or age-related
variation in test performance is needed to firmly conclude about these results. Moreover,
acquiring knowledge on antibody kinetics in ibex could have the additional advantage of
developing alternative approaches that improve the derivation of the FOI by estimating the
time since infection using individual data on antibody titres (Borremans et al., 2016; Pepin et
al., 2017).

Finally, we also assumed, as classically done in catalytic models, that all individuals are born
without infection. In domestic ruminants, congenital transmission of brucellosis exists but is

486 rare, i.e., in only ca. 5% of kids born from infectious mothers (FAO and WHO, 1986;

487 Godfroid et al., 2013). This transmission route is also suspected in ibex (Lambert et al.,

488 2018), but is probably as uncommon as in domestic ruminants. Moreover, individuals

489 congenitally infected seroconvert and become infectious only at the age of sexual maturity

490 (Plommet et al., 1973). Therefore, we assumed that the effects would be negligible on our

491 results.

492

493 **Temporal variation of the force of infection**

494 In Marchand et al. (2017), statistical analyses of seroprevalence data from 2012 to 2015 did 495 not detect changes in seroprevalence before and after the mass culling of autumn 2013. Using 496 a Bayesian approach to estimate seroprevalence, Calenge et al. (2021) found a strong 497 decrease in the overall seroprevalence between 2015 and 2016. In the present study, both 498 datasets suggest that the FOI decreased between mid-2014 and late 2015 (Figure 3), which 499 could explain the seroprevalence drop also observed. Our results support a decrease in 500 transmission during the study period, probably due to management interventions and natural 501 changes in the infection dynamics. Quantitatively, seroprevalence was halved between 2015

502 and 2016 (Calenge et al., 2021), while FOI as estimated using age-seroprevalence data was divided by approximately ten at the same period $(0.115 \text{ year}^{-1} \text{ vs. } 0.016 \text{ year}^{-1})$. 503 504 In the process of estimating the seroprevalence in marked animals, Calenge et al. (2021) also 505 estimated the FOI exerted on marked females in the core area, assuming it was proportional 506 to the proportion of seropositive unmarked ibex with an active infection among the 507 population of females in the core area. This analysis showed a decrease between 2015 and 508 2016, with estimates that were consistent with the values obtained with our longitudinal event 509 time dataset where we did not make such assumptions on the relation between the FOI and 510 active infections in the population (Calenge et al., 2021). Finally, an individual-based, 511 stochastic SEIR mechanistic model, also showed a decrease in incidence between 2013 and 512 2016 in the whole population, and a low increase of new cases between 2016 and 2018. 513 However, this increase in new cases was associated with a similar increase of population size 514 which could indicate a constant incidence rate (Lambert et al., 2020). Therefore, the temporal 515 dynamics appear to be consistent between the different approaches. The approach developed 516 here has the advantage of being much simpler than the mechanistic model, and of bringing 517 complementary information to previous seroprevalence analyses which can inform 518 management decisions. 519 The FOI did not decrease between 2012 and mid-2014, even though the population size was 520 halved due to massive culling operations in autumn 2013-spring 2014. On the contrary, our 521 results showed that the FOI increased in 2012, which could be related to management 522 interventions or undetermined natural changes in infection dynamics, such as abortion events 523 that play a critical role in brucellosis transmission. We also demonstrated a decrease in the 524 FOI in 2014 or 2015, which is probably related to management interventions. At the same 525 time, test-and-remove during captures replaced massive culling as the main component of management strategies. However, our analyses cannot disentangle the effects of each 526

527 management intervention that were implemented between 2012 and 2015, and the intense 528 test-and-remove operation of 2015 could have benefited from cumulative effects from the 529 previous years. These hypothesis could be tested in the future using the stochastic SEIR 530 mechanistic model developed in Lambert *et al.* (2020).

531 Unfortunately, we could not test for differences in temporal variation in the FOI between 532 socio-spatial units (interactions between calendar time and spatial unit), due to low sample 533 sizes. In a context where the FOI increases, such interactions could help to identify hotspots 534 of infection, i.e., spatial units where the FOI increases faster, or simply different course of 535 epidemics in the different spatial units (Heisey et al., 2010). Conversely, in a context of 536 decreasing FOI such as in our study, interactions between calendar time and spatial units 537 could reveal socio-spatial units with the sharpest decrease of FOI, which could correspond to 538 areas where the management interventions are the most effective. In particular, management 539 interventions since 2016 have been targeted towards the socio-spatial units at the centre of 540 the massif, where the seroprevalence levels (and the FOI as demonstrated here) are the 541 highest. It would therefore be interesting in the future to see if the FOI decreased faster in 542 those units, and to test whether the FOI keeps decreasing in the units with less management 543 interventions.

544

545 Force of infection in marked and unmarked animals

546 Our study aimed to compare the FOI between marked and unmarked animals. Most captures 547 targeted unmarked individuals to maximise chances to remove seropositive individuals 548 (Calenge et al., 2021). As a result, only a small number of marked animals were recaptured 549 and a few cases of seroconversion were observed, leading to high uncertainty around the 550 estimates of the FOI from the longitudinal event time dataset. Despite this uncertainty, the 551 estimates from both datasets are comparable in each period (2012-2015 and 2015-2018) and

552 support a potential decrease of the FOI of similar magnitude. These results indicate that the 553 FOI is similar in susceptible individuals from both groups, and therefore that marked 554 individuals have similar patterns of exposure as unmarked individuals. Assuming similar 555 FOI, a unified framework, using marked and unmarked animals, could be then further 556 investigated to gain in precision of FOI estimates, even with relatively small longitudinal 557 sample sizes, and better evaluate the requested sampling size of the monitoring (Gamble et 558 al., 2020). An integrated framework combining cross sectional and longitudinal capturerecapture setups would also decrease the needed sample size in comparison with separate 559 560 approaches (Gamble et al., 2020). It is worth noting that both our cross-sectional and 561 longitudinal datasets are interval censored event time dataset: from birth to the positive test 562 for the age-seroprevalence data or from negative to positive test for the longitudinal data. 563 Therefore, future research could use both datasets in the same analyses, which will allow to 564 increase the statistical power and to use more complex models of time-varying FOI.

565

566 **CONCLUSIONS**

567 In conclusion, using two distinct methods and datasets, our analyses revealed a decrease in 568 the FOI between mid-2014 and late 2015, possibly as a result of the successive management 569 operations implemented since 2012. Estimating the FOI can therefore provide valuable 570 insights on the epidemiological dynamics, as a complement to classical seroprevalence 571 analyses. This is especially the case in the context of ongoing disease management 572 interventions in wildlife populations, where the FOI is expected to change. Insights on the 573 effect of managing on transmission dynamics can be obtained using rather simple models as 574 we did here, provided that there is no strong evidence that the basic assumptions are violated. 575 Comparing two different sources of data, cross-sectional and cohort studies, with appropriate 576 models may contribute to provide robust inference about the FOI. Nonetheless, it is possible

to derive FOI estimates from cross-sectional serological data only, which are the mostcommonly available in wild populations.

579

580 Acknowledgments

581 The authors thank the two anonymous reviewers whose comments <u>helped improve and</u>

582 <u>clarify</u> this manuscript. The authors warmly thank all the field staff who performed

583 monitoring and management of the Alpine ibex population. They also thank the scientific

584 experts of the ANSES working groups on brucellosis of Alpine ibex in the Bargy massif, as

585 well as Charlotte Dunoyer (ANSES; UERSABA's chief unit) and Jean Hars (ONCFS). This

586 study was coordinated by the French Biodiversity Agency (Office Français de la Biodiversité,

587 OFB, formerly Hunting and Wildlife Agency, Office National de la Chasse et de la Faune

588 Sauvage, ONCFS). This work was performed within the framework of the LABEX

589 ECOFECT (ANR-11-LABX-0048) of Université de Lyon and of the program

590 "Investissements d'Avenir" (ANR- 11-IDEX-0007) operated by the French National

591 Research Agency (ANR).

592

593 Funding

594 This work was co-supported by the French Ministry of Research, the French Biodiversity

595 Agency (Office Français de la Biodiversité, OFB, formerly Hunting and Wildlife Agency,

596 Office National de la Chasse et de la Faune Sauvage, ONCFS), ANSES, and VetAgroSup

597 Lyon. The funders of the study had no role in the study design, the collection, analysis and598 interpretation of data, the writing of the report, or the decision to submit the article for

599 publication.

600

601 **Data availability statement:** The raw data required to reproduce the above findings are

602 available from the authors on reasonable request.

603

604 **Declarations of interest:** none

605 **CRediT author statement**

- 606 Sébastien Lambert: Conceptualisation, Methodology, Software, Formal Analysis, Data
- 607 Curation, Writing Original Draft, Writing Review & Editing, Visualisation. Emmanuelle
- 608 Gilot-Fromont, Anne Thébault: Conceptualisation, Methodology, Writing Review &
- 609 Editing, Supervision. Carole Toïgo, Sophie Rossi: Conceptualisation, Investigation, Writing
- 610 Review & Editing, Supervision. **Pascal Marchand, Elodie Petit:** Investigation, Data
- 611 Curation, Writing Review & Editing.
- 612

613 LITERATURE CITED

- ANSES, 2015. Mesures de maîtrise de la brucellose chez les bouquetins du Bargy. Rapport
 ANSES.
- ANSES, 2014. Note d'appui scientifique et technique de l'ANSES relatif au dépistage des
 bouquetins infectés de brucellose sur le terrain. Rapport ANSES.
- Artois, M., Bengis, R., Delahay, R.J., Duchêne, M.-J., Duff, J.P., Ferroglio, E., Gortázar, C.,
 Hutchings, M.R., Kock, R., Leighton, F.A., Mörner, T., Smith, G.C., 2009. Wildlife
 disease surveillance and monitoring, in: Management of Disease in Wild Mammals.
 pp. 187–214.
- Begon, M., Bennett, M., Bowers, R.G., French, N.P., Hazel, S.M., Turner, J., 2002. A
 clarification of transmission terms in host-microparasite models: numbers, densities
 and areas. Epidemiol. Infect. 129, 147–153.
- Benavides, J.A., Caillaud, D., Scurlock, B.M., Maichak, E.J., Edwards, W.H., Cross, P.C.,
 2017. Estimating loss of *Brucella abortus* antibodies from age-specific serological
 data in elk. EcoHealth 14, 234–243. https://doi.org/10.1007/s10393-017-1235-z
- Blasco, J.M., Garin-Bastuji, B., Marin, C.M., Gerbier, G., Fanlo, J., Jimenez de Bagues,
 M.P., Cau, C., 1994a. Efficacy of different Rose Bengal and complement fixation
 antigens for the diagnosis of *Brucella melitensis* infection in sheep and goats. Vet.
 Rec. 134, 415–420. https://doi.org/10.1136/vr.134.16.415
- Blasco, J.M., Marin, C.M., Jiménez-de-Bagués, M., Barberán, M., Hernández, A., Molina, L.,
 Velasco, J., Diaz, R., Moriyon, I., 1994b. Evaluation of allergic and serological tests
 for diagnosing *Brucella melitensis* infection in sheep. J. Clin. Microbiol. 32, 1835–
 1840.

- Bollaerts, K., Aerts, M., Shkedy, Z., Faes, C., Van der Stede, Y., Beutels, P., Hens, N., 2012.
 Estimating the population prevalence and force of infection directly from antibody
 titres. Stat. Modelling 12, 441–462. https://doi.org/10.1177/1471082X12457495
- Borremans, B., Hens, N., Beutels, P., Leirs, H., Reijniers, J., 2016. Estimating time of
 infection using prior derological and individual information can greatly improve
 incidence estimation of human and wildlife infections. PLOS Comput. Biol. 12,
 e1004882. https://doi.org/10.1371/journal.pcbi.1004882
- Burnham, K.P., Anderson, D.R., 2002. Model selection and multimodel inference: a practical
 information-theoretic approach, 2nd ed. ed. Springer, New York.
- Burnham, K.P., Anderson, D.R., Huyvaert, K.P., 2011. AIC model selection and multimodel
 inference in behavioral ecology: some background, observations, and comparisons.
 Behav. Ecol. Sociobiol. 65, 23–35. https://doi.org/10.1007/s00265-010-1029-6
- Buzdugan, S.N., Vergne, T., Grosbois, V., Delahay, R.J., Drewe, J.A., 2017. Inference of the
 infection status of individuals using longitudinal testing data from cryptic
 populations: towards a probabilistic approach to diagnosis. Sci. Rep. 7, 1–11.
 https://doi.org/10.1038/s41598-017-00806-4
- Calenge, C., Lambert, S., Petit, E., Thébault, A., Gilot-Fromont, E., Toïgo, C., Rossi, S.,
 2021. Estimating disease prevalence and temporal dynamics using biased capture
 serological data in a wildlife reservoir: the example of brucellosis in Alpine ibex
 (*Capra ibex*). Prev. Vet. Med. 187, 1–12.
- 656 https://doi.org/10.1016/j.prevetmed.2020.105239
- 657 Carvalho Neta, A.V., Mol, J.P.S., Xavier, M.N., Paixão, T.A., Lage, A.P., Santos, R.L., 2010.
 658 Pathogenesis of bovine brucellosis. Vet. J. 184, 146–155.
 659 https://doi.org/10.1016/j.tvjl.2009.04.010
- Corde, Y., Drapeau, A., Game, Y., Maucci, E., Hars, J., Jaÿ, M., Garin-Bastuji, B., 2014. A
 rapid test for evaluating *B. melitensis* infection prevalence in an Alpine ibex (*Capra ibex*) reservoir in the French Alps. Presented at the Brucellosis 2014 International
 Research Conference, including the 67th Annual Brucellosis Research Meeting,
 Berlin, Germany, p. 221.
- Courtejoie, N., Salje, H., Durand, B., Zanella, G., Cauchemez, S., 2018. Using serological
 studies to reconstruct the history of bluetongue epidemic in French cattle under
 successive vaccination campaigns. Epidemics 25, 54–60.
 https://doi.org/10.1016/j.epidem.2018.05.005
- Dohoo, I., Martin, W., Stryhn, H., 2009. Veterinary epidemiologic research, 2. ed. ed. VER
 Inc, Charlottetown, Canada: University of Prince Edward Island.
- Durán-Ferrer, M., Léon, L., Nielsen, K., Caporale, V., Mendoza, J., Osuna, A., Perales, A.,
 Smith, P., De-Frutos, C., Gómez-Martín, B., Lucas, A., Chico, R., Delgado, O.D.,
 Escabias, J.C., Arrogante, L., Díaz-Parra, R., Garrido, F., 2004. Antibody response
 and antigen-specific gamma-interferon profiles of vaccinated and unvaccinated
 pregnant sheep experimentally infected with *Brucella melitensis*. Vet. Microbiol. 100,
 219–231. https://doi.org/10.1016/j.vetmic.2004.02.008
- Ebinger, M., Cross, P., Wallen, R., White, P.J., Treanor, J., 2011. Simulating sterilization,
 vaccination, and test-and-remove as brucellosis control measures in bison. Ecol. Appl.
 21, 2944–2959. https://doi.org/10.1890/10-2239.1
- European Commission, 2001. Brucellosis in sheep and goats (*Brucella melitensis*) (No.
 SANCO.C.2/AH/R23/2001). Health & Consumer Protection Directorate-General,
 Brussels, Belgium.
- Farrington, C.P., 1990. Modelling forces of infection for measles, mumps and rubella. Stat.
 Med. 9, 953–967. https://doi.org/10.1002/sim.4780090811

- Ferreira, A.C., Cardoso, R., Travassos Dias, I., Mariano, I., Belo, A., Rolão Preto, I.,
 Manteigas, A., Pina Fonseca, A., Corrêa De Sá, M.I., 2003. Evaluation of a modified
 Rose Bengal test and an indirect Enzyme-Linked Immunosorbent Assay for the
 diagnosis of *Brucella melitensis* infection in sheep. Vet. Res. 34, 297–305.
 https://doi.org/10.1051/vetres:2003005
- Forbes, L.B., Tessaro, S.V., Lees, W., 1996. Experimental studies on <i>Brucella abortus<i
 in moose (*Alces alces*). J. Wildl. Dis. 32, 94–104. https://doi.org/10.7589/0090-355832.1.94
- Freycon, P., Game, Y., Hars, J., Gilot-Fromont, E., 2017. Lesional aspects of *Brucella melitensis* in *Capra ibex*. Bull. Acad. Vét. France 170, 1–5.
 https://doi.org/10.4267/2042/62276
- Fuller, J.A., Garrott, R.A., White, P.J., Aune, K.E., Roffe, T.J., Rhyan, J.C., 2007.
 Reproduction and survival of Yellowstone bison. J. Wildl. Manage. 71, 2365–2372.
 https://doi.org/10.2193/2006-201
- 699 Gamble, A., Garnier, R., Chambert, T., Gimenez, O., Boulinier, T., 2020. Next-generation
 700 serology: integrating cross-sectional and capture–recapture approaches to infer
 701 disease dynamics. Ecology 101, e02923. https://doi.org/10.1002/ecy.2923
- Garin-Bastuji, B., Hars, J., Drapeau, A., Cherfa, M.-A., Game, Y., Le Horgne, J.M.,
 Rautureau, S., Maucci, E., Pasquier, J.J., Jaÿ, M., Mick, V., 2014. Reemergence of *Brucella melitensis* in wildlife, France. Emerg. Infect. Dis 20, 1570–1571.
 https://doi.org/10.3201/eid2009.131517
- Gilbert, A.T., Fooks, A.R., Hayman, D.T.S., Horton, D.L., Müller, T., Plowright, R., Peel,
 A.J., Bowen, R., Wood, J.L.N., Mills, J., Cunningham, A.A., Rupprecht, C.E., 2013.
 Deciphering serology to understand the ecology of infectious diseases in wildlife.
 EcoHealth 10, 298–313. https://doi.org/10.1007/s10393-013-0856-0
- Godfroid, J., Nielsen, K., Saegerman, C., 2010. Diagnosis of brucellosis in livestock and
 wildlife. Croat. Med. J. 51, 296–305. https://doi.org/10.3325/cmj.2010.51.296
- Gorsich, E.E., Ezenwa, V.O., Cross, P.C., Bengis, R.G., Jolles, A.E., 2015. Contextdependent survival, fecundity and predicted population-level consequences of
 brucellosis in African buffalo. J. Anim. Ecol. 84, 999–1009.
 https://doi.org/10.1111/1365-2656.12356
- Hars, J., Rautureau, S., Jaÿ, M., Game, Y., Gauthier, D., Herbaux, J.P., Le Horgne, J.M.,
 Maucci, E., Pasquier, J.J., Vaniscotte, A., Mick, V., Garin-Bastuji, B., 2013. Un foyer
 de brucellose chez les ongulés sauvages du massif du Bargy en Haute-Savoie. Bull.
 Epidémiol. Santé Anim. Alim. 60, 2–7.
- Heisey, D.M., Joly, D.O., Messier, F., 2006. The fitting of general force-of-infection models
 to wildlife disease prevalence data. Ecology 87, 2356–2365.
 https://doi.org/10.1890/0012-9658(2006)87[2356:TFOGFM]2.0.CO;2
- Heisey, D.M., Osnas, E.E., Cross, P.C., Joly, D.O., Langenberg, J.A., Miller, M.W., 2010.
 Linking process to pattern: estimating spatiotemporal dynamics of a wildlife epidemic from cross-sectional data. Ecol. Monogr. 80, 221–240. https://doi.org/10.1890/09-0052.1
- Hens, N., Aerts, M., Faes, C., Shkedy, Z., Lejeune, O., Van Damme, P., Beutels, P., 2010.
 Seventy-five years of estimating the force of infection from current status data.
 Epidemiol. Infect. 138, 802–812. https://doi.org/10.1017/S0950268809990781
- Jackson, R., 2016. Flexsurv: a platform for parametric survival modeling in R. J. Stat. Softw.
 70, 1–33. https://doi.org/10.18637/jss.v070.i08
- Joly, D.O., Messier, F., 2005. The effect of bovine tuberculosis and brucellosis on
 reproduction and survival of wood bison in Wood Buffalo National Park. J. Anim.
 Ecol. 74, 543–551. https://doi.org/10.1111/j.1365-2656.2005.00953.x

735 Joseph, M.B., Mihaljevic, J.R., Arellano, A.L., Kueneman, J.G., Preston, D.L., Cross, P.C., 736 Johnson, P.T.J., 2013. Taming wildlife disease: bridging the gap between science and management. J. Appl. Ecol. 50, 702-712. https://doi.org/10.1111/1365-2664.12084 737 738 Katzelnick, L.C., Ben-Shachar, R., Mercado, J.C., Rodriguez-Barraquer, I., Elizondo, D., 739 Arguello, S., Nuñez, A., Ojeda, S., Sanchez, N., Mercado, B.L., Gresh, L., Burger-740 Calderon, R., Kuan, G., Gordon, A., Balmaseda, A., Harris, E., 2018. Dynamics and 741 determinants of the force of infection of dengue virus from 1994 to 2015 in Managua, 742 Nicaragua. PNAS 115, 10762-10767. https://doi.org/10.1073/pnas.1809253115 743 Keiding, N., 1991. Age-specific incidence and prevalence: a statistical perspective. J. R. Stat. 744 Soc. Ser. A Stat. Soc. 154, 371-412. https://doi.org/10.2307/2983150 745 Kleinbaum, D.G., Klein, M., 2005. Survival analysis: a self-learning text, 2. ed. ed, Statistics 746 for biology and health. Springer, New York, NY. 747 Lambert, S., Gilot-Fromont, E., Freycon, P., Thébault, A., Game, Y., Toïgo, C., Petit, E., 748 Barthe, M.-N., Reynaud, G., Jaÿ, M., Garin-Bastuji, B., Ponsart, C., Hars, J., Rossi, 749 S., 2018. High shedding potential and significant individual heterogeneity in 750 naturally-infected Alpine ibex (Capra ibex) with Brucella melitensis. Front. 751 Microbiol. 9, 1-15. https://doi.org/10.3389/fmicb.2018.01065 752 Lambert, S., Gilot-Fromont, E., Toïgo, C., Marchand, P., Petit, E., Garin-Bastuji, B., 753 Gauthier, D., Gaillard, J.-M., Rossi, S., Thébault, A., 2020. An individual-based 754 model to assess the spatial and individual heterogeneity of Brucella melitensis transmission in Alpine ibex. Ecol. Modell. 425, 109009. 755 756 https://doi.org/10.1016/j.ecolmodel.2020.109009 757 le Roex, N., Cooper, D., van Helden, P.D., Hoal, E.G., Jolles, A.E., 2015. Disease control in 758 wildlife: evaluating a test and cull programme for bovine tuberculosis in African 759 buffalo. Transbound. Emerg. Dis. [Epub ahead of print]. 760 https://doi.org/10.1111/tbed.12329 Lewis, F.I., Otero-Abad, B., Hegglin, D., Deplazes, P., Torgerson, P.R., 2014. Dynamics of 761 762 the force of infection: insights from Echinococcus multilocularis infection in foxes. 763 PLoS Negl. Trop. Dis. 8, e2731. https://doi.org/10.1371/journal.pntd.0002731 Mailles, A., Rautureau, S., Le Horgne, J.M., Poignet-Leroux, B., d'Arnoux, C., Dennetière, 764 765 G., Faure, M., Lavigne, J.P., Bru, J.P., Garin-Bastuji, B., 2012. Re-emergence of 766 brucellosis in cattle in France and risk for human health. Euro Surveill. 17, 1–3. 767 Marchand, P., Freycon, P., Herbaux, J.P., Game, Y., Toïgo, C., Gilot-Fromont, E., Rossi, S., 768 Hars, J., 2017. Sociospatial structure explains marked variation in brucellosis 769 seroprevalence in an Alpine ibex population. Sci. Rep. 7, 15592. https://doi.org/10.1038/s41598-017-15803-w 770 771 Mazerolle, M.J., 2019. AICcmodavg: model selection and multimodel inference based on 772 (Q)AIC(c). R package version 2.2-2. 773 McDonald, R.A., Delahay, R.J., Carter, S.P., Smith, G.C., Cheeseman, C.L., 2008. Perturbing 774 implications of wildlife ecology for disease control. Trends Ecol. Evol. 23, 53–56. 775 https://doi.org/10.1016/j.tree.2007.10.011 776 Michallet, J., Grand, B., Bonardi, J., 1988. La population de bouquetins des Alpes du massif 777 de Belledonne-Sept Laux (département de l'Isère). Bull. Mens. ONC 19-24. 778 Mick, V., Le Carrou, G., Corde, Y., Game, Y., Jaÿ, M., Garin-Bastuji, B., 2014. Brucella melitensis in France: persistence in wildlife and probable spillover from Alpine ibex 779 780 to domestic animals. PLoS ONE 9, e94168. 781 https://doi.org/10.1371/journal.pone.0094168 Muench, H., 1934. Derivation of rates from summation data by the catalytic curve. J. Am. 782 783 Stat. Assoc. 29, 25–38. https://doi.org/10.2307/2278457

Müller, T., Freuling, C.M., Wysocki, P., Roumiantzeff, M., Freney, J., Mettenleiter, T.C., 784 785 Vos, A., 2015. Terrestrial rabies control in the European Union: historical 786 achievements and challenges ahead. Vet. J. 203, 10-17. 787 https://doi.org/10.1016/j.tvjl.2014.10.026 788 Mysterud, A., Rolandsen, C.M., 2019. Fencing for wildlife disease control. J. Appl. Ecol. 56, 519-525. https://doi.org/10.1111/1365-2664.13301 789 790 Mysterud, A., Rolandsen, C.M., 2018. A reindeer cull to prevent chronic wasting disease in 791 Europe. Nat. Ecol. Evol. 1-3. https://doi.org/10.1038/s41559-018-0616-1 792 Nielsen, K., Gall, D., Smith, P., Balsevicius, S., Garrido, F., Durán-Ferrer, M., Biancifiori, F., 793 Dajer, A., Luna, E., Samartino, L., Bermudez, R., Moreno, F., Renteria, T., Corral, 794 A., 2004. Comparison of serological tests for the detection of ovine and caprine 795 antibody to Brucella melitensis. Rev. Sci. Tech. Off. Int. Epizoot. 23, 979-987. 796 Papoz, L., Simondon, F., Saurin, W., Sarmini, H., 1986. A simple model relevant to 797 toxoplasmosis applied to epidemiologic results in France. Am. J. Epidemiol. 123, 154-161. https://doi.org/10.1093/oxfordjournals.aje.a114210 798 799 Pepin, K.M., Kay, S.L., Golas, B.D., Shriner, S.S., Gilbert, A.T., Miller, R.S., Graham, A.L., 800 Riley, S., Cross, P.C., Samuel, M.D., Hooten, M.B., Hoeting, J.A., Lloyd-Smith, J.O., 801 Webb, C.T., Buhnerkempe, M.G., 2017. Inferring infection hazard in wildlife 802 populations by linking data across individual and population scales. Ecol. Lett. 20, 803 275-292. https://doi.org/10.1111/ele.12732 804 Perrin, J.-B., Rautureau, S., Bronner, A., Hosteing, S., Dufour, B., Garin-Bastuji, B., Jaÿ, M., 805 2016a. Absence of bovine brucellosis confirmed in 2014, but vigilance must be 806 maintained. Bull. Epidémiol. Santé Anim. Alim. 71, 12-16. 807 Perrin, J.-B., Rautureau, S., Bronner, A., Hosteing, S., Jaÿ, M., Garin-Bastuji, B., Dufour, B., 2016b. Brucellosis in small ruminants in 2014: 95 départements of metropolitan 808 809 France are now officially disease-free. Bull. Epidémiol. Santé Anim. Alim. 71, 17–21. Reynolds, J.J.H., Carver, S., Cunningham, M.W., Logan, K.A., Vickers, W., Crooks, K.R., 810 811 VandeWoude, S., Craft, M.E., 2019. Feline immunodeficiency virus in puma: 812 estimation of force of infection reveals insights into transmission. Ecol. Evol. 9, 11010-11024. https://doi.org/10.1002/ece3.5584 813 814 Schlafer, D.H., Miller, R.B., 2007. Female genital system, in: Maxie, M.G. (Ed.), Jubb, 815 Kennedy & Palmer's Pathology of Domestic Animals (Fifth Edition). Elsevier, Edinburgh, pp. 429-431. https://doi.org/10.1016/B978-070202823-6.50174-3 816 Therneau, T.M., Grambsch, P.M., 2000. Modeling survival data: extending the Cox model, 817 818 Statistics for biology and health. Springer, New York. 819 Treanor, J.J., Geremia, C., Crowley, P.H., Cox, J.J., White, P.J., Wallen, R.L., Blanton, D.W., 2011. Estimating probabilities of active brucellosis infection in Yellowstone 820 821 bison through quantitative serology and tissue culture. J. Appl. Ecol. 48, 1324–1332. 822 https://doi.org/10.1111/j.1365-2664.2011.02058.x 823 Vicente, J., Apollonio, M., Blanco-Aguiar, J.A., Borowik, T., Brivio, F., Casaer, J., Croft, S., 824 Ericsson, G., Ferroglio, E., Gavier-Widen, D., Gortázar, C., Jansen, P.A., Keuling, O., 825 Kowalczyk, R., Petrovic, K., Plhal, R., Podgórski, T., Sange, M., Scandura, M., 826 Schmidt, K., Smith, G.C., Soriguer, R., Thulke, H.H., Zanet, S., Acevedo, P., 2019. 827 Science-based wildlife disease response. Science 364, 943.2-944. 828 https://doi.org/10.1126/science.aax4310 829 Vynnycky, E., White, R.G., 2010. An introduction to infectious disease modelling. Oxford 830 University Press, New York. 831 Wagenmakers, E.J., Farrell, S., 2004. AIC model selection using Akaike weights. Psychon. 832 Bull. Rev. 11, 192–196. https://doi.org/10.3758/BF03206482 833

Table 1: Model selection table to analyse the age-seroprevalence data using a constant force of infection with age and time, and testing for the effect of infection-related mortality μ , year

| | | - | - | | |
|-----|-------------------|----------------------|---------------------|-----------------------------------|---------|
| 836 | and age (linear o | or quadratic), takin | ng into account the | e effect of sex and socio-spatial | l unit. |

| N° | Year | Age | Age ² | μ | Unit | Sex | DF | LL | AICc | ΔAICc | W |
|----|------|-----|------------------|---|------|-----|----|---------|--------|-------|------|
| 11 | + | | + | | + | + | 12 | -176.91 | 378.73 | 0.00 | 0.20 |
| 9 | + | + | | | + | + | 12 | -176.99 | 378.89 | 0.16 | 0.19 |
| 4 | + | | | + | + | + | 12 | -177.16 | 379.24 | 0.50 | 0.16 |
| 3 | + | | | | + | + | 11 | -178.31 | 379.38 | 0.65 | 0.15 |
| 12 | + | | + | + | + | + | 13 | -176.71 | 380.48 | 1.75 | 0.09 |
| 10 | + | + | | + | + | + | 13 | -176.99 | 381.04 | 2.31 | 0.06 |
| 7 | | | + | | + | + | 7 | -183.68 | 381.69 | 2.96 | 0.05 |
| 5 | | + | | | + | + | 7 | -184.20 | 382.73 | 3.99 | 0.03 |
| 2 | | | | + | + | + | 7 | -184.34 | 383.01 | 4.27 | 0.02 |
| 1 | | | | | + | + | 6 | -185.42 | 383.09 | 4.35 | 0.02 |
| 8 | | | + | + | + | + | 8 | -183.60 | 383.62 | 4.88 | 0.02 |
| 6 | | + | | + | + | + | 8 | -184.20 | 384.82 | 6.09 | 0.01 |

For each model, the table gives the number of Degrees of Freedom (DF), the Log-Likelihood (LL),
the Akaike Information Criterion corrected for small sample sizes (AICc), the difference between their

839 AICc and the lowest AICc value (Δ AICc), and the Akaike Weights (W), which are relative model 840 likelihoods normalized over the likelihoods of all possible submodels. The models with Δ AICc<2 are 841 in bold.

843 Figure legends

844

845 Figure 1: Timing of the monitoring and management operations during which ibex of

846 the Bargy massif (France) were sampled for serological tests between 2012 and 2018.

847 In green: months during which captures were performed; in yellow: months during which

recaptures were performed; in blue: months during which culling operations were performed.

849 The number of individuals captured, recaptured or culled is indicated for each operation.

850

Figure 2: Serological datasets for brucellosis between 2012 and 2018 in the Bargy massif (France)

853 (A) Age-seroprevalence data, for ibex culled or captured for the first time, with black points

representing the observed seroprevalence for each age, with the size of the points

proportional to the sample taken (n=363). (B) Distribution of the duration between the first

856 capture (where the individuals were seronegative, marked and released) and the last

- recapture, where the ibex either tested seronegative, or converted to seropositive (the event of
- 858 interest), for ibex that were recaptured at least once (n=50). (B) Dates of first capture and last
- recapture of marked individuals, for ibex that were recaptured at least once (n=50). The solid
- horizontal lines represent the life of marked animals between the date of their first capture,
- 861 where they tested seronegative (white dots), and their last recapture, where they either tested
- 862 seronegative or seropositive (black dots). The dashed vertical line represents a possible
- threshold date that separates the first period where individuals are exposed to a certain force of infection (FOI 1) from the second period where individuals are exposed to a different force
- 865 of infection (FOI 1) from the se
- 866

867 Figure 3: Estimates of time-varying forces of infection.

868 (A) AICc of the models considering different forces of infection for three periods on the 869 calendar time for the cross-sectional age-seroprevalence dataset, comparing all possible pairs

- of threshold years between 2012 and 2017 to divide the 1998-2018 period into three. (C)
- AICc of the models considering different forces of infection for two periods on the calendar
- time for the longitudinal event time dataset, comparing all possible threshold months to
- divide the 2012-2018 period into two. In (A) and (C), the grey areas represents the AICc
- values where the difference between the AICc and the lowest AICc value (Δ AICc) were
- below two. (B) and (D) Estimates of the FOI (year⁻¹) for the best model considering time-
- 876 varying forces of infection for the age-seroprevalence and the event time datasets,
- 877 respectively, without effects of sex and unit. The lines represent the means, and the areas
- 878 represent the 95% confidence intervals. The dashed vertical lines represent the threshold
- ates that separate the different periods corresponding to different forces of infection in the
- 880 models with the lowest AICc values.
- 881





