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Development of a primary cell model derived from porcine dorsal soft palate for foot-and-mouth disease virus research and diagnosis.

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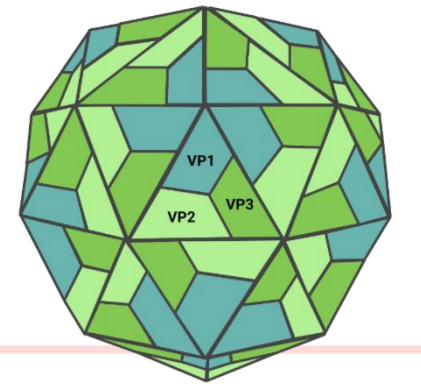
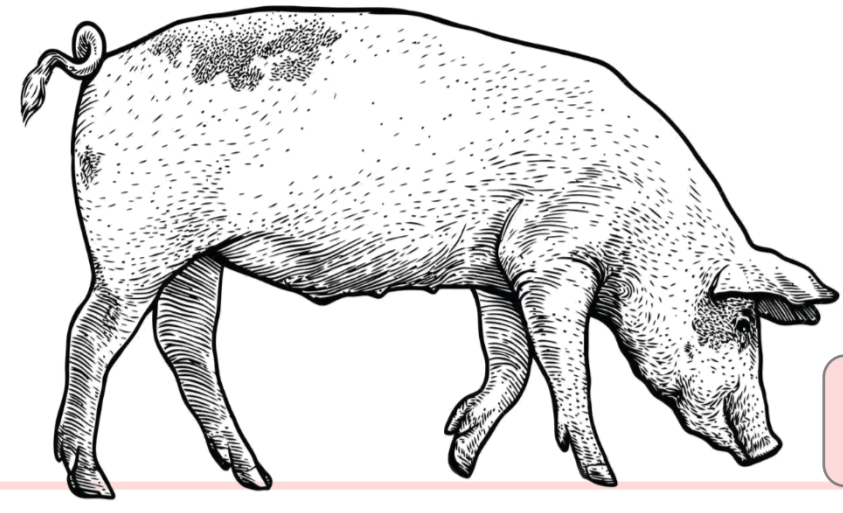
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INTRODUCTION & OBJECTIVES

- Foot-and-mouth disease is a highly contagious disease affecting wild and domestic artiodactyls, caused by a Picornavirus named Foot-and-mouth disease virus (FMDV).
- FMDV is able to persist (i.e. infectious virus detected after 28 days post-infection (dpi)) for several years in the pharyngeal epithelium of ruminants after resolution of clinical signs¹. However, this is not the case in pigs where only viral RNA could be detected in vivo for more than 60 dpi in the oropharyngeal tonsils and dorsal soft palate^{2,3}.
- No porcine cell model from a tissue of interest is currently available.

➤ **Aim of the study:** To develop a specific cellular model to study FMDV infection in porcine cells.

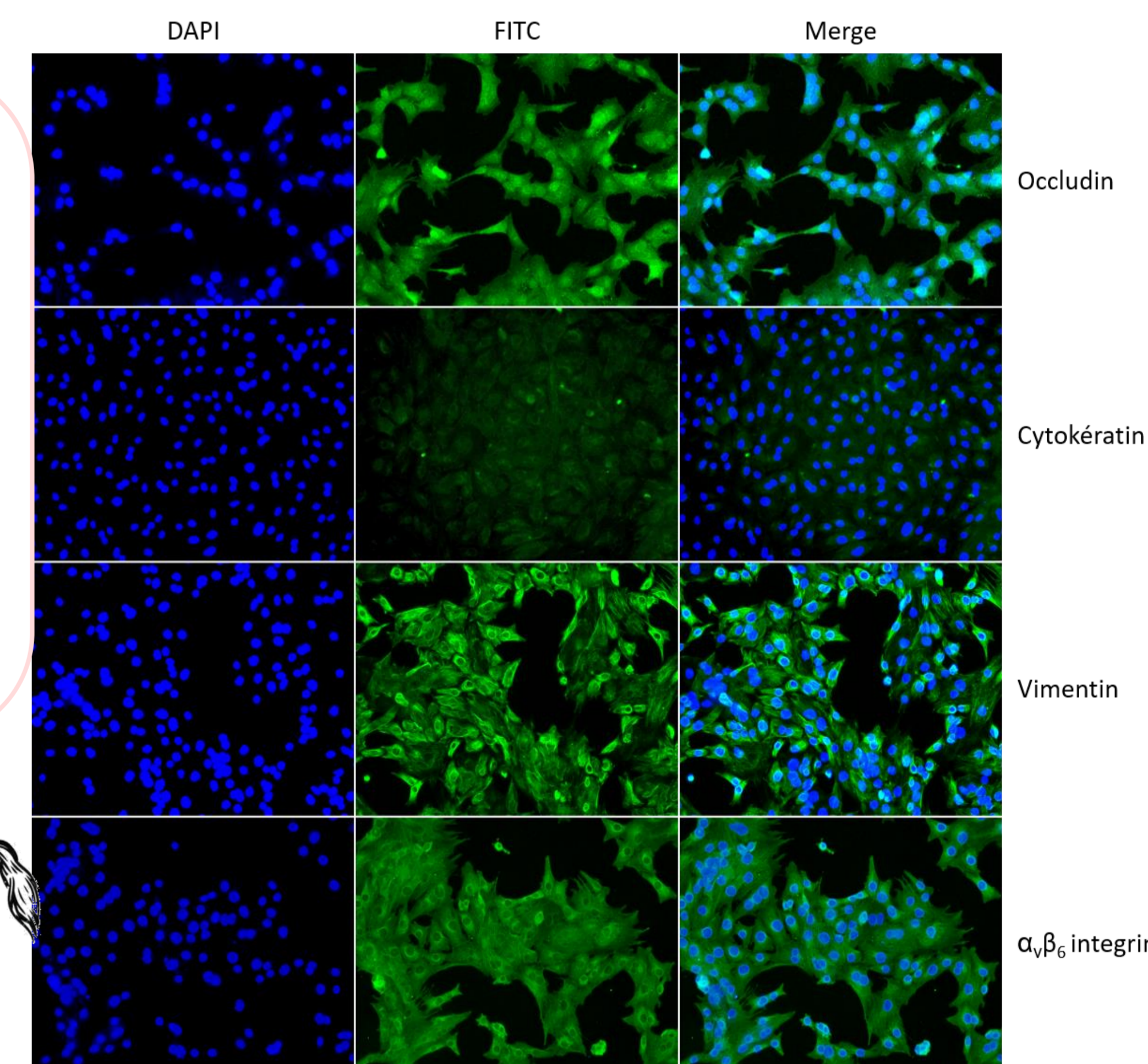
Workflow

- Swine oropharyngeal tonsils and dorsal soft palate (DSP) were sampled.
- Epithelial structures were identified and isolated ①.
- Epithelial cells derived from DSP were propagated in monolayers, characterized ②, then infected with FMDV ③.
- Epithelial cells were then grown in multilayers at the air-liquid interphase to mimic a pluristratified epithelium ④.

② DSP cells characterization

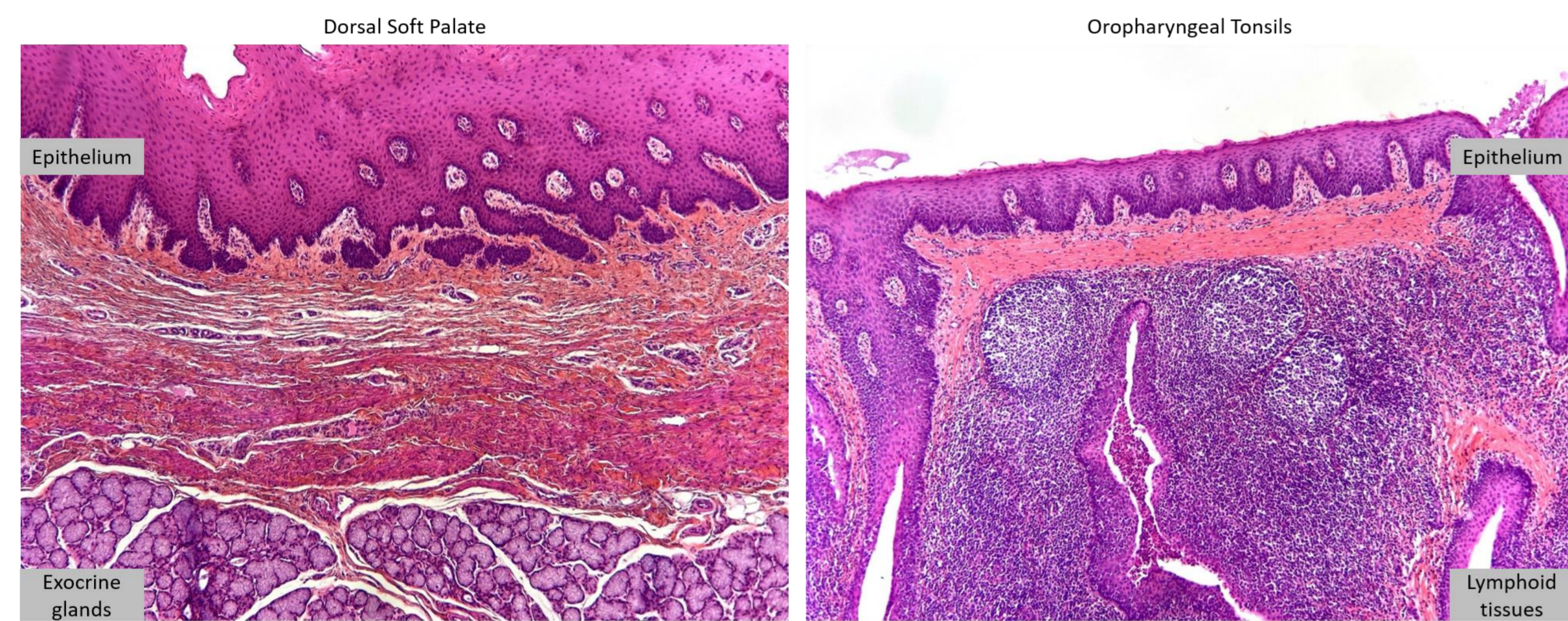
- DSP cells were cultured in monolayers, fixed, then labelled with antibodies targeting epithelial markers (occludin, cytokeratin), fibroblastic markers (vimentin) and FMDV receptor ($\alpha_v\beta_6$ integrin).

- Significant expression of occludin was detected, indicating an epithelial nature.
- Greater expression of vimentin than cytokeratin was detected, indicating an epithelial-mesenchymal transition.
- High expression of the major receptor for FMDV was detected.



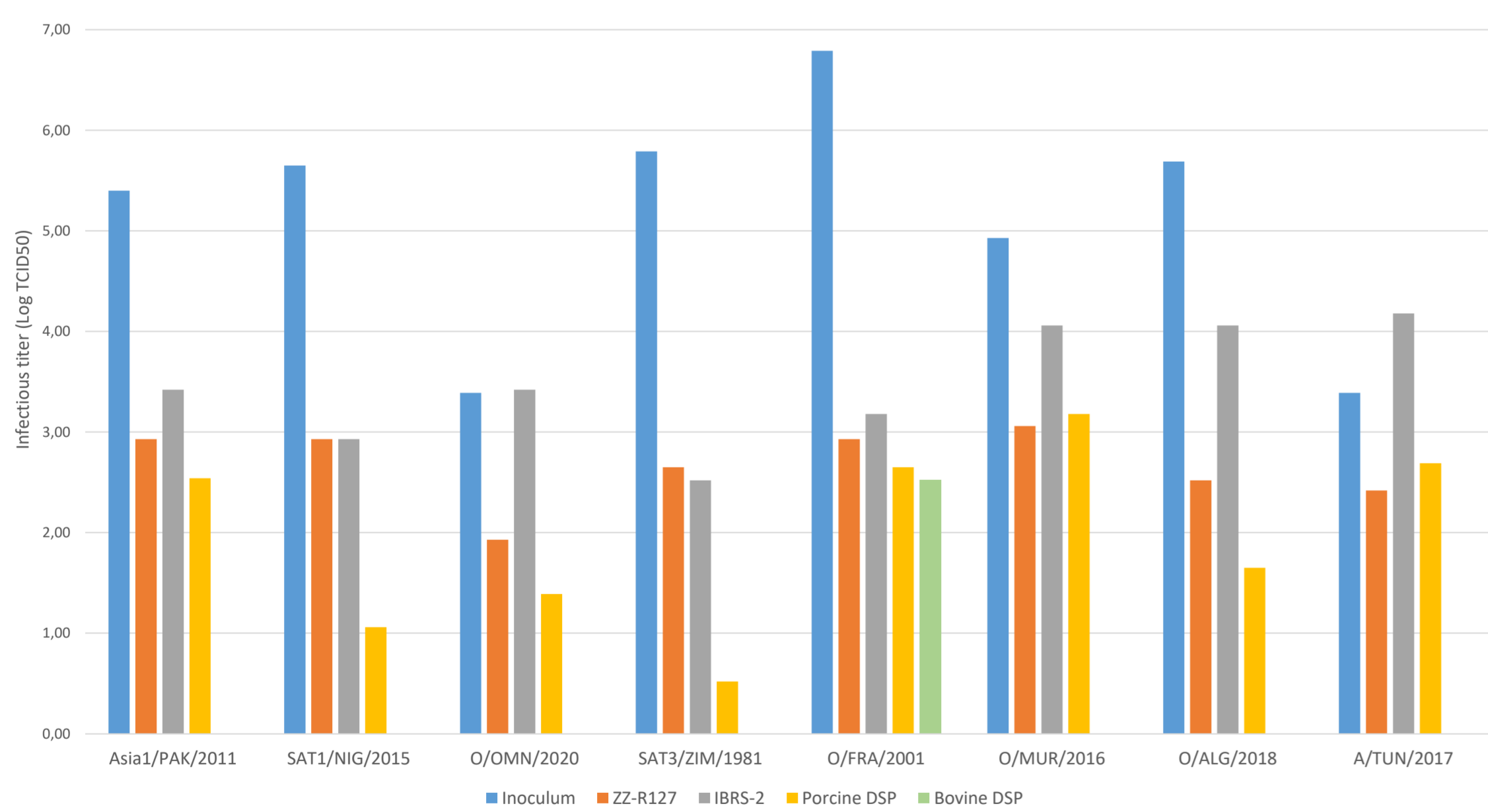
① Epithelial structures characterization

- Oropharyngeal tonsils and DSP samples were stained using HES (hematoxylin-eosin-saffron) in order to localize epithelial structures.
- Epithelium were identified in both oropharyngeal tonsils and DSP samples



③ Assessment of DSP susceptibility to FMDV infection

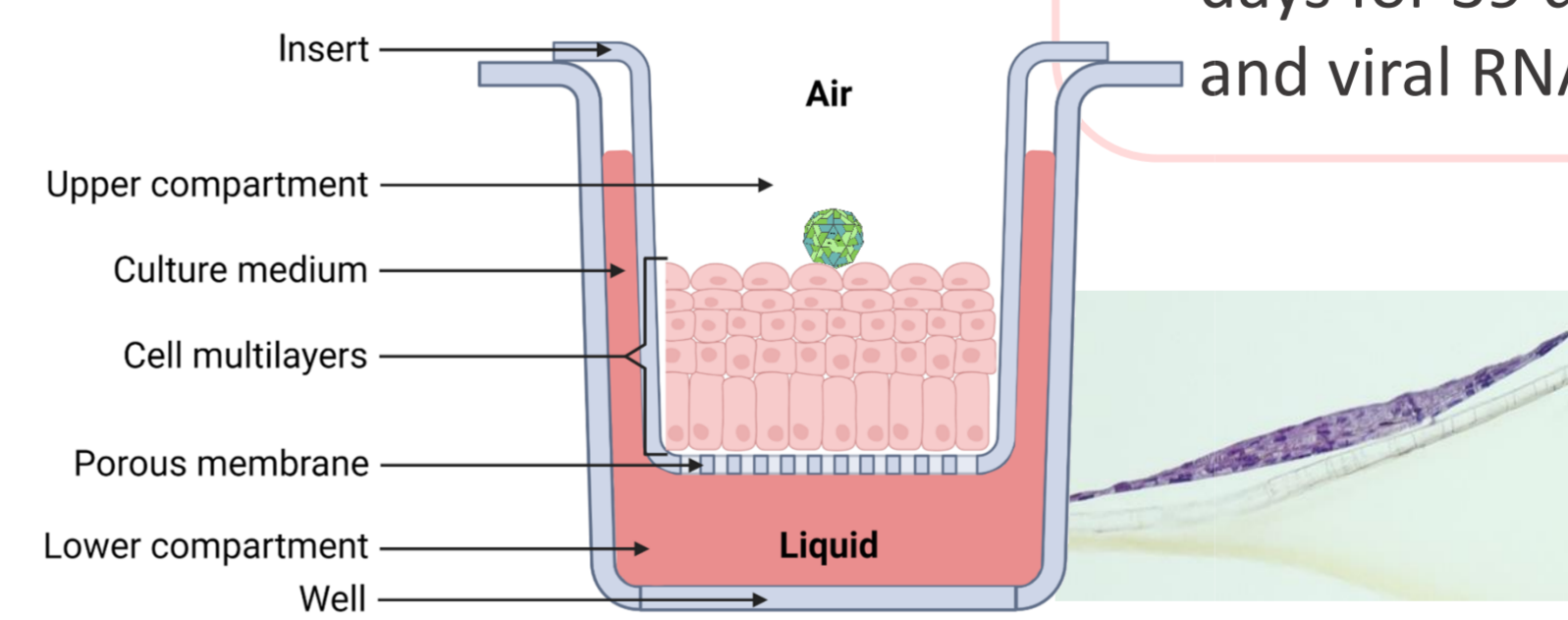
- DSP cells were infected with a panel of FMDV strains in order to compare their susceptibility to other cell lines : ZZ-R127, IBRS-2 and bovine DSP.
- Culture supernatants were collected 48 hpi in order to perform viral titration on BHK-21 cells.



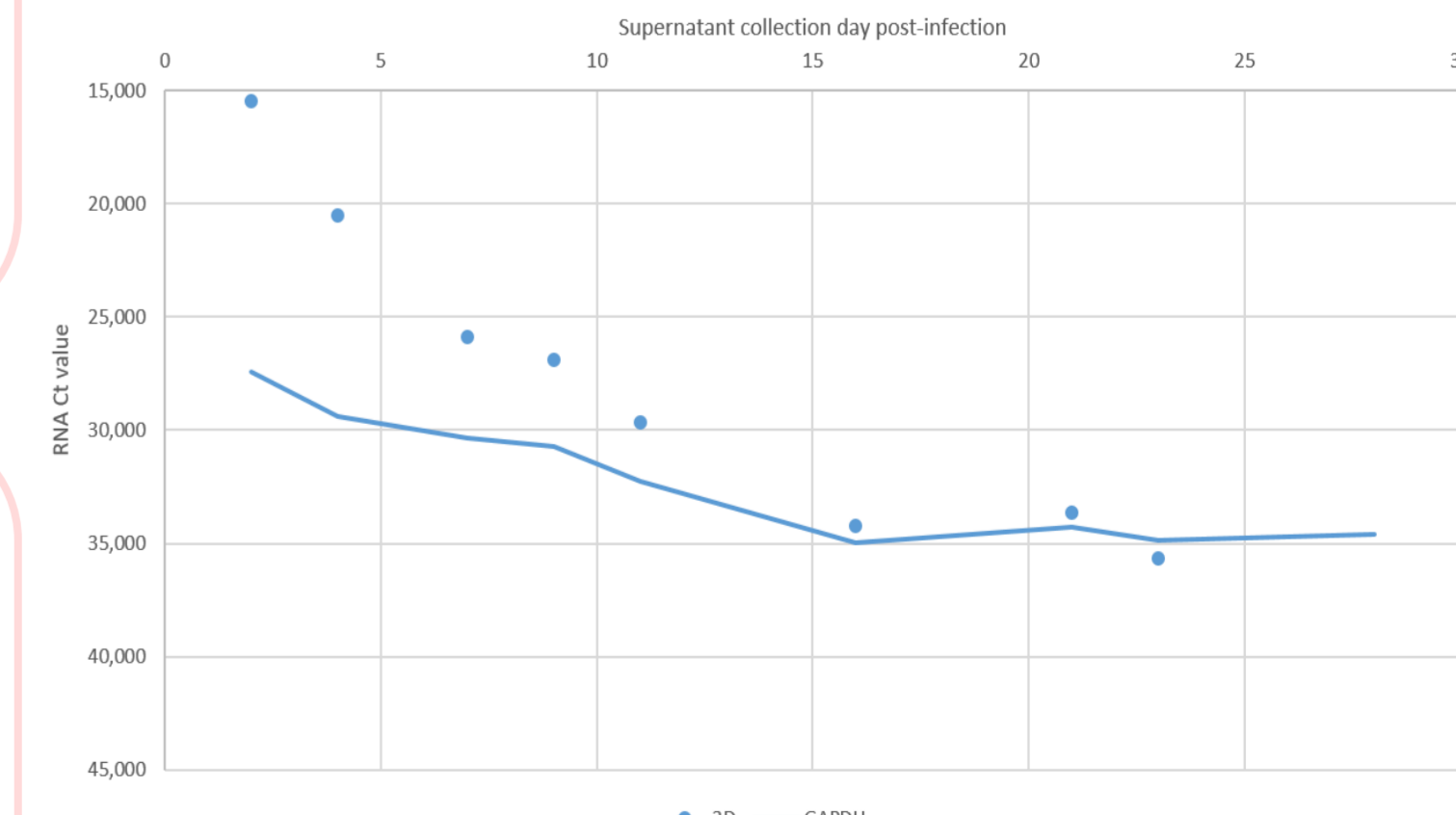
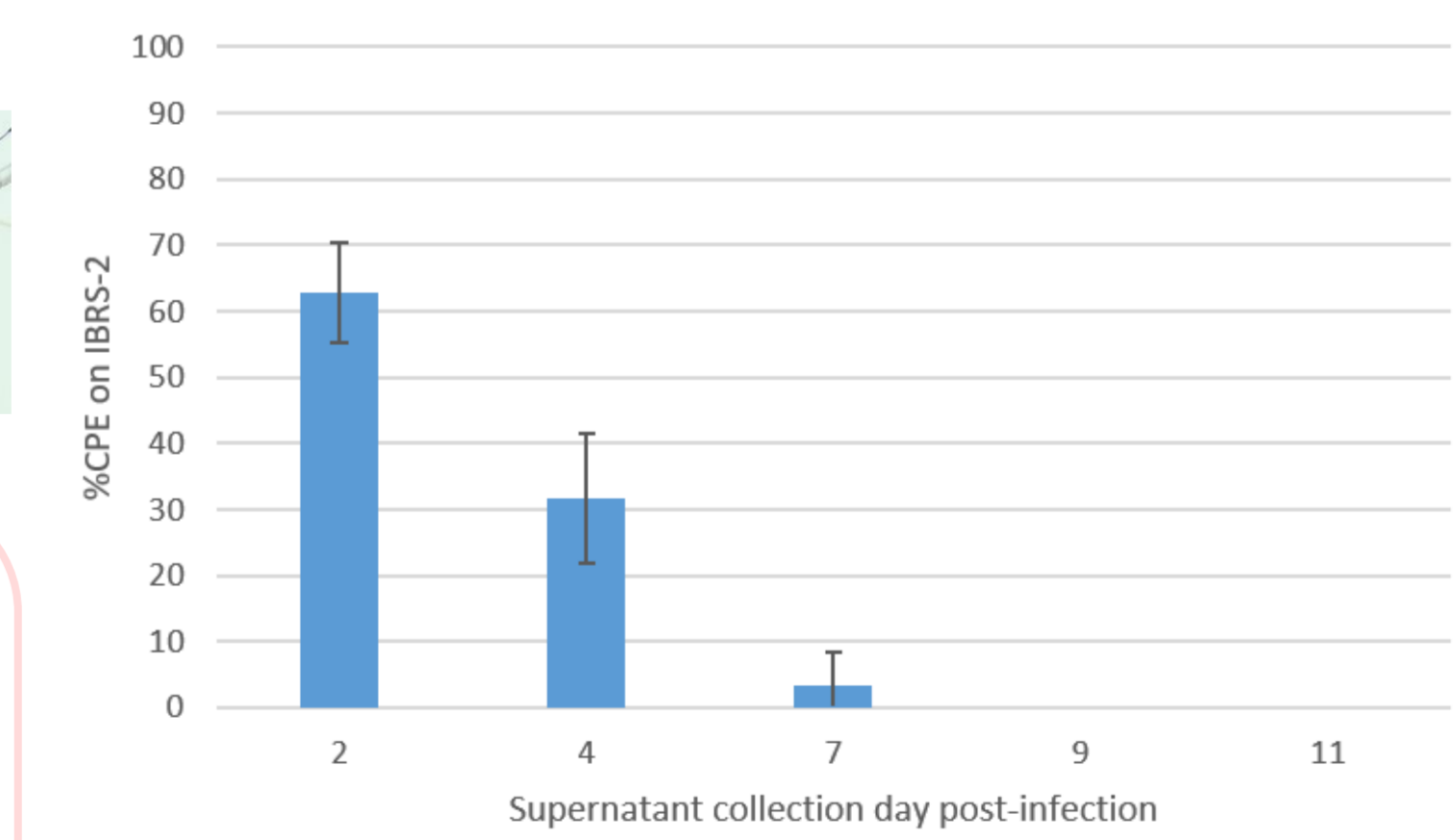
- Porcine DSP were susceptible to the eight strains tested.
- Cytopathic effect on porcine DSP were easy to observe for all the strains.
- Porcine DSP susceptibility to certain strains, in particular from SAT1 and SAT3 serotypes, was lower than that of the cells currently used.

④ Infection of DSP multilayers

- DSP cells were grown in multilayers, on collagen membrane, at the air-liquid interface and then infected with a type-O FMDV at MOI 1.
- Culture supernatants were collected every two or three days for 39 dpi and analyzed for infectious virus, antigen and viral RNA.



- Multilayer depth was determined using HES staining.
- Infectious virus were detected by virus isolation up to 7 dpi in culture supernatants.
- Viral antigen were detected up to 9 dpi by immunofluorescence.
- Viral RNA were detected up to 23 dpi by RTqPCR.



Conclusion & perspectives

- We have succeeded in developing a primary porcine dorsal soft palate cell model susceptible to FMDV and other vesicular disease viruses infection.
- A similar model based on oropharyngeal tonsil cells, could not be developed since these cells did not maintain in culture.
- Consistent with what has been observed in vivo in pigs, no evidence of FMDV persistence could be detected in our cell model (no infectious virus detected after 28 dpi).

- Porcine DSP cells could be immortalized to obtain a cell line suitable for FMDV research and diagnosis.
- These cells could constitute a privileged tool in the differential diagnosis of FMDV.

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