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Proficiency study for the determination of nitrofuran metabolites in shrimps.

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Proficiency study for the determination of nitrofuran metabolites in shrimp

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Methods/Techniques:	Proficiency testing, LC/MS
Additives/Contaminants:	Veterinary drug residues
Food Types:	Fish and fish products

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Table 1 : Results of the analysis of variance for the homogeneity study.

	Source of variation	Sum of square	df	Mean-square	F-ratio
Mat 4 - AOZ	Between containers	0.2794	9	0.031	1.03
	Analytical	0.3	10	0.030	
Mat 5 - AOZ	Between containers	299.65	9	33.294	1.13
	Analytical	294.4	10	29.437	
Mat 5 -SEM	Between containers	0.55	9	0.061	4.21
	Analytical	0.1	10	0.014	
Mat 6 - AOZ	Between containers	5.27	9	0.585	0.71
	Analytical	8.2	10	0.822	
Mat 6 -SEM	Between containers	2.86	9	0.317	0.91
	Analytical	3.5	10	0.350	
Mat 7 - AOZ	Between containers	0.091	9	0.010	0.18
	Analytical	0.6	10	0.0570	

Critical value of $F(p=0.05, v_1 = 9, v_2 = 10)$ is 3.02.

Table 2 : characteristics of the methods used.

Lab. code	Technique	Calibration range $\mu\text{g}/\text{kg}$ (Number of levels)	Internal standard	CC α ($\mu\text{g}/\text{kg}$)				CC β ($\mu\text{g}/\text{kg}$)			
				AOZ	AMOZ	SEM	AHD	AOZ	AMOZ	SEM	AHD
A	LC/MS-MS	0.1 – 60 (8)	D4-AOZ D5-AMOZ	0.7	0.7	0.7	0.7	1	1	1	1
B	LC/MS-MS	0.25 – 5 (6)	D4-AOZ D5-AMOZ	0.3 [#]	0.3 [#]	1 [#]	1 [#]				
H	LC/MS-MS	0 – 5 (7)	D4-AOZ D5-AMOZ	0.5 [°]	0.5 [°]	0.5 [°]	0.5 [°]				
L	LC/MS-MS	0.5 – 2 (4)	D4-AOZ D5-AMOZ	0.10	0.17	0.22	0.18	0.26	0.30	0.39	0.33
M	LC/MS-MS	0 – 10 (5)	D4-AOZ D5-AMOZ	0.5	0.5	0.5	0.5	0.5	0.5	1.2	0.5
N	LC/MS-MS	0 – 5 (5)	D4-AOZ D5-AMOZ	0.23	0.77	0.17	0.77	0.6	0.83	1.06	1.07
P	LC/(MS) ⁿ	0.5 – 5 (4)	D4-AOZ D5-AMOZ								
S	LC/MS-MS	0 – 7 AOZ (5) 0 – 1.5 SEM (5)	D4-AOZ D5-AMOZ C ₁₃ N ₁₅ - N ₁₅ SEM								
T	LC/(MS) ⁿ	1-25 (5)	D5-AMOZ		0.5				0.7		
U	LC/MS-MS	0.1-4.0 (6) AOZ, AMOZ 0.33-13.36 (6) SEM, AHD	D5-AMOZ	0.3 ⁺	0.3 ⁺	1 ⁺	1 ⁺				
V	LC/MS-MS	0 - 5 (5)	D4-AOZ D5-AMOZ	0.3	0.3	0.5	0.5	0.4	0.4	0.8	0.8
W	LC/MS-MS	0 - 10 or 0 - 50 (6 or 7)	D4-AOZ	0.18	0.07	0.37	0.47	0.51	0.31	0.71	1.91
X	LC/(MS) ⁿ	0.25-10 (7)	D4-AOZ D5-AMOZ	0.1	0.1	0.1	0.1				
Z	LC/MS-MS	0.125 – 2.0 (6)	D4-AOZ D5-AMOZ	0.12	0.13	0.25	0.30	0.17	0.25	0.37	0.41
E [□]	LC/MS	5 – 50 (6)	D4-AOZ	3.8				8			
F	LC/MS-MS	0.25 – 5 (5)	D4-AOZ D5-AMOZ	0.2 ⁺	0.2 ⁺	0.5 ⁺	0.5 ⁺				
C	LC/MS-MS	1 – 10 (4)	No	0.2	0.2	0.2	0.2	0.5	0.5	0.5	0.5
K	LC/MS	2 – 50 (4)	No	0.3 [°]	0.3 [°]	0.3 [°]	0.3 [°]	0.7 [*] 1 ⁺	0.7 [*] 1 ⁺	1 [*] 2 ⁺	0.7 [*] 1 ⁺
Q	LC/(MS) ⁿ	0.5 – 5 (4)	D4-AOZ D5-AMOZ	1 [°]	1 [°]	1 [°]					
R	LC/(MS) ⁿ	0.5 – 5 (4)	D4-AOZ D5-AMOZ								

[□] : determination of AOZ only.

[#] : decision limit based on lowest level of validation

⁺ : limits of reporting

[°] : LOD

^{*} : LOQ

empty field : no data available

Table 3 : assigned values ($\mu\text{g}/\text{kg}$) and their standard deviation in test materials.

	AOZ	AMAZ	SEM	AHD
Material 1	-	-	-	-
Material 2	-	-	-	-
Material 3	-	-	-	-
Material 4				
Assigned value ($\mu\text{g}/\text{kg}$)	1.2	-	-	-
Standard deviation ($\mu\text{g}/\text{kg}$)	0.5			
Relative standard deviation (%)	42			
Number of data	16			
Material 5				
Assigned value ($\mu\text{g}/\text{kg}$)	32.1	-	1.3	-
Standard deviation ($\mu\text{g}/\text{kg}$)	16.1		0.6	
Relative standard deviation (%)	50.1		41.7	
Number of data	18		14	
Material 6				
Assigned value ($\mu\text{g}/\text{kg}$)	7.9	-	1.4	-
Standard deviation ($\mu\text{g}/\text{kg}$)	4.2		0.6	
Relative standard deviation (%)	52.9		47.1	
Number of data	18		13	
Material 7				
Assigned value ($\mu\text{g}/\text{kg}$)	0.9	-	-	-
Standard deviation ($\mu\text{g}/\text{kg}$)	0.3			
Relative standard deviation (%)	36.3			
Number of data	15			

Table 4 :

Expected negative result : -

Expected positive result : +

Lab. code	Technique	Mat 1	Mat 2	Mat 3	Mat 4-A	Mat 4-B	Mat 5	Mat 6	Mat 7
A	LC/MS-MS	-	-	-	+	+	+	+	+
B	LC/MS-MS	-	-	-	+	+	+	+	+
H	LC/MS-MS	-	-	-	+	+	+	+	+
L	LC/MS-MS	-	-	-	+	+	+	+	+
M	LC/MS-MS	-	-	-	+	+	+	+	+
N	LC/MS-MS	-	-	-	+	+	+	+	+
P	LC/(MS) ⁿ	-	-	-	+	+	+	+	+
S	LC/MS-MS	-	-	-	+	+	+	+	+
T	LC/(MS) ⁿ	+ (AMoz)	-	+ (AMoz)	-	-	-	+ (AMoz)	-
U	LC/MS-MS	-	-	-	+	+	+	+	+
V	LC/MS-MS	-	-	-	+	+	+	+	+
W	LC/MS-MS	-	-	-	+	+	+	+	+
X	LC/(MS) ⁿ	-	-	-	+	+	+	+	-
Z	LC/MS-MS	-	-	-	+	+	+	+	+
E	LC/MS *	-	-	-	-	-	+	+	-
R	LC/(MS) ⁿ	-	-	-	+	+	+	+	+
K	LC/MS	-	-	-	+	+	+	+	+
C	LC/MS-MS	-	-	-	+	+	+	+	+
Q	LC/(MS) ⁿ	+ (AOZ)	+ (AOZ)	-	+ ^{**} (AMoz + AOZ)	-	+	+	**
F	LC/MS-MS	-	-	-	+	+	+	+	+
False positive		2	1	1					4/60 : 6.7 %
					1			1	6/160 : 3.7 %
False negative					2	3	1		3 9/98 : 9.2 %

- : declared compliant

+ : declared non-compliant

in brackets : false identification

* : determination of AOZ only.

** : possible confusion with material 4-A and 7 for laboratory Q.

Table 5 :Z-scores for contaminated samples

LAB	AOZ 1.2 µg/kg				AOZ 0.9 µg/kg		SEM 1.3 µg/kg		SEM 1.4 µg/kg	
	Mat 4-1		Mat 4-2		Mat 7		Mat 5		Mat 6	
	Za ($\sigma = 0.25$)	Zr ($\sigma_r = 0.17$)	Za ($\sigma = 0.25$)	Zr ($\sigma_r = 0.17$)	Za ($\sigma_r = 0.19$)	Zr ($\sigma_r = 0.13$)	Za ($\sigma_r = 0.29$)	Zr ($\sigma_r = 0.19$)	Za ($\sigma_r = 0.29$)	Zr ($\sigma_r = 0.19$)
X	-3.2	1.2	-3.4	1.7						
U	-3.0	0.8	-2.8	0.4	-2.7	0.5	-2.4	0.7	-2.5	0.0
L	-1.6	0.5	-1.7	0.5	-1.5	0.2	7.9	18.3	-1.7	0.4
M	-1.6	0.6	-0.8	1.2	-1.4	0.0				
F	-1.0	0.8	-1.4	0.0						
K	-0.9	0.5	-1.2	0.1	-1.3	0.4	-2.6	1.1	1.8	7.7
B	-0.8	0.1	-1.2	0.0	-0.8	0.2	1.2	0.9	2.3	0.5
S	-0.2	0.0	-0.2	0.0	-0.8	0.0	-0.2	1.9	-0.4	0.4
N	-0.2	1.4	-0.4	0.8	-0.3	0.8	-1.7	0.5	-2.2	0.5
V	0.4	1.2	0.0	2.1	-0.3	1.1	-0.4	0.0	-0.7	0.4
H	0.8	1.2	1.4	2.5	1.5	0.5	2.4	7.4	2.0	7.4
Z	0.8	0.4	-0.6	0.0	-0.3	0.3	1.0	0.7	0.5	0.0
R	0.9	0.3	2.3	2.6	1.9	0.1				
A	1.2	1.2	3.0	3.3	1.5	1.6	6.5	3.7	3.5	0.0
P	4.6	3.3	1.8	1.7	0.5	1.6	0.0	0.7	-1.2	1.4
W	4.6	0.1	2.0	1.4	1.9	1.5	1.3	0.0	0.8	0.6
C	4.8	0.4	4.8	1.2	2.8	0.0	0.1	2.6	1.0	0.4
Q							-0.8	0.6		
Satisfactory Za-score ($ z \leq 2$): - Number of lab - Rate	12/17 70 %		12/17 70 %		13/15 87 %		9/14 64 %		9/13 69 %	

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8 **Figure 1 : Stability study for AOZ.**

9 **Figure 2 : Stability study for SEM.**

10 **Figure 3 : Accuracy z-scores for material 4. Dotted line corresponds to z-**
11 **score = 2. Solid line corresponds to z-score = 3.**

12 **Figure 4 : Accuracy z-scores for material 5.**

13 **Figure 5 : Accuracy z-scores for material 6.**

14 **Figure 6 : Accuracy z-scores for material 7.**

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Fig 1

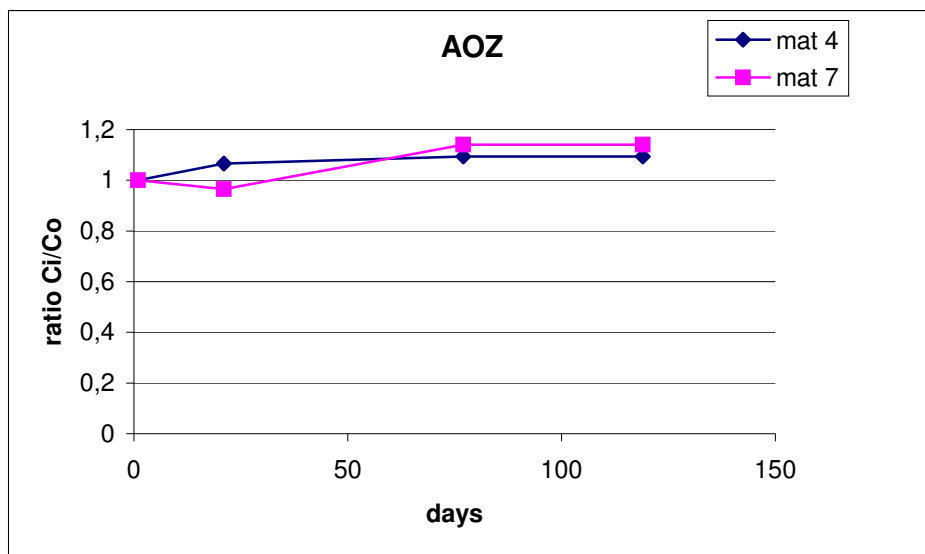


Fig 2

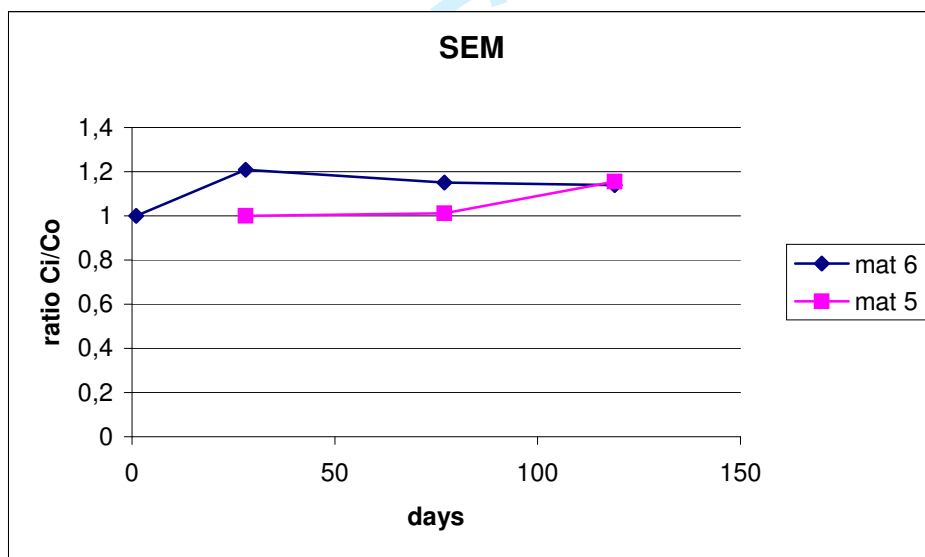


Fig 3
Material 4

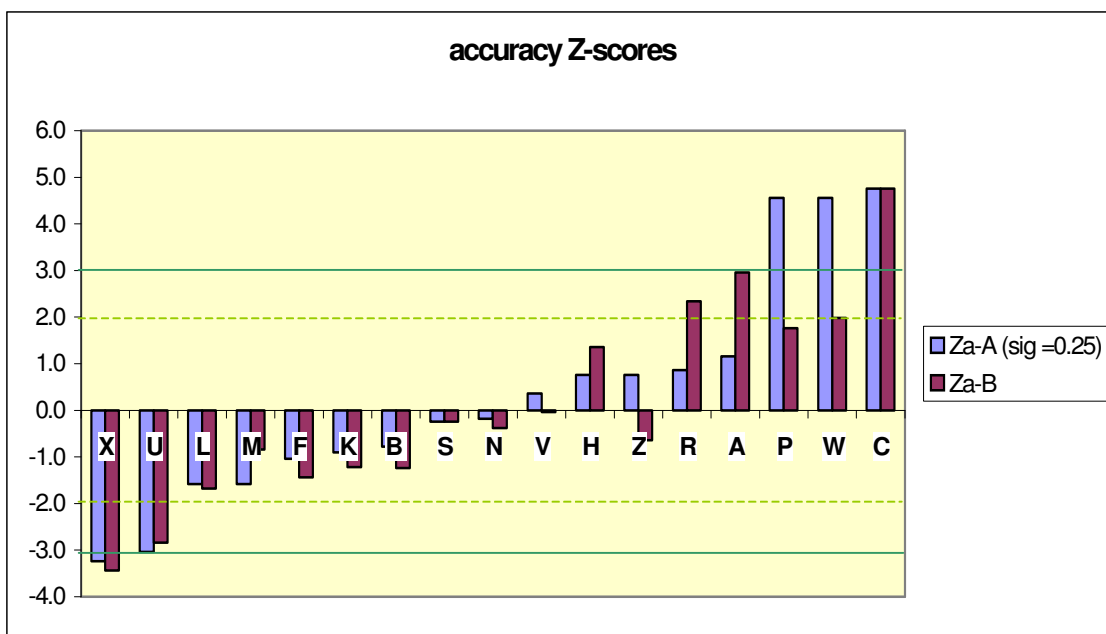


Fig 4
Material 5

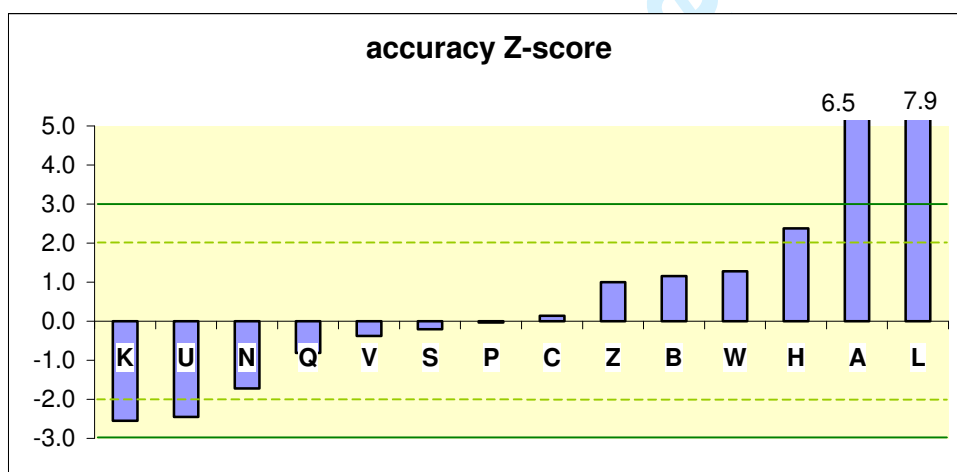


Fig 5
Material 6

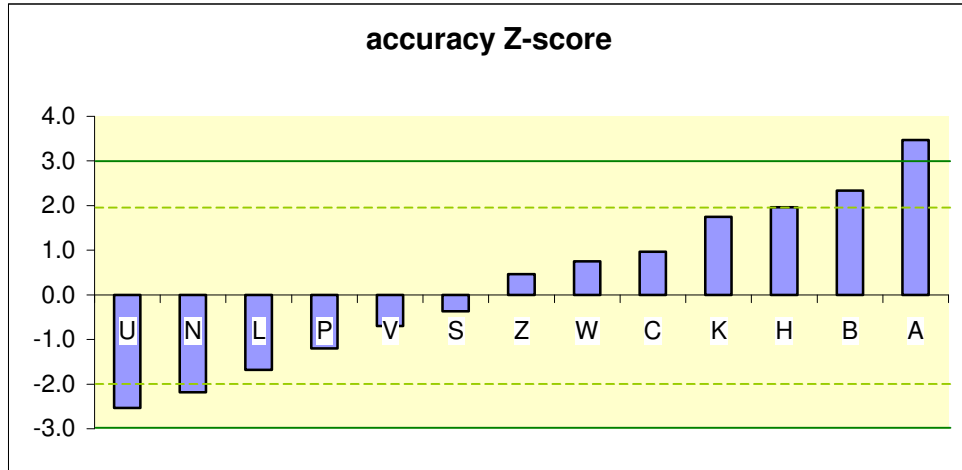
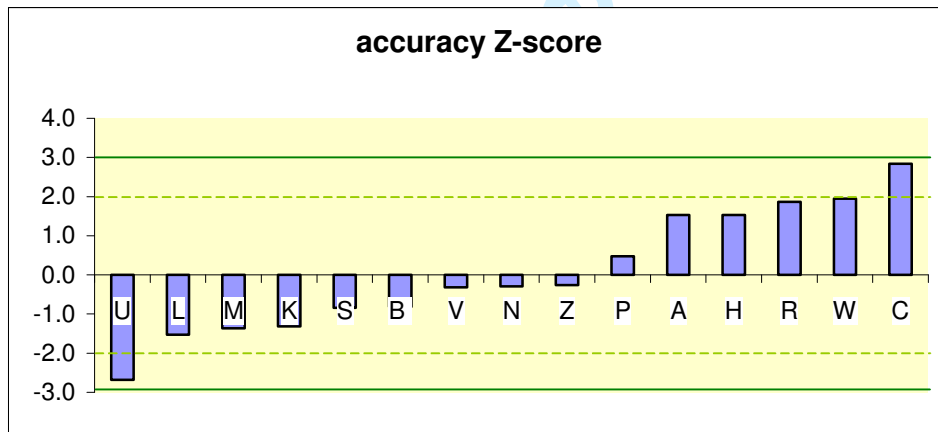


Fig 6
Material 7



Proficiency study for the determination of nitrofuran metabolites in shrimps

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A proficiency test for the determination of nitrofuran metabolites in shrimp tissue was organised in the first half of 2003. This test was intended to allow the participants to use their routine method and to assess their competence on this specific analysis. The participation in this proficiency test was offered to all the National Reference Laboratories (NRLs) of the European Union (EU) in charge of the analysis of nitrofurans, to Official Laboratories of the then 10 Candidate Countries for entry in EU and to some countries exporting food to the EU. The participants (20) analysed nitrofuran metabolites in 8 randomly coded frozen samples including 3 blank samples. All participants performed a confirmatory method using liquid chromatography/mass spectrometry to detect total nitrofuran metabolite residues. Both qualitative and quantitative analysis of the results were investigated. Qualitatively, 16 laboratories out of 20 gave the correct interpretation of the results in term of compliant / non-compliant sample. Quantitatively, laboratory performance was evaluated by calculating the z-scores.

Keywords : nitrofurans, residues, LC-MS/MS analysis, proficiency, interlaboratory study, metabolites.

Introduction

Nitrofurans are antibacterial agents that have been used as feed additives for cattle, pig and poultry. Since 1993 and 1995 (for furazolidone), the use of nitrofurans (furazolidone, furaltadone, nitrofurazone, nitrofurantoin) was banned in EU for food animal production (Council regulation (EEC) No 2901/93, 1993, Commission Regulation (EC) No 1442/95, 1995). However, these substances are still in use in some other countries worldwide and residues of nitrofurans have been found in a wide range of food-animal products (poultry, aquaculture products, eggs). In March 2003, a definitive MRPL (Minimum Required Performance Limit) was set at 1 µg/kg in EU for these drugs in poultry and aquaculture products (Commission decision 2003/181/EC, 2003). So, all laboratories involved in the monitoring of nitrofurans residues have to reach at least this limit. Within the EU, the control of nitrofurans in food from animal products started ten years ago. The testing for residues of the parent drugs was ineffective because nitrofurans compounds are rapidly metabolized. *In vivo*, they formed stable and persistent tissue-bound residues. A few years ago, the ability of EU NRLs to test for tissue-bound nitrofurans residues was poor. This is why a liquid chromatography tandem mass spectrometry (LC-MS/MS) method developed through the FoodBrand EU project (QLK1-CT1999-00142, available from www.afsni.ac.uk/foodbrand) was transferred and most of the laboratories involved in the veterinary drug residue control in EU implemented this LC-MS/MS method for screening and confirmation of nitrofurans residues in tissue. This method focused on AOZ (3-amino-2-oxazolidinone), AMOZ (3-amino-5-morpholinomethyl-2-oxazolidinone), AHD (1-aminohydantoin) and SEM (semicarbazide) the marker residues of the nitrofurans banned parent drugs furazolidone, furaltadone, nitrofurantoin and nitrofurazone respectively. It is based on acid hydrolysis of protein-bound residues followed by the derivatization using nitrobenzaldehyde to lead to the nitrophenyl derivatives. The literature describes few methods using LC-MS/MS for the analysis of nitrofurans metabolites (Leitner *et al.* 2001, Mottier *et al.* 2005, Cooper *et al.*

2005). Therefore, it became important that laboratories had the opportunity to test their ability to control nitrofurans and to prove the reliability of their result. As the organization of proficiency tests for European Union-National Reference Laboratories (EU-NRLs) is one of the tasks of the Community Reference Laboratory (CRL), a proficiency test for the analysis of nitrofurans was organized during the period February-July 2003. The participation in this test was offered to all EU-NRLs, but also to Official Laboratories of Candidate Countries for 2004 entry in EU and to some countries exporting food into EU. The study was conducted according to the recommendations of ISO guide 43-1 (1997). This paper reports the results of the first proficiency testing study for the determination of nitrofurans residues in shrimps.

Experimental

Sample material preparation.

Materials for the test were shrimps obtained from the French Border Inspection Services. 7 different batches of frozen raw shrimps were chosen as follows : 3 batches of shrimps where nitrofurans residues were found to be absent (i.e. below the LOD of 0.1 to 0.2 µg/kg depending on the nitrofurans) were selected as blank matrix and 4 other batches of shrimps where nitrofurans residues were found to be present were selected as naturally contaminated matrices. Shrimps from a same batch were thawed, deshelled and only the tails of the shrimps were minced. Each material was homogenized in a mixing bowl, sampled into containers (10 g) and stored at -20°C until dispatching. Each container was randomly coded. One material (material 4) was sent blindly in duplicate samples. A total of eight frozen samples was shipped to each participant.

Homogeneity and stability studies.

Before sending the samples, it was necessary to test the materials for sufficient homogeneity. The sample homogeneity was determined for each contaminated material by analysing 10 randomly selected containers in duplicate and comparing the sampling variance to the analytical variance, according to the 'International Harmonised Protocol for Proficiency Testing of (Chemical)

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Analytical Laboratories' (Thompson *et al.* 1993). For each material, the procedure was carried out as follows : 10 containers were randomly selected among the containers stored at -20°C and thawed at room temperature. 2 test portions were taken from each container and the analysis of the 2 x 10 portions were carried out under the same day. The samples were analysed by the CRL-AFSSA Fougères using the LC/MS-MS method. Homogeneity was assessed by means of a one-way analysis of variance (ANOVA) of the concentration values. The results are given in table 1.

The analysis of variance proved that the materials were found to be homogeneous. The statistical F-test showed that the sampling variance doesn't differ from the analytical variance with 5% risk assessment (F-ratio < F critical) except for SEM in material 5. For this material, although the ratio of the sampling variance $S_s / \sigma = 0.4$ (where S_s is the sampling variance and σ is the target value for reference standard deviation derived from Horwitz equation) is higher than the recommended value 0.3, it was decided to use it for proficiency study, as the material was homogeneous for AOZ.

[insert table 1 about here]

Stability studies were performed during the proficiency test to prove that the analyte content did not significantly change with regard to analytical variations. The materials were analysed at different time over a period of 3 months being stored at -20°C . The ratio C_i/C_o was determined for each period : C_i was the concentration at the time T_i and C_o was the concentration at initial time T_o . The time T_o was the day of the first analysis carried out at CRL-AFSSA Fougères during the period of the interlaboratory study. No loss of AOZ or SEM content could be observed during this period under storage at -20°C (see figures 1-2).

[insert figures 1 and 2 about here]

Results and discussion

The participants were asked to analyse the 8 samples for their possible content of nitrofurans metabolites using the method of their choice. Each sample had to be analysed in duplicate. Detected nitrofurans had to be quantified and confirmed according to Commission Decision 2002/657/EC (2002). Moreover,

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5 for each sample, participants had to give an interpretation of the result as
6 compliant or non-compliant in line with Decision 2002/657/EC. 20 laboratories
7 participated to the study : 14 were EU-NRLs and 6 were outside EU laboratories
8 (in 2003).
9

10 11 12 13 *Methods and analytical techniques.*

14 All participants used LC/MS to analyse the samples but with different mass
15 instruments : LC/MS single quadrupole (10 %), LC/(MS)ⁿ ion trap (25 %) and
16 LC/MS-MS triple quadrupole (65%).
17

18 All methods used to detect total nitrofurans metabolite residues were based on
19 the following principle : hydrolysis and derivatization with nitro-benzaldehyde
20 during one night at 37 °C, extraction of the nitrophenyl derivatives, evaporation
21 and reconstitution of the sample before injection. Most of the participants took
22 advantage of deuterated internal standards. The analytical characteristics of the
23 methods are displayed in table 2. Not all the participants gave the analytical
24 limits of their method. One can observe that, except for laboratory E, the limits
25 of decision CC_α comply with the MRPL of 1 µg/kg. However, the participants
26 did not give informations on the way they calculated the limits. Depending on
27 the participant, the given limit is either the limit of reporting, either the lowest
28 level of validation or either the limit of detection.
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30 31 32 33 *Assigned values.*

34 The assigned values of the materials were determined as a consensus value of
35 the results of all participants (except one laboratory for its extra-deadline
36 sending of results). They were calculated as the robust average of the results,
37 as it is described in the Annex C of ISO/DIS 13528 (2000). The robust
38 estimates of x^* and s^* are derived by an iterative calculation, updating the
39 values of x^* and s^* several times, until the process converges. This calculation
40 provides robust values of the average and standard deviation of the data to
41 which it is applied with no need to discard any of the data. The results are given
42 in Table 3.
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60 [insert table 3 about here]

Qualitative analysis

Participants analysed the samples for the presence of the 4 metabolites : AHD, AOZ, AMOZ and SEM. One participant analysed only for AOZ. Table 5 shows laboratory qualitative results. For a blank sample, the expected result is – (compliant) and for a contaminated sample, the expected result is + (non compliant).

Over the 20 participants, only two laboratories (T and Q) found false positive results. They confirmed the presence of AMOZ and AOZ in materials 1, 2 or 3 with LOD of 0.5 µg/kg for laboratory T and 1 µg/kg for laboratory Q. The LOD of the others participants are nearly all inferior. For the blank samples, the rate of false positive is 6.7 %. For the contaminated sample, the determination of an analyte which is actually not present counted also as a false positive result. It is the case both for material 6 and material 4 for which laboratory T and laboratory Q respectively, confirmed AMOZ instead of AOZ. Considering these samples, the rate of false positive is 3.7 %. Usually, the false positive result can often be explained by cross-contamination between samples. Among the 8 samples sended, one sample (material 5) was highly contaminated with AOZ (~ 30 µg/kg) and might lead to cross-contamination. For laboratory T finding three AMOZ contaminated samples and for laboratory Q finding one AMOZ contaminated sample, no explanation is proposed.

Taking into account that, when SEM and AOZ were both present in a material, only one of them had to be detected to declare the sample as no-compliant, it is assumed that the rate of false negative is approximately 9 %. Laboratory E found 3 false negative results. Obviously the 3 samples with contaminated level around 1 µg/kg were not confirmed because of the inadequate limit of its method ($CC\alpha = 3.8 \mu\text{g/kg}$). Laboratories T, X and Q gave also false negative results.

[insert table 4 about here]

Quantitative analysis

The statistical approach used for the analysis of the results was based on the z-score [ISO, 1997].

The accuracy z-score was given by : $Z_a = (x - \hat{X}) / \sigma$ where Z_a is the "accuracy z-score", x is the laboratory result (mean of the duplicate analyses), \hat{X} is the assigned value and σ is the target value for standard deviation. The target value for standard deviation σ was determined according to the equation proposed by M. Thompson (2000):

$$\sigma = 0.22 c \quad \text{if } c < 1.2 \times 10^{-7}. \quad (\equiv 120 \text{ ppb}) \text{ where } c \text{ is the concentration..}$$

As each laboratory analysed every material in duplicate, repeatability z-score was also calculated as : $Z_r = SD / \sigma_r$ where Z_r is the "repeatability z-score", SD is the standard deviation of the 2 results and σ_r is the target value for standard deviation determined as follows : $\sigma_r = 2/3 \sigma$ **

** When the analyses are carried out under repeatability conditions, the Commission decision 2002/657/CE (2002) specifies that the intralaboratory RSD to be between one-half and two third of the reproducibility RSD.

The z-scores give a numerical approach of the performance of a laboratory in a proficiency test. It allows a classification as follows :

$$\begin{aligned} |Z| \leq 2 &\Rightarrow \text{Satisfactory} \\ 2 < |Z| < 3 &\text{Questionable} \\ |Z| \geq 3 &\Rightarrow \text{Unsatisfactory} \end{aligned}$$

The z-score values have not been calculated for AOZ content in material 5 and 6 because the high values of AOZ (32.1 and 7.9 $\mu\text{g}/\text{kg}$ respectively) reported by the participants came for most of them from extrapolation of a too short calibration range, usually centred around the MRPL level concentration of 1 $\mu\text{g}/\text{kg}$. Even if it would be expected that a calibration curve should be made in a range which covers all the samples analysed, many participants focused on the accuracy at the MRPL level. Extra high concentration has not practical

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importance from the point of the possible legal action. The accuracy of the quantification beyond 2 MRPL is of lesser importance in the case of banned substances. For that, the z-scores were only calculated for SEM content of these materials 5 and 6 and for AOZ content of material 4 and 7. Table 5 displays an overview of the z-scores. Histograms presenting the data for accuracy z-scores are given in figures 3-6.

[insert table 5 about here]

[insert figures 3-6 about here]

Material 4 (1.2 µg/kg AOZ) was sent in duplicate blindly. From figure 3, for this material, it is noteworthy that 4 laboratories (R, A, P and W) have obtained very different Za-scores for the two identical samples. For material 7 (0.9 µg/kg AOZ), none Za-scores are unsatisfactory and 13 over 15 are satisfactory. For materials containing SEM (5 and 6), 2 laboratories (A and L) are involved in high Za-score values. Laboratory A obtained unsatisfactory results both for material 5 and material 6. For laboratory L which obtained a high Za-score for material 5, this value could be due to a bad repeatability in the analysis done in duplicate, shown by a high Zr-score.

Considering only the participants who gave quantitative results (excluding the false positive and the false negative responses), the global z-score evaluation showed that :

- the rate of laboratories producing a satisfactory accuracy z-score for total AOZ content around the MRPL (between 0.9 and 1.2 µg/kg) is around 70 to 87 %.
- the rate of laboratories producing a satisfactory accuracy z-score for total SEM content around the MRPL (between 1.3 and 1.4 µg/kg) is around 64 to 69 %.

Even if all false positive or false negative results have come from laboratories (T, Q and X) using ion trap MS instrumentation, it is not possible to see a relationship between the type of MS used and the quality of the results. The number of participants using ion trap MS is too small to allow to give a

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significant conclusion from this study. The evidence is given by 2 others laboratories (P and R) which used ion trap MS : Laboratory P showed satisfactory Za-scores for materials 4-2, 5, 6 and 7 and laboratory R obtained 2 satisfactory Za-scores for material 4-1 and 7 and 1 questionable for AOZ. The bad results are rather due to the operators only. Nevertheless, with our experiment in the field of residues analysis, it is clear that triple quadrupole analyser is the type of instrument the best suitable to obtain reliable quantitative results.

Conclusions

For the control of banned substances, the rate of false positive / false negative results is of primary importance for official residue control. False negative results have implications for health protection. False positive result leads to direct trouble for the producer. The rate of false positive / false negative results cannot be evaluated without binding it to the analytical limit of the methods and to the concentration levels of the incurred samples. From this proficiency study, 16 laboratories out of 20 found the correct compliant and non-compliant samples according to the Decision 2002/657/EC.

The quantitative analysis of this study is important too because it shows that all z-scores were not satisfactory for samples containing SEM or AOZ around the MRPL level. The participants who obtained unsatisfactory results have to find the causes of that fact. In order to harmonize the control of nitrofurans metabolites from a point of view of legal action, it is necessary that quantitative results are homogeneous around the MRPL level.

However, it should be stressed that decision limits CC_{α} reported by the participants varied from 0.1 to 1 $\mu\text{g}/\text{kg}$ except for one participant (for which CC_{α} is 3.8 $\mu\text{g}/\text{kg}$ for AOZ using LC/MS single quadrupole). They are in accordance with the MRPL.

This proficiency test was very useful providing for the first time an overview of the control of nitrofurans metabolites inside and outside EU. It allowed participants to check their ability to confirm analytes with their own routine method. From that point, participants can either prove the reliability of their

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results or either take measures to improve their method. To keep on with efforts on harmonization of control of nitrofurans metabolites residues in EU, another proficiency test for nitrofurans metabolites in chicken is in progress.

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